# Molecular physiological responses and acclimation of the seagrass species *Z. muelleri* to light limitation



A thesis submitted to the University of Technology Sydney for the degree of DOCTOR OF PHILOSOPHY (PhD)

Submitted January, 2017

Peter A. Davey, BSc. Hons, MSc. Supervisors: Professor Peter Ralph, Dr Mathieu Pernice and Dr Rudy Dolferus

The Thesis presented meets the standards and requirements set out by the University of Technology Sydney

#### **Certificate of original ownership**

I certify that the work in this thesis has not previously been submitted for a degree nor has it been submitted as part of requirements for a degree except as part of the doctoral degree and/or fully acknowledged within the text. I also certify that this thesis has been written by me (Peter A. Davey). Any help that I have received in my research work and the preparation of this thesis itself has been acknowledged. In addition, I certify that all information sources and literature used are indicated in this thesis.

#### Acknowledgments

I would first like to thank Professor Peter Ralph for giving me the exciting opportunity to work within the Climate Change Cluster (C3) at the University of Technology Sydney, and my co-supervisors, Dr Mathieu Pernice and Dr Rudy Dolferus for the advice and time they dedicated to helping me over the duration of the project. I would also like to thank Dr Martin Schliep, for his support as my co-supervisor in the first year of my PhD. This research was supported by the UTS President's Research Scholarship, UTS International Research Scholarship, and a CSIRO Office of Chief Executive post-graduate scholarship.

I acknowledge Dr Gaurav Sablok for his bioinformatics based teachings, and thank the following colleagues for providing fruitful advice and discussion; Manoj Gupta Kumar; Unnikrishnan Kuzhiumparambil; Justin Ashworth; Chris Evenhuis; Nasim Mohammadi; Kasper Brodersen; Alex Thomson; Stacey Trevathan-Tackett; Milan Szabo; Tony Larkum; Rachel Levin; and John Raven. In relation to fieldwork, the TropWater team associated with James Cook University (Queensland), Katherina Petrou (UTS), Frederic Cadera (visiting scholar) and Vincent Schols (visiting scholar) should be thanked. I thank the lab technicians; Paul Brooks; Stacey Ong; Gemma Armstrong; Peter Jones; Graham Powolski; and Kun Xie for providing first-class support in the laboratories. On many occasions, Mike Lake (IT Department, UTS) Brian Haas (Broad Institute, USA), Don Gilbert (Indiana University, USA), members of the 'biostars.org' and 'seqanswers' online bioinformatics forums should be thanked for providing me with insight on how to solve numerous computational, bioinformatics and software related issues.

Finally, I wish to express my gratitude for the love and support that my family and friends gave me during this journey; My parents; my brother Steven; my late grandmother and my partner Emma; I thank them for encouraging me to undertake a PhD degree on the other side of the world, and for their continuous support and encouragement during my highs and lows.

#### Preface

The Chapters within this PhD thesis have been written with the intention of submission to scientific journals. The chapters are therefore presented in a typical journal format, ready for submission. Chapter 1 has already been published in a scientific journal (Functional and Integrative Genomics; IF = 2.265) as a critical literature review. Chapters 4 and 5 will be submitted in the near future to scientific journals as original research articles. Scientific work, which I have contributed to, is listed in Appendix 4, one of these pieces of work has been published in another journal (Frontiers in Plant Science; IF = 4.495), whilst the other piece of work (a book chapter) is in preparation. Given that this thesis is presented as a series of ready to submit manuscripts, there is an element of repetition in the introduction of some of the chapters.

#### A foreword

## "Look deep into nature, and then you will understand everything better" - Albert Einstein

Personally for me, the above quote sums up my PhD journey over the past three and a half years. Only through bioinformatics analyses, did true meaning come from the observations I made in the laboratory at UTS. To unravel the complexity of one organism over three and a half years has been a huge accomplishment for me, one that I have immenseley enjoyed; however, with the satisfaction came the challenge, one that I found testing at times. By undertaking this PhD, I feel that I have come a long way, learning about myself, and seagrasses in many ways. A journey, which gave me appreciation for how complex nature can be. As the saying goes - "There's more than meets the eye."

### General Abbreviations A to Z

2D-IEF	Two Dimensional- Isoelectric Focusing
ABA	Abscisic Acid
AFLP	Amplified Fragment Length Polymorphism
AL-fact	Actinic Light Factor
AMSA	Australian Marine Sciences Association
ANU	Australian National University
ATP	Adenosine 5'-triphosphate
AU	Arbitrary Units
BBH	Bidirectional Best BLAST Hit
BCV	Biological Coefficient of Variation
BLAST	Basic Local Alignment Search Tool
bp	Base pairs
BP	Biological Process
BUSCO	Benchmarking Universal Single-Copy Orthologs
BWA	Burrows-Wheeler Aligner
C3	C3 photosynthesis / Climate Change Cluster
C4	C4 photosynthesis
CA	Carbonic Anhydrase
CAP3	Contig Assembly Program 3
CC	Cellular Component
ССМ	Carbon Concentrating Mechanism
cDNA	complementary Deoxyribonucleic Acid
CEGMA	Core Eukaryotic Genes Mapping Approach
CHIP-Seq	Chromatin Immunoprecipitation-Sequencing
chl	Chlorophyll
Ci	Inorganic Carbon
СРМ	Counts Per Million
CRBB	Conditional Reciprocal Best BLAST
Ct	Cycle threshold
Cyt.b6f	Cytochrome b6f
ddRAD	double digest Restriction Associated DNA
DNA	Deoxyribonucleic Acid
dNTP	Deoxyribonucleotide triphosphate
DOGMA	Dual Organellar GenoMe Annotator
DVC A	Divinyl chlorophyllide A
EBI	European Bioinformatics Institute
EMBL	European Molecular Biology Laboratory
EST	Expressed Sequence Tag
ETR	Electron Transport Rate
FC	Fold Change
FDM-SSR	Functional Domain Marker - Simple Sequence Repeat
FDR	False Discovery Rate

FPKM	Fragments Per Kilobase of transcript per Million mapped reads
Fv/Fm	Maximum quantum yield of photosystem II
G3P	Glyceraldehyde 3-phosphate
GABA	Gamma-aminobutyric acid
GBS	Genotype By Sequencing
GC-MS	Gas Chromatography Mass Spectrometry
gDNA	genomic Deoxyribonucleic Acid
GGPP	Geranylgeranyl diphosphate
GLM	Generalized Linear Model
GO	Gene Ontology
GSEA	Gene Set Enrichment Analysis
HPC	High Performance Computing
HPLC	High Performance Liquid Chromatography
Ik	Half-saturation constant
ISBW	International Seagrass Biology Workshop
KAAS	KEGG Automatic Annotation Server
KEGG	Kyoto Encyclopedia of Genes and Genomes
LC-MS/MS	Liquid Chromatography Tandem-Mass Spectrometry
LHC	Light Harvesting Complex
LHCI / LHCA	Photosystem I Light Harvesting Complex
LHCII/ LHCB	Photosystem II Light Harvesting Complex
LL	Light Limited
MDS	Multi-Dimensional Scaling
MF	Molecular Function
Meas. Int	Measuring Intensity
MEME	Multiple EM for Motif Elicitation
MEP	2-C-methyl-D-erythritol 4-phosphate
MIQE	Minimum Information for Publication of Quantitative. Real- Time PCR Experiments
miRNAs	micro Ribonucleic Acid
MISA	MIcroSAtellite identification tool
mRNA-Seq	Messenger RNA-Sequencing
MTDB	Mangrove Transcriptome Database
mya	million years ago
NADP	Nicotinamide adenine dinucleotide phosphate
NADPH / NADP(H)	Reduced nicotinamide adenine dinucleotide phosphate
NCBI	National Center for Biotechnology Information
ncRNAs	non-coding Ribonucleic Acids
NGS	Next Generation Sequencing
No-RT	No Reverse Transcriptase
NPQ	Non-Photochemical Quenching
NRQ	Normalised Relative Quantity
NSW	New South Wales

NZGL	New Zealand Genomics Limited
ORF	Open Reading Frame
P-I	Photosynthesis - Irradiance
PAM	Pulse Amplitude Modulated
PC	Plastocyanin
PCR	Polymerase Chain Reaction
PDA	Photodiode Array Detector
PEPC	Phosphoenolpyruvate carboxylase
PhiX	Enterobacteria Phage PhiX174
polyA	Polyadenylated
P(MC)	Probability (Multiple Correction)
PPOs	Polyphenol Oxidases
PQ	Plastoquinone
PSI	Photosystem I
PSII	Photosystem II
psu	Practical salinity units
PTFE	Polytetrafluoroethylene
qPCR	Quantitative Polymerase Chain Reaction
QC	Quality Check
QTL	Quantitative Trait Loci
RAPD	Random Amplification of Polymorphic DNA
RBH	Reciprocal Best BLAST Hit
rETRmax	relative maximum Electron Transport Rate
RFLP	Random Fragment Length Polymorphism
RIN	RNA Integrity Number
RLC	Rapid Light Curve
RNA	Ribonucleic Acid
RNA-Seq	RNA-Sequencing
ROI	Reactive Oxygen Intermediate
ROS	Reactive Oxygen Species
RPKM	Reads Per Kilobase of transcript per Million mapped reads
RSEM	RNA-Seq by Expectation-Maximization
RT-qPCR	Real Time-quantiative Polymerase Chain Reaction
RuBisCo	Ribulose-1,5-bisphosphate carboxylase/oxygenase
RubP	Ribulose-1,5-bisphosphate
SAM	Sequence Alignment Map
Sat. Width	Saturation Width
Sat.Int	Saturation Intensity
SNP	Single Nucleotide Polymorphism
sRNA	small Ribonucleic Acid
SRP	Signal Recognition Particle
SSL	Super Saturating Light
SSR	Simple Sequenc Repeat

STAR	Spliced Transcripts Alignment to a Reference
TAIR	The Arabidopsis Information Resource
TBAA	tert-Butyl acetoacetate
TCA	Tricarboxylic Acid
TF	Transcription Factor
Tm	Melting temperature
TMM	Trimmed Mean of M-Values
TPM	Transcripts Per Million
UNIX	UNIX programming environment
UPLC	Ultra Performance Liquid Chromatography
UV	Ultra Violet
Yi	Initial quantum yield of photosystem II

#### List of figures

**Figure 1:** Illustrative diagram collectively showing the specialised traits of seagrasses, which allow for seagrasses to live a submerged life in the coastal marine environment. Information sources: Marbà et al., 2002; Ackerman, 2006; Larkum et al., 2006; Touchette, 2007; Broderson et al., 2015; Hasler-Sheetal et al., 2015; Olsen et al., 2016. Illustrative model concept is based on the *Zostera* genus.

**Figure 2:** The advances in seagrass molecular profiling and omics to date. The technologies that have been utilised are shown along the bottom of the illustration, whilst types of study conducted to date are shown at the top of the illustration.

Figure 3: The physical light environment of A. terrestrial plants and, B. sub-43tidal marine plants. Black arrows represent light scattering and attenuation9processes. Spectral light availability is represented by a wavelength9spectrum bar.9

Figure 4: The light dependent reaction of photosynthesis (Z-scheme) in
45 higher plants. PQ = Plastoquinone; Cyt.b6f = Cytochrome b6/f complex; PC
= Plastocyanin. P680 refers to photosystem II and P700 refers to photosystem I. Positive (+) characters refer to excited state of the photosystems.

Figure 5: Schematic diagram of chlorophyll excitation in response to high49light, the various fates of the excited chlorophyll molecule are represented.In association with NPQ, the role of the psbS protein and the xanthophyllcycle are shown.

**Figure 6:** Experimental set-up of the artificial mesocosm study with varying 64 light intensities:  $\sim 1,000 \mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>;  $\sim 250 \mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>; and  $\sim 10 \mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>. Aquarium related equipment is labelled in the legend above. Temperature was kept at 24°C and salinity between 32 and 36 units.

Figure 7: A schematic flow diagram representing the methodology used to66assemble, characterise and profile the Z. *muelleri* transcriptome.

Figure 8: Venn diagrams representative of the GO terms enriched in each of71the light conditions. Labelling units are in  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>.

8

9

Figure 9: The transcription factors family/types (represented by a minimum74of 50 occurrences) identified in the Z. muelleri transcriptome.

Figure 10: Pie chart showing the composition (in %) of SSR sequence types75in Z. muelleri.

Figure 11: Scaled topographical map of Narrabeen Lakes. Sampling site is92indicated by a blue star. North arrow and scale represented. Map createdusing ArcGIS online (esri).

**Figure 12:** Experimental protocol and layout. A. Mesocosm layout detailing **93** acclimation period, experimental treatments and sample collection time points. Light units are  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>. B. mRNA-Seq genome guided pipeline to determine transcriptome and differential gene expression, green reads = 200  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>; red = 20  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup> and orange = genome. C. Physiological and photobiology measurements recorded including the pigments analysed. Plants for RNA-sequencing, pigment analysis and rapid light curve analysis were all independent of one another.

**Figure 13:** A – G. Pigment concentrations ( $\mu$ g/ gram fresh weight in control and light limited seagrass plants; n = 3). Significance is represented by an asterix (\*). H. Relative electron transport rate maximum of control and light limited plants across Days 2 and 6 (n = 5). All error bars are representative of standard deviation of mean. Ctl = Control, LL = Light limited.

99

**Figure 14:** Differential gene expression represented by hierarchical **101** clustered heatmaps. Normalised LogCPM gene counts are centered and z-scaled. D2 = Day 2, D6 = Day6. Light limited replicates (L1-3; 20  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>) and control plant replicates (C1-3; 200  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup>) are indicated.

Figure 15: Bootstrapped hierarchical clustering of correlated transcripts. A. 102A cluster of correlated transcripts that decreased in expression under lightlimitation. B. Three closely related clusters of transcripts that increased in expression under light limitation on Day 6.

Figure 16: Significant photosynthetic gene regulation on Day 2 (FDR104threshold = 0.05; fold change of 1). Significant down-regulation of many<br/>genes can be observed. CCS1 = Cytochrome C biogenises protein; NDF1 =NDH-DEPENDENT CYCLIC ELECTRON FLOW 1; Cyt b6/f =

Cytochrome b6/f sub-unit; PGR5 = Proton Gradient Regulation 5; LTD = Light harvesting complex translocation defective protein; OHP = One helix proteins; FtsH = ATP-dependent Zinc metalloproteases; YCF = hypothetical chloroplast open reading frame genes; psb = photosystem two sub units; ELIPs = Early light induced protein genes.

**Figure 17:** Significant gene regulation (FDR = 0.05; fold change of 1) associated with the carotenoid, xanthophyll, tocopherol, chlorophyll and ABA synthesis pathways on Day 2 of light limitation. *PSY* = Phytoene synthase; *PDS* = Phytoene desaturase; *ZDS* = Zeta carotene desaturase; *CRTISO* = Carotenoid isomerase / Prolycopene isomerase; *LYCB* = Lycopene beta-cyclase;  $\beta$ -OHase =  $\beta$ -carotene hydroxylase;  $\epsilon$ -OHase =  $\epsilon$  hydroxylase; *ZEP* = Zeaxanthin epoxidase; *VDE* = Violaxanthin deepoxidase; *CCDs* = Carotenoid cleavage dioxygenases; *NCED* = Nine cisepoxycarotenoid dioxygenase; *POR* = Protochlorophyllide; *FLU* = Blue fluorescent light; *Chl.red B NOL* = Chlorophyllide B reductase NOL; *ChlH* = Mg chelatase sub-unit H; *DVR* = Divinyl chlorophyllide A 8-vinyl-reductase; *VTE* = Tocopherol cyclase; *gamma-TMT* = *Tocopherol O*-methyltransferase; *ABA 8'-OHase 3* = Absicic acid 8' hydroxylase 3; *NSY* = Neoxanthin synthase.

Figure 18: Relative ETR<sub>max</sub> of Z. muelleri plants during pre-treatment and135Day 4 in control (dark grey) and light limited treatments (white). Asterix (\*)= statistical significance observed (p<0.05) between treatments.</td>

Figure 19: Gene stability measure of chloroplast-encoded reference genes137only. Names and average M-stability values are shown on the plot.

Figure 20: Gene stability measure of chloroplast-encoded and nuclear-137encoded reference genes combined. Names and average M-stability valuesare shown on the plot. ndhC, rpoC2, rps11 and rps2 are chloroplast-encodedgenes. GDPH and PP2Aa are nuclear-encoded genes.

Figure 21: A. Target genes normalised to nuclear-encoded reference genes 139 (Normalised Relative Quantity – NRQ). B. Target genes normalised to chloroplast-encoded reference genes. Asterix (\*) and brackets indicate statistically significant difference between control and light limited (low light) *Z. muelleri* plants. n = 3.

105

Figure 22: Schematic diagram highlighting the trends of how Z. muelleri156responds to light limitation over a period of 14 days.

A.1; Figure 1: Plant GO-Slim characterisation of the Z. *muelleri* 168 transcriptome 'Biological Processes'. A threshold of 250 or more unigenes per GO term was applied. Level 3 and below (Blast2GO software) are only shown.

A.1; Figure 2: Plant GO-Slim characterisation of the Z. *muelleri* 169 transcriptome 'Molecular Functions'. A threshold of 250 or more unigenes per GO term was applied. Level 3 and below (Blast2GO software) are only shown.

A.1; Figure 3: Plant GO-Slim characterisation of the Z. *muelleri*170 transcriptome 'Cellular Compartments'. A threshold of 250 or more unigenes per GO term was applied. Level 3 and below (Blast2GO software) are only shown.

A.2; Figure 1: RLC analysis of Z. *muelleri* plants from Narrabeen Lakes (n
191
= 6) to determine control irradiance for the experiment during Australian winter, 2015. Standard deviation represented by error bars.

A.2; Figure 2: Odyssey light logger data from the experiment across 6 days: 191
18/09/15 to the 23/09/15. Mid-day spikes are caused by single time point sampling effort disturbance.

A.2; Figure 3: Chromatogram of the pigment peaks identified in HPLC 192 analysis. Retention time (minutes) shown on X axis. Intensity (AU) shown on Y-axis.

A.2; Figure 4: Biological coefficient of variation between RNA-Seq
192 samples within the experiment. Common dispersion is represented in green.
A BCV value of 0.24 was obtained based on the common dispersion.
Trended and tagwise dispersions are also represented; however, common dispersion is used to gauge variation within EdgeR (Mccarthy et al. 2012).

A.2; Figure 5: Redundant or duplicated transcripts that are present in the 193 draft genome of *Z. muelleri* (Lee et al. 2016) and are correlated in expression. The correlation of expression is plotted for 1,051 differentially expressed transcript models in this study versus the percent nucleotide identity against redundant, similar or duplicated transcript models in the

draft genome.

A.2; Figure 6: Multi-dimensional scaling (MDS) plot showing similarities
between RNA samples (Leading LogFC dimension 1 Vs Leading logFC dimension 2). D2 = Day 2, D6 = Day 6, L1-L3 = LL plants 1 – 3, C1-C3 = Control plants 1-3.

A.2; Figure 7: DNA sequence motifs detected by MEME in the putative 194 upstream promoter regions of A. Genes down-regulated under light limitation by Day 2 and, B. Genes up-regulated under light limitation by Day 6.

#### List of tables

**Table 1:** Known to us, the molecular profiling and omic studies published since the establishment of the seagrass NGS era in 2006. A comprehensive search was conducted on 'Web of science' for the topics 'seagrass sequencing', 'seagrass gene expression', 'seagrass proteomics', 'seagrass metabolomics'. Additional studies based on personal knowledge are also provided. Last accessed: June, 2016.

**Table 2:** The number of polymorphic SSR loci and SNP loci validated in25seagrass species to date. Organised by number of polymorphic loci perspecies. A search was conducted using 'Web of Science' with the searchterms 'microsatellite loci seagrass', 'microsatellite markers seagrass','polymorphic microsatellite seagrass' and 'polymorphic loci seagrass'. Lastaccessed: June, 2016.

**Table 3:** Summary of statistics of the original and final Z. muelleri de novo69assemblies.

**Table 4:** Coverage distribution of the Z. muelleri transcriptome across related69plant species. Proteome = number of genome predicted proteins, Z. muelleriunigenes =52,616. CRBB = Conditional Reciprocal Best BLAST. Referenceproteomes were downloaded from ftp://ftp.ensemblgenomes.org/pub/plants/release-32. The Spirodela polyrhiza proteome was downloaded fromhttp://spirodelagenome.org, Zostera marina from http://uniprot.org.

**Table 5:** Number of unigenes distributed across GO categories putatively72involved in light associated processes and light harvesting.

**Table 6:** Statistical comparisons between control and light limited plants on**98**Days 2 and 6. Median, p-values, U values and Z-test scores are reported forMann-Whitney-U statistical tests. Significant (P<0.05) results are indicated</td>by an asterix (\*). The number of biological replicates was n = 5, except forDay 2 when n = 4 for light limited plants (loss of one replicate sample).

**Table 7:** Chloroplast / nuclear-encoded reference genes, and chloroplast-131encoded target genes validated and profiled in this study. Gene names areprovided (abbreviated and in full), along with primer melting temperatures(Tm, °C), oligo sequences, amplicon size (AS; bp), amplification efficiencies(E; %) and standard curve co-efficient values (R²).

**Table 8:** Statistical comparisons between control and light limited plants on136Day 4. Median, p-values, U values and Z-test scores are reported for Mann-<br/>Whitney-U statistical tests. Significant results are indicated by an asterix (\*),<br/>n = 5.

**Table 9:** Significance of gene expression change between control and light138limited plants when normalised to chloroplast-encoded (cp) reference genesand nuclear-encoded (nc) reference genes. P(MC) = P-value (multiplecorrection), T = pairwise t-test value. Statistical significance (p<0.05) is</td>represented by an asterix (\*).

A.1; Table 1: Number of paired end reads (bp) in each library used to 171 assemble the *Z. muelleri* transcriptome.

A.1; Table 2: Functional distribution hit analysis showing top 5 most 171 represented plant species in DIAMOND protein alignment searches (E-value of  $1.0 \ge 10^{-5}$ .); 52,616 = total number of best hits retained.

**A.1: Table 3:** Enriched GO terms in ~10  $\mu$ mol photons m<sup>-2</sup>s<sup>-1</sup> and ~250  $\mu$ mol photons m<sup>-2</sup>s<sup>-1</sup> of light combined. GO = Gene Ontology, NS = Classification, Name = Name of GO term, FDR threshold = 0.05. BP = Biological Process; CC = Cellular Compartment and MF = Molecular Function.

A.1; Table 4: Enriched GO terms in ~10  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup> of light. GO = 174 Gene Ontology, NS = Classification, Name = Name of GO term, FDR threshold = 0.05. BP = Biological Process; CC = Cellular Compartment and MF = Molecular Function.

A.1; Table 5: Enriched GO terms in ~250  $\mu$ mol photons m<sup>-2</sup>s<sup>-1</sup> of light. GO = 179 Gene Ontology, NS = Classification, Name = Name of GO term, FDR threshold = 0.05. BP = Biological Process; CC = Cellular Compartment and MF = Molecular Function.

**A.1; Table 6:** Enriched GO terms in ~1,000  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup> of light. GO **182** = Gene Ontology, NS = Classification, Name = Name of GO term, FDR = 0.05. BP = Biological Process; CC = Cellular Compartment and MF = Molecular Function.

**A.1: Table 7:** PlantGSEA metabolic pathway enrichment of terms in ~1,000 **184**  $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup> of light. FDR threshold = 0.05.

A.1: Table 8: PlantGSEA metabolic pathway enrichment of terms in ~250 186

 $\mu$ mol photons m<sup>-2</sup> s<sup>-1</sup> of light. FDR threshold = 0.05.

A	.1:	Table	9:	PlantGSEA	metabolic	pathway	enrichment	of terms	s in /	~10	187
μr	nol	photor	ıs n	n <sup>-2</sup> s <sup>-1</sup> of light	FDR three	shold $= 0.0$	05.				

A.1; Table 10: The top ten most common SSR repeat motifs found within the 189*Z. muelleri* transcriptome.

A.2; Table 1: Xanthophyll and carotenoid pigments profiles on days 2 and 6 (n = 3). Significant results are highlighted with an asterix (\*) character (P<0.05). Independent student t-tests were conducted.

A.2; Table 2: Chlorophyll pigments profiles on days 2 and 6 (n = 3). 195 Significant results are highlighted with an asterix (\*) character (P<0.05). Independent student t-tests were conducted.

A.2; Table 3: Number of paired end reads (bp) in each library used for the 196 assembly of the genome-guided transcriptome.

A.2; Table 4: Top 10 species/ genus groups with functional best hits to the Z.197*muelleri* transcriptome.

A.2; Table 5: Differentially regulated genes associated with photosynthesis 198 and carbon fixation on Day 2. FC = Fold change, FDR = 0.05.

**A.2; Table 6:** Differentially regulated genes associated with photosynthesis **200** and carbon fixation on Day 6. FC = Fold change, FDR = 0.05.

**A.2; Table 7:** Differentially regulated genes associated with photosynthetic **201** pigments and ABA (absicic acid) on Day 2. FC = Fold change, FDR = 0.05.

**A.2; Table 8:** Differentially regulated genes associated with photosynthetic **203** pigments and ABA (absicic acid) on Day 6. FC = Fold change, FDR = 0.05.

**A.2; Table 9:** Differentially regulated genes associated with light reception 205 and signaling on Day 2. FC = Fold change, FDR = 0.05.

A.2; Table 10: Differentially regulated genes associated with light reception 207 and signaling on Day 6. FC = Fold change, FDR = 0.05.

A.2; Table 11: Enriched GO terms in the up-regulated sub-set of genes in LL
208
plants on Day 2. FDR = 0.05. NS = Classification; MF = Molecular Function;
BP = Biological Process; CC = Cellular Compartment.

A.2; Table 12: Enriched GO terms the down-regulated sub-set of genes in LL
209
plants on Day 2. FDR = 0.05. NS = Classification; MF = Molecular Function;
BP = Biological Process; CC = Cellular Compartment.

A.2; Table 13: Enriched GO terms in the up-regulated sub-set of genes in LL
plants on Day 6. FDR = 0.05. NS = Classification; MF = Molecular Function;
BP = Biological Process; CC = Cellular Compartment.

A.2; Table 14: Enriched GO terms in the down-regulated sub-set of genes in
LL plants on Day 6. FDR = 0.05. NS = Classification; MF = Molecular
Function; BP = Biological Process; CC = Cellular Compartment.

A.2; Table 15: Enriched GO terms in the down-regulated sub-set of genes in
216
LL plants on Day 6 compared to LL plants on Day 2. FDR = 0.05. NS =
Classification; MF = Molecular Function; BP = Biological Process; CC =
Cellular Compartment.

A.3; Table 1: Comparison of genome-guided assembly and *de novo* super 218 assembly statistics.

#### CONTENTS

Thes	iesis summaryX						
PhD	thesis aims and objectives	1					
СНА	PTER 1	2					
Abst	ract	3					
1.1	Introduction						
1.2	Seagrasses: A unique group of plants						
1.3	The current status of omics and molecular profiling in seagrass biology	5					
1.4	Seagrass light perception and response at the molecular level	.10					
1.5	Carbon fixation in seagrasses - challenging old beliefs with new technology	.13					
1.6	Are stress and environmental response signatures between land and marine plants different?	.14					
1.7	Osmoregulation at the molecular level	.16					
1.8	Tolerance to anoxia and phytotoxic sediment	.18					
1.9	Other areas needing investigation and development	.19					
1.10	Genetic marker utilisation in seagrass biology	.22					
1.11	Development and advancement of molecular markers	.22					
1.12	SSR markers: High-resolution popularity	.23					
1.13	Is there a need for further advances in the field of genetic markers?	.24					
1.14	Concluding remarks	.26					
1.15	Acknowledgements	.27					
1.16	References	.28					
CILL		40					
	PTER 2	.40					
2.1	Light and photosynthesis: terrestrial vs. marine environments	.42					
2.2	Organelles responsible for light absorption in higher plants	.44					
2.3	Light-dependent reaction of photosynthesis	.44					
2.4	Photosystem-light harvesting complexes4						
2.5	Chlorophyll pigments						
2.6	Carotenoids and the xanthophyll pigments	.47					
2.7	The xanthophyll cycle and non-photochemical quenching	.48					
2.8	Anthocyannins	.49					
2.9	Sensing quality and quantity of environmental light	.50					
2.10	Common low light responses in plants	.51					
2.11	Our knowledge of light response in Z. <i>muelleri</i> to date	.52					
2.12	References	.54					
СНА	PTER 3	.60					
Abst	ract	.61					
3.1	Introduction	.62					
3.2	Materials and methods	.63					
3.2	2.1 Sample collection, experimental layout and sample preparation	.63					
3.2	2.2 Quality checks and transcriptome assembly	.65					
3.2	2.3 Annotation and filtering of unigenes	.66					
3.2	2.4 Evaluation of the transcriptome assembly	.67					
3.2	2.5 Functional classification and detection of genetic elements	.67					
3.2	2.6 Enrichment analysis and in silico profiling of light responsive orthologs	.68					
3.3	Results	.68					
3.3	3.1 Transcriptome annotation and assessment	.68					
3.3	3.2 Functional classification and enrichment analysis	.70					
3.3	3.3 Metabolic pathway enrichment across light irradiances	.72					
	•						

3.	.3.4	Antennae complex genes involved in light capture and response	73
3.	.3.5	Identification of transcriptional regulators and microsatellites	73
3.4.	Dis	cussion	75
3.5	Cor	nclusion	80
3.6	Acl	knowledgements	81
3.7	Ref	erences	82
СЦ	л рт		00
	AFI.		00 09
AUS	Intr	aduction	09 00
4.1	Ma	tarials and matheds	90 01
4.2 1	1VIA	Field analysis and sample transplantation	91 01
4.	.2.1	A querium set up and acclimation period	91 01
4.	.2.2 7 3	Aqualium set-up and acclimation period	91 04
4.	.2.3 7 1	L as f tissue sampling and processing	94 04
4.	.2.4 25	Pead processing, transcriptome assembly and appotation	94 05
4.	2.5	Differential gape expression analysis and gape entelogy enrichment	95 06
4.	2.0	Statistical testing of photobiology and pigmont data	90 07
4.	.2.1 Doc	statistical testing of photobiology and pignicit data	رو 07
4.5	2 1	Dhatabialagy	رو 07
4.	27	Pigmant shifts absorved through UPL C profiling	<i>ا</i> و ۵۵
4.	.J.Z 2 2	Transprintome assembly	100 1
4.		Differential expression analysis	100 100
4.	3.4	Pagulation of genes involved in photosynthesis and carbon fixation	100 103
4.	3.5	Regulation of genes associated with pigment pathways	105 104
4.	3.0	Regulation of genes involved in light perception and signaling	10 <del>4</del> 106
4.	3.7	Gene Optology (GO) enrichment analysis	100 106
4.	.J.0 3 8 1	Dene Ontology (OO) enforment analysis	100 106
4.	3.0.1	Light and pigment enriched GO terms	100 107
4.	3.0.2	Secondary metabolism and ROS enriched GO terms	107 107
4.		cussion	107
4.4 15		cussion	100 116
4.5		znowledgements	110 118
4.0 17	Pof	anow reagements	110 110
4.7	KU		117
CH	APT	ER 5	126
Abs	tract		127
5.1	Intr	oduction	128
5.2	Ma	terials and methods	129
5.	.2.1	Sample transplantation and experimental set-up	129
5.	.2.2	Rapid light curve assessment and photobiology	130
5.	.2.3	RNA sampling, extraction and quality checks	132
5.	.2.4	Prediction of chloroplast genes and primer design	132
5.	.2.5	Reverse transcription and qPCR protocol	133
5.	.2.6	RT-qPCR data analysis and normalisation	134
5.3	Res	sults	134
5	.3.1	Photobiology	134
5	.3.2	Selection of reference genes	136
5.	.3.3	Target gene expression profiling to nuclear-encoded reference genes	138
5.4	Dis	cussion	139
5.5	Cor	nclusion	144

5.6	Acknowledgements	.145
5.7	References	.146
SYN	THESIS, OUTLOOK AND CONCLUSIONS	.152
6.1	Overview	.153
6.2	Photo-physiological acclimation occurs early in Z. muelleri	.154
6.3	Both nuclear and chloroplast-encoded genes play important roles in	
	Z. muelleri, in response to light limitation	.154
6.4	Trends observed in Z. muelleri over 14 days in response to light limitation	.155
6.5	Genome-guided assembly versus de novo transcriptome assembly; Research	
	challenges and implications for the future	.157
6.6	Future research direction from this thesis	.160
6.0	6.1 The seagrass holobiont paradigm and consequences for <i>Z. muelleri</i>	
	under light limited environments	.160
6.0	6.2 Further investigation of light response and acclimation in Z. muelleri	.161
6.0	6.3 Design and implementation of molecular markers for reactive	
	monitoring of light-related stress through targeted gene expression assay	.161
6.7	Conclusions	.162
6.8	References	.164
APP	ENDIX 1	.167
APP	PENDIX 2	.190
APP	PENDIX 3	.217
APP	PENDIX 4	.219

#### **Thesis summary**

Understanding how a keystone marine species responds to its extrinsic environment is important to ensure adequate conservation measures are in place, especially with increasing reports of climate change and anthropogenic disturbance events. For the Southern Hemisphere seagrass, Zostera muelleri, this scenario is no different. This keystone species is native to Australia and New Zealand, providing many socio-economic benefits to the coastal zone. Over the past few decades, a reduction in water quality (light limitation) has led to numerous reports of Z. muelleri meadow loss in Australia and New Zealand. Although seagrass biologists have a firm understanding of the physiological, morphological and ecological changes within light limited Z. muelleri meadows, no current knowledge exists on how Z. muelleri responds to light limitation at the transcriptional level. By investigating transcriptional regulation, new knowledge was obtained on how this species responds to light limitation, allowing for more appropriate conservation measures. Encompassing the advances in RNA-Seq, this project has examined how Z. muelleri responds to light limitation over a 14-day period, through transcriptional regulation, photobiology and physiology, both at the nuclear and chloroplastic level. Main findings indicate that important regulational shifts occur in genes associated with photosynthesis, photo-pigments, carbon metabolism, reactive oxygen species (ROS) homeostasis and secondary defence metabolism. Both nuclear and chloroplast encoded genes involved in photosynthetic processes have been shown to be correlated with downstream changes in photophysiology, and thus are both crucial for the response as well as the acclimation to light limitation. This research also compared genome-guided transcriptome assembly versus de novo assembly, indicating the superiority of genome-guided protocols when a genome is available. Whilst this PhD thesis offers a new level of knowledge to seagrass biologists, it also provides candidate molecular markers, which can be used in future monitoring efforts and population genetic studies.

#### PhD thesis aims and objectives

The overall aim of this thesis is to investigate how *Zostera muelleri* responds to light limitation using a multi-disciplinary approach. By combining mRNA-seq and RTqPCR protocols with already established photobiology and pigment profiling techniques, we will not only obtain a new level of understanding on how this seagrass species responds at the transcriptional level, but also how transcriptional regulation is linked to downstream changes in photophysiology. Such work is timely, given that seagrasses are increasingly threatened by light limitation within the coastal environment.

#### Objectives

- To examine and provide a critical literature review on the current state of molecular profiling and omics techniques in seagrass biology, whilst identifying key knowledge gaps in previous and current research.
- To address the background knowledge associated with light perception in higher plants, seagrasses and *Zostera muelleri*: fundamental knowledge and further direction for research will be discussed.
- To complete *in silico* characterisation and data mining of the *Z. muelleri de novo* transcriptome, based on whole plant tissue.
- To characterise leaf tissue-specific responses of *Z. muelleri* to light limitation; to establish links between transcriptional regulation of nuclear-encoded genes and downstream photophysiology.
- To investigate the expression of chloroplast-encoded photosynthetic genes in *Z*. *muelleri* in response to light limitation. To designate suitable reference genes and link chloroplastic-encoded gene expression with downstream photobiology.