

1 **Performance of titanium salts compared to conventional FeCl₃ for the**
2 **removal of algal organic matter (AOM) in synthetic seawater: Coagulation**
3 **performance, organic fraction removal and floc characteristics.**

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15 **Abstract:**

16 During algal bloom periods, operation of seawater reverse osmosis (SWRO) pretreatment processes
17 (e.g. ultrafiltration (UF)) has been hindered due to the high concentration of algal cells and algal
18 organic matter (AOM). The present study evaluated for the first time the performance of titanium salts
19 (i.e. titanium tetrachloride (TiCl₄) and polytitanium tetrachloride (PTC)) for the removal of AOM in
20 seawater and results were compared with the conventional FeCl₃ coagulant. Previous studies already
21 demonstrated that titanium salts not only provide a cost-effective alternative to conventional
22 coagulants by producing a valuable by-product but also minimise the environmental impact of sludge
23 production. Results from this study showed that both TiCl₄ and PTC achieved better performance than
24 FeCl₃ in terms of turbidity, UV₂₅₄ and dissolved organic carbon (DOC) removal at similar coagulant
25 dose. Liquid chromatography – organic carbon detection (LC-OCD) was used to determine the
26 removal of AOM compounds based on their molecular weight (MW). This investigation revealed that
27 both humic substances and low MW organics were preferentially removed (i.e. up to 93% removal)
28 while all three coagulants showed poorer performance for the removal of high MW biopolymers (i.e.
29 less than 50% removal). The detailed characterisation of flocs indicated that both titanium coagulants
30 can grow faster, reach larger size and present a more compact structure, which is highly advantageous
31 for the design of smaller and more compact mixing and sedimentation tanks. Both titanium coagulants
32 also presented a higher ability to withstand shear force, which was related to the higher amount of
33 DOC adsorbed with the aggregated flocs. Finally, TiCl₄ had a better recovery after breakage
34 suggesting that charge neutralization may be the dominant mechanism for this coagulant, while the
35 lower recovery of both PTC and FeCl₃ indicated that sweep flocculation is also a contributing
36 mechanism for the coagulation of AOM.

37

38 **Keywords:** Coagulation, Algal organic matter (AOM), Polytitanium tetrachloride, Titanium
39 tetrachloride, Floc characteristics

40 **1 Introduction**

41 The sustainability of water and energy resources is being threatened due to rapid population and
42 industry growth and therefore, developing low-energy separation technologies is crucial to meet the
43 increasing water demand through unconventional sources (Chung et al., 2012; Elimelech and Phillip,
44 2011; Rasul and Sharma, 2016; Zhao et al., 2012). Nowadays, membrane processes are the most
45 widely used technology to produce clean water and, among them, reverse osmosis (RO) is currently
46 the most employed separation technology for desalination (up to 70% of installed desalination plants)
47 (Lee et al., 2011). One of the major obstacles for cost-effective operation of seawater RO (SWRO) is
48 membrane fouling, which significantly reduces the process efficiency in terms of quality and quantity
49 of produced water (Matin et al., 2011). Therefore, pretreatment processes such as micro- or ultra-
50 filtration (MF/UF) are generally required to mitigate this issue. UF has gained increased popularity as
51 an alternative pretreatment process to SWRO (Basha et al., 2013; Brehant et al., 2002). In fact, UF
52 systems have lower footprint, better permeate quality and lower overall chemical consumption
53 compared with conventional pretreatment process (e.g. dual media filtration) (Wilf and Schierach,
54 2001). However, similar to RO, membrane fouling remains one of the major challenges impeding the
55 high production capacity of this membrane system (Pearce, 2009). During seawater desalination,
56 membrane fouling in UF pretreatment systems is largely associated with algal bloom events and it has
57 been recently shown that algal cells and more specifically algal organic matter (AOM) were
58 responsible for the severe permeability decline (Li et al., 2016; Merle et al., 2016; Pivokonsky et al.,
59 2016; Villacorte, 2014).

60 AOM released by bloom-forming algal species mainly comprises extracellular biopolymers (i.e. high
61 molecular weight, hydrophilic, anionic compounds such as polysaccharides and glycoproteins) but the
62 composition of AOM varies from one species to another, and changes over the course of an algal
63 bloom (Fogg, 1983). These high molecular weight (MW) biopolymers are also known as transparent
64 exopolymer particles (TEP) and have been found to play a key role in the aggregation of algae during
65 bloom events (Myklestad, 1995). Previous studies have demonstrated that AOM can deposit on the
66 membrane surface and/or within the pores, which increases the resistance to permeate flow but also
67 reduces the effectiveness of hydraulic cleaning (i.e. backwashing) (Qu et al., 2012; Schurer et al.,
68 2013; Villacorte et al., 2010). Besides, if a fraction of the AOM passes through the UF membranes, it
69 affects the downstream RO membranes by increasing the fouling and biofouling potential of the feed
70 water.

71 Coagulation using conventional metal salts (i.e. aluminium and iron) has been effective in controlling
72 and reducing membrane fouling in UF systems (Busch et al., 2010; Choi and Dempsey, 2004),
73 including in the context of AOM removal during algal bloom events (e.g. (Schurer et al., 2012;
74 Schurer et al., 2013; Tabatabai et al., 2014)). Coagulation can reduce UF membrane fouling by AOM

75 via several mechanisms (which depends on the coagulant dose) including partial complexation of
76 algal biopolymers and formation of biopolymer-colloidal metal complexes (e.g., for low coagulant
77 dose < 1 mgFe/L) and adsorption and enmeshment of biopolymers in metal hydroxide precipitates at
78 higher coagulant dose (Tabatabai et al., 2014; Yan et al., 2017). These mechanisms have been found
79 to be quite different from the coagulation of natural organic matter and are still relatively poorly
80 understood (Tabatabai et al., 2014). To date, ferric salts, and more specifically ferric chloride (FeCl₃),
81 are the most commonly used coagulants in seawater desalination mainly because of their lower
82 solubility compared to aluminium salts, resulting in lower dissolved metal concentration and thus
83 lower scaling potential in the treated water (Edzwald and Haarhoff, 2011). However, the
84 coagulation/flocculation process performed by ferric salts generates a large volume of chemical
85 sludge which has to be disposed, creating an environmentally-challenging burden.

86 To overcome the issue of sludge disposal, titanium salts can be used as alternative coagulants since
87 the sludge generated after titanium salt flocculation can be recycled to produce an economically
88 valuable by-product via calcination: titanium dioxide (TiO₂) (Lee et al., 2009; Okour et al., 2010;
89 Shon et al., 2009; Zhao et al., 2011a). Therefore, the use of titanium-based coagulants offers a more
90 attractive and cost-effective solution to the sludge disposal approach. In fact, an early study on TiCl₄
91 (Shon et al., 2007) showed that a medium size wastewater treatment plant (i.e. treatment capacity of
92 25,000 m³/day) could produce an average of 446.5 kg of TiO₂ photocatalyst per day, which is quite
93 significant considering the increasing market demand for TiO₂. Besides, the performance of recycled
94 TiO₂ (i.e. photocatalytic activity and surface area) was found to be higher than the commercially
95 available TiO₂ (Shon et al., 2007). Finally, another study (Okour et al., 2009), focusing on the
96 application of TiCl₄ and Ti(SO₄)₂ as pretreatment for SWRO, found that the residual Ti after
97 flocculation caused significantly less fouling compared to the residual Fe after conventional FeCl₃
98 flocculation.

99 Floc characteristics (i.e. floc size, floc growth rate, strength and recovery factors and floc structure)
100 are crucial factors to assess for the design and the performance evaluation of a coagulation process.
101 For instance, large flocs with high degree of compaction will settle more rapidly than smaller flocs
102 with large open pore structure. This will lead to not only higher removal efficiency, but also faster
103 settling time resulting in reduced sedimentation tank size requirement. The ability of flocs to resist a
104 shear force and to recover after breakage is also important to evaluate since it will have significant
105 impact on the process design and its capital and operational costs (McCurdy et al., 2004).

106 The objective of this study was therefore to investigate, for the first time, the performance of titanium-
107 based coagulants (i.e. TiCl₄ and polytitanium tetrachloride (PTC)) as an alternative to conventional
108 ferric salts for the removal of AOM in seawater. Coagulation performance has been assessed in terms
109 of turbidity, zeta potential, UV₂₅₄ and dissolved organic carbon (DOC) removal and compared with

110 FeCl₃. The coagulation performance of Ti-salts was also investigated for the removal of AOM
111 fractions (i.e. based on MW). Finally, a detailed comparative study was also conducted to understand
112 the growth, breakage and structural parameters of flocs formed by all three coagulants.

113 **2 Materials and methods**

114 **2.1 Coagulants**

115 TiCl₄ stock solution was obtained by adding continuously drop by drop a concentrated TiCl₄ solution
116 (>99% purity, density $\rho = 1.26$ g/mL; Sigma Aldrich, Australia) to frozen cubes of deionized water to
117 obtain a final concentration of 20% w/w. The concentration of the stock solution was checked using a
118 microwave plasma-atomic emission spectrometer (4100 MP-AES, Agilent Technologies, US) and
119 adjusted to the desired concentration as necessary. FeCl₃ stock solution (i.e. 10 g/L) was prepared by
120 dissolving 2 g of FeCl₃ powder in 200 mL of deionized (DI) water. The preparation of PTC coagulant
121 was developed in a previous study (Zhao et al., 2013). In brief, a predetermined volume (i.e. 63.3 mL)
122 of concentrated sodium hydroxide (i.e. 200 g/L) was added to 200 mL of the TiCl₄ stock solution
123 using a slow alkaline titration under intensive agitation. The basicity value (i.e. OH/Ti ratio) was fixed
124 to 1.5 since it was previously found to generate the best coagulation performance (Zhao et al., 2013).
125 Coagulant dose is given in units of g-Ti/L and g-Fe/L for Ti-coagulants and FeCl₃, respectively.

126 **2.2 Algae cultivation and AOM harvesting**

127 To simulate seawater bloom conditions, the marine diatom species *Chaetoceros muelleri* was
128 cultivated for laboratory-scale production of AOM. In fact, the diatoms of the genus *Chaetoceros* are
129 a suitable choice since they bloom regularly in coastal waters (Ignatiades, 1969; Lewin and Mackas,
130 1972; Passow and Alldredge, 1995) and they were found to generate significant volume of
131 extracellular polysaccharides during their growth cycle (Dam and Drapeau, 1995; Mykkestad, 1995;
132 Watt, 1969).

133 An aquaculture strain of the diatom *Chaetoceros muelleri* (Department of Primary Industries, Port
134 Stephens, Australia) was grown in synthetic seawater enhanced with F/2 medium (Cell-hi F2P,
135 Varicon Aqua) and silicate (Guillard and Ryther, 1962). The microalgae were grown in a 10 L
136 transparent carboy bubbled with filtered air, at an incident irradiance of approximately 120
137 $\mu\text{mol}\cdot\text{photon}/\text{m}^2\cdot\text{s}^2$ and a temperature of 25°C.

138 The algae culture was collected approximately 14 days after inoculation (i.e. during the stationary
139 phase of growth) when the production of extracellular polysaccharides has been found to be maximal
140 (Mykkestad, 1974). Samples were collected regularly and the culture was stopped when the

141 biopolymer concentration reached a steady value of 5 mgC/L. The bottle containing the culture was
142 first gently shaken to ensure homogeneity. Then, the sample was filtered through 0.45 µm nylon
143 membrane filters (Whatman) to harvest the AOM and obtain a stock solution which was then used for
144 characterization and preparation of the feed solution. The AOM stock solution therefore includes
145 mainly extracellular organic matter (EOM) but may also contain intracellular organic matter (IOM) if
146 some of the algae cells were broken during the filtration process.

147 **2.3 AOM characterization**

148 The detailed organic fraction of the AOM stock solution (Table 1) and samples before and after
149 flocculation was determined using liquid chromatography with organic carbon detection (LC-OCD)
150 (LC-OCD model 8, DOC Labor, Germany), with organic carbon, organic nitrogen and
151 UV₂₅₄ detectors and two TSK HW-50S (Toso, Japan) chromatographic columns in series (double
152 column setup) (Huber et al., 2011). The longer retention time (240 mins) of the double column setup
153 was used to analyse organic sub-fractions of seawater (Naidu et al., 2015). LC-OCD separates the
154 hydrophilic fraction of the total dissolved organic carbon (DOC) by molecular size per retention time,
155 with an initial peak of high molecular-sized biopolymer followed by humic substance and their related
156 building blocks and finally low MW organics (LMW organics) (Huber et al., 2011; Naidu et al.,
157 2015). The proportion of each organic fraction was calculated by specialised software (ChromCALC
158 DOC-LABOR, Karlsruhe, Germany).

159

160 **Table 1**

161 The AOM harvested from *Chaetoceros muelleri* (Table 1) mostly comprised of biopolymers (i.e.
162 38.5% of the total DOC) and a relatively large amount of building blocks and LMW organics. The
163 composition of AOM, and more specifically the MW distribution of the organic compounds, has been
164 found to have significant impact on the coagulation process (Henderson et al., 2010).

165 **2.4 Jar-tests**

166 For coagulation performance studies, the AOM stock solution was diluted with synthetic seawater
167 (TDS: 31.8 g/L; Conductivity: 49.7 mS/cm; pH: 7.7 ± 0.1) following the recipe of Lyman and
168 Fleming (1940) to obtain a biopolymer concentration of 0.5 mgC/L. This concentration corresponds to
169 AOM concentrations during algal bloom events in the North Sea, which resulted in significant
170 biofouling of UF systems (Schurer et al., 2013).

171 Coagulation experiments were performed using a programmable jar-tester (PB-900TM, Phipps and
172 Bird, USA). The coagulant was added to 500 mL of AOM in synthetic seawater at the beginning of
173 the rapid mixing step. Since metal hydrolysis occurs immediately after coagulant addition, solution

174 pH decreases and, depending on the coagulant dose, needs to be adjusted. Therefore, to maintain pH
175 in the operational range of SWRO plants (i.e. pH 7-8), pH adjustment was necessary, especially for
176 the higher coagulant doses as shown in Table 2.

177 **Table 2**

178 It is worth noting that TiCl_4 required a higher NaOH dose for pH adjustment which is likely related to
179 the release of large amount of hydrogen ions during titanium hydrolysis (Shon et al., 2007; Zhao et
180 al., 2011b). This is one of the main reasons behind the development of polytitanium salts through the
181 prehydrolysis of TiCl_4 to reduce the need for pH adjustment and thus chemical consumption and cost.

182 Experimental conditions are summarized as follow: rapid mixing was first applied for 1.5 min (i.e.
183 200 rpm), followed by slow mixing for 20 min at 40 rpm and finally 20 min of quiescent settling.
184 Water samples were collected from 2 cm below the water surface for subsequent measurements. The
185 water samples were pre-filtered using a 0.45 μm membrane syringe filter before testing UV_{254}
186 (absorbance at 254 nm using a UV-754 UV/VIS spectrophotometer) while residual turbidity and zeta
187 potential were directly measured using a turbidity meter and Zetasizer (Malvern Instruments, UK),
188 respectively. Turbidity, UV_{254} and zeta potential were measured on three independent samples. Data
189 are presented as mean \pm standard error.

190 **2.5 Floc characterization**

191 The variation of floc size was continuously monitored via an on-line monitoring system using a static
192 light scattering instrument (Mastersizer 2000, Malvern, UK) with an array of photosensitive detectors,
193 positioned at different angles between 0.01° and 40.6° . The floc size was represented by the median
194 equivalent diameter d_{50} , although similar trends for d_{10} and d_{90} floc sizes were also observed (data not
195 shown). The schematic diagram of the on-line monitoring system for continuous floc size
196 measurement can be found elsewhere (Chekli et al., 2015).

197 Floc characterization experiments were conducted under the optimum conditions determined during
198 the jar tests (i.e. at optimum coagulant dose and pH 7-8). The floc properties (i.e. growth rate, strength
199 and regrowth capacity) were determined using the following protocol: 15 min at 40 rpm (i.e. initial
200 growth phase followed by steady state phase), 1 min at 200 rpm (high shear force to induce breakage
201 of aggregated flocs) and finally 20 min at 40 rpm (slow mixing to allow floc regrowth). During these
202 different stages, the suspended flocs were sent through the measurement cell and pumped back into
203 the jar using a peristaltic pump. A preliminary study showed that a flow rate of 2.5 L/h was the
204 optimum flow rate for these experiments. In fact, higher flow rates gave rise to floc breakage while
205 lower ones induced flocs settling within the tubing. These experiments were run in duplicate for each
206 tested coagulant and the data presented are the mean \pm standard error.

207 The flocs strength factor (SF) and recovery factor (RF) were calculated using the following equations
208 (Jarvis et al., 2005; Yukselen and Gregory, 2002):

$$209 \quad SF = \frac{d_2}{d_1} \times 100 \quad (1)$$

$$210 \quad RF = \frac{d_3 - d_2}{d_1 - d_2} \times 100 \quad (2)$$

211 where d_1 is the average floc size at steady-state before breakage, d_2 is the average floc size after
212 breakage, and d_3 is the average floc size after regrowth to a new steady-state.

213 The floc growth rate was evaluated by determining the slope during the growth period (assuming
214 linear growth) (Xiao et al., 2010):

$$215 \quad \text{Growth rate} = \frac{\Delta_{\text{size}}}{\Delta_{\text{time}}} \quad (3)$$

216 The highly disordered structure of floc aggregates can be defined by its scaling behaviour which is
217 known as the mass fractal dimension (FD) (Rice and Lin, 1993; Weitz et al., 1985). The FD describes
218 the actual space occupied by the solid and defines the degree of compactness of the floc aggregate
219 structure.

220 Previous studies have used static light scattering instrument to evaluate the FD of formed flocs (Jarvis
221 et al., 2005; Lin et al., 2008; Wei et al., 2009). The total scattered light intensity I , the scattering
222 vector Q and the FD followed a power law as per the following equation (Rieker et al., 2000):

$$223 \quad I \propto Q^{-\text{FD}} \quad (4)$$

224 Besides, the scattering vector Q can be defined as follow (Lin et al., 2008):

$$225 \quad Q = \frac{4n\pi \sin\left(\frac{\theta}{2}\right)}{\lambda} \quad (5)$$

226 where n is the refractive index of the medium, λ , the laser light wavelength in vacuum, and θ , the
227 scattering angle.

228 High FD values generally represent densely packed flocs while low FD values characterise highly-
229 branched flocs with loosely-bound structure.

230 3 Results and discussion

231 3.1 Coagulation performance: Zeta potential and Turbidity, UV₂₅₄ and DOC removal

232 Coagulation performance of the three coagulants was assessed in terms of zeta potential and turbidity,
233 UV₂₅₄ and DOC removal efficiencies (Figure 1).

234 Zeta potential gives a measurement of the apparent surface charge and is therefore very useful to
235 evaluate the ability of a coagulant to destabilize the components in the feed water. In fact, the
236 reduction in magnitude of the negative zeta potential implies a reduction in the repulsive electrostatic
237 forces up to the point of zero charge (PZC) whereby the attractive van der Waals forces overcome the
238 repulsive forces and particles start to agglomerate (Gregory, 2005). Besides, the changes in floc zeta
239 potential have also been commonly used to identify the mechanisms involved in the coagulation
240 process (Gregory and Duan, 2001). Figure 1a shows that the magnitude of the negative zeta potential
241 values first decreased, up to a coagulant dose of 5 mg/L for all three coagulants, and then charge
242 reversal occurs at higher dose (floc zeta potential values become positive). These results suggest that
243 charge neutralization somewhat plays a dominant role in the coagulation of AOM for all three
244 coagulants. However, charge neutralization is unlikely to be the sole mechanism involved since the
245 highest removal efficiencies were achieved at a coagulant dose of 5 mg/L at which the floc zeta
246 potential values after coagulation were slightly negative (i.e. -4.3 mV, -2.03 mV and -4.1 mV for
247 FeCl₃, TiCl₄ and PTC respectively). These results corroborate with another study (Henderson et al.,
248 2008a) where it was suggested that a zeta potential operational window of -5 to 0 mV would provide
249 optimum removal of AOM. Besides, coagulation experiments were run at a pH between 7 and 8
250 which has been found to be the optimum pH range where the highest turbidity removal was achieved
251 for all three tested coagulants (Chekli et al., 2015; Zhao et al., 2011a; Zhao et al., 2011b). It is
252 interesting to note that similar removal efficiencies were observed at positive zeta potentials which
253 have been found to be typical of coagulation experiments conducted at pH 7. In fact, at the relatively
254 high dose required to induce charge reversal, removal efficiency is generally attributed to a change in
255 the dominant coagulation mechanism from charge neutralization to sweep flocculation whereby
256 coagulation is enhanced by adsorption to and/or enmeshment in metal hydroxide precipitates (Duan
257 and Gregory, 2003).

258 The coagulants performance in terms of turbidity (Figure 1b), UV₂₅₄ (Figure 1c) and DOC removal
259 (Figure 1d) shows that both Ti-based coagulants achieved better removal efficiencies than FeCl₃ and
260 the optimum coagulant dose was 5 mg/L for all three tested coagulants. This coagulant dose is much
261 lower than those found in previous studies focusing on the removal of organic matter in wastewater
262 (e.g. 8-20 mg/L in (Galloux et al., 2015)). One possible explanation could be that AOM can act as a
263 polymer aid which will decrease the amount of coagulant required (Henderson et al., 2008b). In an

264 earlier study (Zhao et al., 2013), it was found that PTC performed better or similarly to TiCl_4 in terms
265 of turbidity, UV_{254} and DOC removal, which is not consistent with the results obtained in the present
266 study. The difference in coagulation performance between TiCl_4 and PTC may be explained by the
267 difference in hydrolysed Ti species formed in the AOM feed solution. The dominant hydrolyzates for
268 TiCl_4 are most likely $\text{Ti}(\text{OH})_2^{2+}$ or $\text{Ti}(\text{OH})_3^+$ which can both form complex with negatively charged
269 AOM compounds (Zhao et al., 2013). For PTC, the lower performance can be due to the formation of
270 $\text{Ti}_n(\text{OH})_m^{4n-m}$ complexes with high m value such as $\text{Ti}(\text{OH})_3^+$ or $\text{Ti}(\text{OH})_4$ which will have lower
271 efficiency for the removal of negatively charged AOM compounds (Zhao et al., 2013).

272 **Figure 1**

273 **3.2 Detailed organic carbon fraction removal**

274 The detailed composition of the AOM feed solution before and after flocculation has been analysed
275 and fractionated (i.e. based on MW) by LC-OCD (Table 3). LC-OCD results can provide further
276 understanding of the mechanisms involved in AOM coagulation. In fact, LMW AOM has been shown
277 to cause inefficient flocculation and therefore demands a high coagulant load because crosslinking of
278 the LMW AOM with metal compounds is necessary to build flocs (Henderson et al., 2010). On the
279 other hand, high MW AOM (e.g. protein-rich AOM) can act as polymer aid and thereby increase the
280 coagulation efficiency (Henderson et al., 2010).

281 Following dilution with synthetic seawater, the AOM feed solution was mainly composed of building
282 blocks and LMW organics (i.e. 42% and 41%, respectively, of the total DOC) while the fractions of
283 biopolymers and humic substances were relatively low (i.e. less than 10%).

284 Table 3 shows the removal efficiencies of the different AOM fractions by all three coagulants. It can
285 be seen that the highest removal efficiencies were obtained for humic substances and LMW organics
286 (i.e. up to 90% removal efficiency) while the removal of biopolymers was significantly less efficient
287 (i.e. maximum removal efficiency of 48.1% obtained with TiCl_4 at 10 mg-Ti/L). These results are
288 consistent with the high turbidity and UV_{254} removal efficiencies obtained with all three coagulants
289 (Figure 1). In fact, biopolymers are dominated by hydrophilic polysaccharides that do not absorb
290 UV_{254} and since these high MW compounds (also known as TEPs) are transparent, turbidity
291 measurements neither detect their presence nor quantify their concentration (Li et al., 2016;
292 Tabatabai, 2014). Thus, these two parameters are not adequate for the measurement of biopolymers
293 but may provide useful information for the other organic carbon fractions such as UV_{254} for the
294 aromatic organic compounds (i.e. humic substances). It is therefore not surprising that the removal
295 efficiency for humic substances obtained with LC-OCD measurements follows the trend as the UV_{254}
296 removal efficiency shown in Figure 1c. Similarly, the high turbidity removal efficiency obtained in

297 this study reveals that biopolymers were not effectively removed by coagulation, as confirmed by the
298 low biopolymer removal efficiency measured by LC-OCD.

299 Both Ti-based coagulants, and more specifically TiCl_4 , performed better than FeCl_3 regardless of the
300 AOM fraction, which corroborates with previous studies where TiCl_4 performed better than FeCl_3 for
301 the removal of different organic carbon fractions in seawater (Jeong et al., 2013; Okour et al., 2009).
302 However, although these studies measured similar removal efficiencies for the LMW organics (i.e. up
303 to 93% at 5 mg-Ti/L), much higher removal efficiencies of biopolymers were obtained at relatively
304 low dose for both FeCl_3 and TiCl_4 (e.g. complete removal of biopolymers at 5 mg-Ti/L). In another
305 recent study (Tabatabai et al., 2014), FeCl_3 achieved a biopolymer removal efficiency of about 80% at
306 10 mg-Fe/L while in the present study, at the same coagulant dose, only 40% removal efficiency was
307 achieved.

308 The main difference between these studies and the present one relies on the composition of the feed
309 solution before coagulation. In fact, the fraction of high MW biopolymers was relatively high (i.e.
310 reaching 51% (Tabatabai et al., 2014) and in more balanced proportion in comparison to the other
311 organic carbon fractions, especially the LMW organics. In another recent study by Henderson et al.
312 (using aluminium sulfate as coagulant) (Henderson et al., 2010), the authors demonstrated that some
313 algae species (and related AOM) require a higher coagulant demand due to the presence of LMW
314 AOM compounds while high removal efficiencies were achieved at relatively low coagulant dose for
315 species that have AOM with high MW compounds. Higher removal efficiencies were also obtained
316 with highly charged species (Henderson et al., 2010). The AOM feed solution used in the present
317 study had a high fraction of LMW organics (i.e. about 40% of the total DOC) combined with a
318 relatively low charge (i.e. initial zeta potential of $-9.98 \text{ mV} \pm 0.39 \text{ mV}$), which probably caused the
319 lower removal efficiencies at similar coagulant dose.

320 **Table 3**

321 **3.3 Dynamic variation of floc size, floc breakage and recovery and floc structural analysis**

322 Initial slow mixing (at 40 rpm) allowed a gradual increase of floc sizes for all three coagulants up to a
323 steady-state floc size plateau after 3 to 5 minutes (Figure 2). When a shear force (200 rpm for 1
324 minute) was introduced after 15 minutes, the floc size immediately decreased to 73%, 55% and 52%
325 of the average floc size at steady state for FeCl_3 , TiCl_4 and PTC, respectively (d_1 and d_2 –Table 4).
326 Upon reintroduction of slow mixing, the flocs started to regrow; however, none of the flocs formed by
327 any of the three coagulants recovered back to their initial size before breakage (d_1 and d_3 –Table 4).
328 This irreversible breakage of flocs is evidence that the flocs were to some extent formed by sweep
329 flocculation, suggesting that the flocs were held together by chemical bonds (e.g. carboxyl and phenol

330 groups of organic molecules), which do not have the capacity to reform after breakage (Jarvis et al.,
331 2005).

332 **Figure 2**

333 Results in Table 4 also show the calculated floc growth rate, SF and RF based on equations (1), (2)
334 and (3). Both Ti-based coagulants exhibited a faster growth rate and formed larger flocs at each stage
335 (i.e. steady-state, after breakage and after regrowth) compared to the conventional FeCl₃ coagulant
336 (Table 4), which corroborates with previous studies comparing the performance of these three
337 coagulants (Chekli et al., 2015; Galloux et al., 2015). The floc sizes obtained at steady state (i.e. d₁ –
338 Table 4) were larger than those obtained in previous studies (Chekli et al., 2015; Galloux et al., 2015).
339 This can be explained by the composition and MW distribution of the organic carbon compounds
340 present in the initial feed solution. In fact, in previous studies, synthetic or coal-mining wastewater
341 were used as feed whereby the DOC concentration was either much lower or mainly composed of
342 humic substances with narrower MW distribution consisting of lower MW compounds. The presence
343 of biopolymers in AOM feed solution, even at low concentration, can enhance the coagulation process
344 by acting as polymer aid resulting in the formation of larger flocs. The larger floc sizes obtained in
345 this study could also be explained by the presence of divalent cations such as Ca²⁺ in the synthetic
346 seawater. In fact, divalent cations can enhance the coagulation performance in the presence of
347 negatively charged organic compounds due to bridging effects resulting in the formation of (Ca-
348 AOM-Metal hydrolyzates) complexes at low coagulant dose and (Ca-AOM-Metal hydroxide
349 precipitates) complexes at high coagulant dose (Zhao et al., 2014).

350 Floc strength and recovery factors (SF and RF) were calculated to investigate the capacity of the flocs
351 to resist an applied shear force and to regrow after exposure to this force. Both TiCl₄ and PTC showed
352 higher SF than FeCl₃ (i.e. 47.9% and 45.1 % for PTC and TiCl₄ against 27.1% for FeCl₃) while TiCl₄,
353 alone showed the highest RF (i.e. 70.5%). These results suggest that charge neutralisation could be the
354 main mechanism for TiCl₄ while sweep flocculation might have also contribute to a certain degree for
355 both FeCl₃ and PTC. In fact, previous studies have reported that flocs formed by sweep flocculation
356 have a weaker recovery after breakage than those formed by charge neutralisation (Aguilar et al.,
357 2003).

358 **Table 4**

359 The floc fractal dimension (FD) was also evaluated for all three coagulants before breakage and after
360 regrowth using equations (4) and (5). Floc FD is an important factor representing the floc density,
361 which impacts greatly on the efficacy of the solid/liquid separation process (Gregory, 1998). Results
362 in Table 4 showed that the floc FD values followed the order of FeCl₃ (1.99) < TiCl₄ (2.14) ≤ PTC
363 (2.2). The FD values obtained in this study are typical for flocs generated in water treatment processes
364 (i.e. 1.5-2.2) (McCurdy et al., 2004). Results also indicate that the floc FD values follow the same

365 trend as the floc SF values; suggesting that flocs having a better ability to resist a shear force will also
366 present a higher degree of compactness. This can be explained by the strong relationship between the
367 floc breakup mechanism and the floc structure. In fact, there are two main breakup mechanisms:
368 surface erosion (slow) and large-scale fragmentation (fast) (Jarvis et al., 2005). Flocs with low FD
369 will preferentially breakup via a fragmentation mechanism during which floc aggregates will split into
370 pieces of comparable size, leading to a low SF. However, compact flocs with a high FD will breakup
371 via surface erosion where small aggregates are separated from the surface of the main aggregated floc,
372 leading to a higher SF. This result has also been demonstrated in a previous study where a close
373 relationship between floc structure and floc strength was established (Wang et al., 2009). The
374 discernible difference between FeCl₃ and Ti-based coagulants could be explained by the amount of
375 AOM adsorbed within the flocs, which is higher for both Ti-based coagulants (i.e. higher DOC
376 removal – Figure 1d). In fact, it has been previously demonstrated for inorganic colloids that the
377 adsorption of organic macromolecules can induce a change in aggregate structure (Baalousha et al.,
378 2008). Christian et al. (2008) also explained that the adsorption of organic macromolecules on
379 aggregate surfaces can occur in two steps. First, the organic macromolecules may cover the surface of
380 the aggregates in a fast adsorption step which could then be followed by a second step whereby slow
381 diffusion of organic macromolecules within the aggregates pores occurs, resulting in the formation of
382 more compact aggregates. Finally, results in Table 4 show that the FD values after regrowth were
383 slightly higher than before breakage. In fact, it has been demonstrated that cyclic breakup and re-
384 aggregation may lead to more compact aggregates through rearrangements at more favourable points
385 resulting in more stable structures (Chakraborti et al., 2003; McCurdy et al., 2004).

386 Figure 3 displays the floc size distribution of the three coagulants before breakage (i.e. at steady state)
387 and after regrowth. Flocs formed by FeCl₃ were polydispersed in size with the presence of smaller
388 flocs (i.e. additional peak with smaller volume at about 700 μm) while the flocs formed by both Ti-
389 based coagulants have a much narrower size distribution with the presence of only one peak at around
390 1100 μm. The presence of smaller flocs in the case of FeCl₃ is not advantageous since they tend to
391 remain in suspension or settle more slowly after flocculation as evidence for the lower turbidity
392 removal efficiency displayed in Figure 1b. After regrowth, a much broader floc size distribution
393 towards lower floc size was obtained for all three coagulants, which corroborates with the lower
394 average floc size obtained after regrowth (i.e. d₃ in Table 4). However, the volume of the main peak
395 (i.e. observed before breakage – Figure 3a) remains relatively high after regrowth for TiCl₄ (Figure
396 3b) which supports the high RF value obtained for this coagulant.

397 **Figure 3**

398

399

400 **Conclusions**

401 This study investigated the performance of titanium salts as alternative coagulants for the removal of
402 AOM in seawater. The results were systematically compared with those obtained with conventional
403 FeCl₃ coagulant. The key findings can be summarised as follow:

- 404 • Both titanium salts (i.e. TiCl₄ and PTC) performed better in terms of turbidity, UV₂₅₄ and
405 DOC removal than conventional FeCl₃ at the same dose. The presence of AOM acting as
406 coagulant aid combined with the presence of divalent cations (e.g. Ca²⁺) promoting bridging
407 mechanisms resulted in similar performance at low dose for all coagulants tested.
- 408 • LC-OCD measurements revealed that both humic substances and LMW organics were
409 preferentially removed by all three coagulants while the removal efficiency for biopolymer
410 was relatively low (i.e. about 15-50% removal efficiency). This was attributed to the presence
411 of high proportion of LMW organics (i.e. 40% of the total DOC) interfering/competing with
412 the flocculation process of other higher MW compounds.
- 413 • Detailed analysis of the floc characteristics showed that both TiCl₄ and PTC can grow faster
414 and reach larger floc size than FeCl₃ while having a better ability to resist shear force (i.e. SF
415 of about 45%, 48% and 27% for TiCl₄, PTC and FeCl₃, respectively). This is highly
416 advantageous since many water treatment processes (e.g. desalination plants) are non-ideal
417 with many potential zones of high shear forces that could induce floc breakage. The high
418 recovery factor obtained with TiCl₄ (i.e. about 70%) is another obvious benefit.
- 419 • Finally, the floc FD followed the same trend as the floc SF indicating that flocs that have a
420 better ability to resist shear force are also more compact and will therefore break up via
421 surface erosion rather than large-scale fragmentation. The higher floc FD obtained with both
422 TiCl₄ and PTC was related to their higher DOC removal efficiency.

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