

# Systemic Administration of a Connexin43 Mimetic Peptide: A Treatment Option for Spinal Cord Injury

A thesis submitted in fulfilment of the requirements for the degree of Doctor of Philosophy

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#### **Certificate of Original Authorship**

I certify that the work in this thesis has not previously been submitted for a degree nor has it been submitted as part of requirements for a degree except as part of the collaborative doctoral degree and/or fully acknowledged within the text.

I also certify that the thesis has been written by me. Any help that I have received in my research work and the preparation of the thesis itself has been acknowledged. In addition, I certify that all information sources and literature used are indicated in the thesis.

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#### **Table of Contents**

Certificate of Original Authorshipi		
Acknowledgementsii		
Table of Contents iv		
List of Figures viii		
List of Tablesx		
Abbreviation xi		
Publications Arising from PhD Researchxiii		
Other Publications during PhD Candidaturexiv		
Conference Proceedings Arising from PhD Researchxv		
Awards and Scholarshipsxvi		
Abstractxvii		
1. Chapter 1 General Introduction1		
1.1 Epidemiology of spinal cord injury2		
1.2 Pathophysiology of spinal cord injury6		
1.2.1 Primary injury		
1.2.2 Secondary injury		
1.2.3 Astroglial scar formation9		
1.2.4 Inflammatory cells 12		
1.2.4.1 Activation of resident microglia13		
1.2.4.2 Penetration of circulating neutrophils14		
1.2.4.3 Recruitment of blood-borne monocytes15		
1.2.4.4 Infiltration of lymphocytes16		
1.2.5 Neurotoxic molecules		
1.2.6 Gap junctional communication19		
1.3 Present clinical approaches for spinal cord injury23		
1.3.1 Methylprednisolone to limit secondary injury 23		
1.3.2 Surgical stabilisation and decompression25		
1.3.3 Multisystem medical management		

	1.3.4 Functional rehabilitation	27
	1.4 Current research on Connexin43 for spinal cord injury repair	28
	1.4.1 Genetic deletion of Connexin43	29
	1.4.2 Gap junction inhibitors	30
	1.4.3 Connexin43 antisense oligodeoxynucleotides	31
	1.4.4 Connexin43 mimetic peptides	34
	1.5 Project overview	37
	1.5.1 Hypotheses and aims	38
2	. Chapter 2 Optimisation of the peptide5 Systemic Administration Protocol for	•
Т	raumatic Spinal Cord Injury	40
	2.1 Introduction	41
	2.2 Materials and methods	43
	2.2.1 Preparation of peptides	43
	2.2.2 Mass spectrometry	44
	2.2.3 Electrophoresis	45
	2.2.4 Animals and general care	46
	2.2.5 Surgical procedures	47
	2.2.6 Peptide treatment	48
	2.2.7 Post-operative care	49
	2.2.8 Behavioural assessments	49
	2.2.9 Euthanasia	51
	2.2.10 Western blot	51
	2.2.11 Histology	52
	2.2.12 Fluorescent immunohistochemistry	53
	2.2.13 Imaging and quantification	54
	2.2.14 Statistical analysis	55
	2.3 Results	56
	2.3.1 Degradation curves of Peptide5 <i>in vitro</i>	56
	2.3.1.1 Mass spectrometry	56
	2.3.1.2 Electrophoresis	
	2.3.2 Consistent spinal cord injury in rats	
	2.3.3 Dosing responses of Peptide5	
	2.3.4 Systemic delivery of Peptide5 to the lesion	
	2.4 Discussion	67

3. Chapter 3 Investigation of Cellular and Functional Improvement in Spinal Cord			
Injury Rats from the Peptide5 Systemic Administration	72		
3.1 Introduction	73		
3.2 Materials and methods	75		
3.2.1 Animals and general care	75		
3.2.2 Surgical procedures	75		
3.2.3 Peptide treatment			
3.2.4 Post-operative care			
3.2.5 Behavioural assessments			
3.2.5.1 Locomotor function	76		
3.2.5.2 Pain hypersensitivity	77		
3.2.6 Euthanasia and histology			
3.2.7 Fluorescent immunohistochemistry			
3.2.8 Imaging and quantification	81		
3.2.9 Statistical analysis			
3.3 Results	83		
3.3.1 Consistent spinal cord injury in rats	83		
3.3.2 Connexin43 and phosphorylated Connexin43	84		
3.3.3 Lesion size	87		
3.3.4 Astrogliosis (GFAP immunohistochemistry)	89		
3.3.5 Microglial/macrophage activation (Iba1&ED1 immunohistochemistry)			
3.3.6 Neuronal survival (NeuN immunohistochemistry)			
3.3.7 Motor function (Open field and horizontal ladder tests)			
3.3.8 Sensory function (Mechanical allodynia)			
3.4 Discussion	99		
4. Chapter 4 Safety Profile of the Peptide5 Systemic Administration for	Traumatic		
Spinal Cord Injury			
4.1 Introduction			
4.2 Materials and methods			
4.2.1 Distribution of Peptide5 <i>in vivo</i>			
4.2.1.1 Animals and general care			
4.2.1.2 Surgical procedures			
4.2.1.3 Euthanasia and tissue preparation			
4.2.2 Connexin43 protein in the heart and lung tissue			

	4.2.2.1 Histology on the heart and lung tissue	111	
	4.2.1.2 Immunohistochemistry on the heart and lung tissue		
	4.2.3 Major connexin proteins in the spinal cord tissue	113	
	4.2.3.1 Cellular distribution of major connexins in the spinal cord tissue		
	4.2.3.1 Quantification of major connexins in the spinal cord tissue	115	
	4.2.4 Imaging and quantification	116	
	4.2.5 Inflammatory effects in plasma	116	
	4.2.6 Statistical analysis	117	
4	I.3 Results	118	
	4.3.1 Distribution of Peptide5 <i>in vivo</i>	118	
	4.3.2 Connexin43 protein in the heart tissue	120	
	4.3.3 Connexin43 protein in the lung tissue	122	
	4.3.4 Major connexin proteins in the spinal cord tissue	123	
	4.3.5 Inflammatory effects in plasma	128	
4	I.4 Discussion	129	
5.	Chapter 5 General Discussion	137	
5	i.1 Project summary	138	
5	i.2 Future directions	141	
5	i.3 Conclusion	144	
Rih	liography	145	
	но <sup>р</sup> . арту		
Арр	Appendix – Reagent Specifications 181		

## **List of Figures**

Figure 1.1 The severity of SCI measured by neurological level of injury and extent of
injury4
Figure 1.2 The formation of astroglial scar following spinal cord injury in humans 11
Figure 1.3 Temporal response of inflammatory cells in spinal cord injury rats
Figure 1.4 Illustration of gap junction (A) and connexin (B) structures in vertebrates. 20
Figure 2.1 Schematic diagram of the Multicentre Animal Spinal Cord Injury Study
impactor
Figure 2.2 Mass spectrometry analysis of Peptide5 degradation in normal rat serum in
<i>vitro</i>
Figure 2.3 Electrophoresis analysis of Peptide5 degradation in normal rat and human
sera <i>in vitro</i>
Figure 2.4 Surgical parameters for Cohort I showing the means of (A) weight-drop
velocity in m/s, (B) compression in mm and (C) compression rate in m/s with standard
error of the mean
Figure 2.5 Behavioural assessment of hind limb locomotor function for Cohort I 60
Figure 2.6 Surgical parameters for Cohort II showing the means of (A) weight-drop
velocity in m/s, (B) compression in mm and (C) compression rate in m/s with standard
error of the mean
Figure 2.7 Western blot analysis of Connexin43 and phosphorylated Connexin43
protein levels at the injury site 24 hours following surgery
Figure 2.8 Changes in lesion size at 8 hours following spinal cord contusion injury 63
Figure 2.9 Immunohistochemistry staining of Connexin43 at 8 hours following spinal
cord injury
Figure 2.10 Immunohistochemistry staining of phosphorylated Connexin43 at 8 hours
following spinal cord injury
Figure 3.1 Immunohistochemistry staining of Connexin43 at 24 hours following spinal
cord injury
Figure 3.2 Immunohistochemistry staining of phosphorylated Connexin43 at 24 hours
following spinal cord injury
Figure 3.3 Changes in lesion size following spinal cord contusion injury

Figure 3.4 Glial fibrillary acidic protein (GFAP) expression following spinal cord
contusion injury
Figure 3.5 Activation of microglia and/or macrophages at the lesion edge following
spinal cord contusion injury
Figure 3.6 Surviving neuronal cells following spinal cord contusion injury
Figure 3.7 Locomotor function assessments following spinal cord injury
Figure 3.8 Sensory function assessments following spinal cord injury
Figure 4.1 Distribution of fluorescein isothiocyanate labelled Peptide5 in normal and
spinal cord injury rats119
Figure 4.2 Histological morphology and the immunohistochemistry staining of
Connexin43 and phosphorylated Connexin43 in the heart tissue
Figure 4.3 Histological morphology and the immunohistochemistry staining of
Connexin43 in the lung tissue
Figure 4.4 Immunohistochemistry co-labelling of major connexin proteins and neural
markers at 8 hours following spinal cord injury
Figure 4.5 Immunohistochemistry staining of Connexin30 in rat spinal cord at 8 and 24
hours following injury
Figure 4.6 Immunohistochemistry staining of Connexin36 in rat spinal cord at 8 and 24
hours following injury
Figure 4.7 Immunohistochemistry staining of Connexin47 in rat spinal cord at 8 and 24
hours following injury

## **List of Tables**

Table 1.1 Medical management for people with spinal cord injury (Becker et al., 2003).
Table 1.2 Summary of clinical trials for Nexagon®
Table 1.3 Summary of animal cohorts and tissue samples obtained in the current study
Table 2.1 Basso, Beattie, and Bresnahan locomotor rating scale (Basso et al., 1996)50
Table 2.2 Summary of filter characteristics for fluorescent imaging. 54
Table 3.1 Summary of antibody characteristics for immunohistochemistry
Table 3.2 Consistent contusion spinal cord injury produced in rats. 83
Table 4.1 Summary of the heart and lung samples used for the Connexin43 study111
Table 4.2 The routine overnight processing programme for histology
Table 4.3 Summary of antibody characteristics for immunohistochemistry co-labelling
of connexins and neural markers
Table 4.4 Summary of the spinal cord samples used for quantification of major
connexins
Table 4.5 Summary of antibody characteristics for immunohistochemistry of major
connexins
Table 4.6 Comparison of cytokines, chemokines and growth factors in rat plasma 128

#### Abbreviation

3Rs:	Replacement, Reduction and Refinement
ACEC:	Animal Care and Ethics Committee
AMPA:	a-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid
ANOVA:	Analysis of Variance
AS-ODN:	Antisense oligodeoxynucleotide
ATP:	Adenosine triphosphate
BBB:	Basso, Beattie and Bresnahan
BSB:	Blood spinal cord barrier
CNS:	Central nervous system
FITC:	Fluorescein isothiocyanate
FL:	Front limb
GABA:	γ-aminobutyric acid
GFAP:	Glial fibrillary acidic protein
H&E:	Haematoxylin and Eosin
HL:	Hind limb
HREC:	Human Research Ethics Committee
IHC:	Immunohistochemistry
MASCIS:	Multicentre Animal Spinal Cord Injury Study
MP:	Methylprednisolone
mRNA:	Messenger ribonucleic acid
MS:	Mass spectrometry
NASCIS:	National Acute Spinal Cord Injury Studies
NeuN:	Neuronal nuclei
NGS:	Normal goat serum
NMDA:	N-methyl-D-aspartate
OSP:	Oligodendrocyte Specific Protein
P5:	Peptide5
PBG:	Phosphate buffer with 5% normal goat serum
PFA:	Paraformaldehyde
ROS:	Reactive oxygen species
SCI:	Spinal cord injury

SDS:	Sodium dodecyl sulphate
SEM:	Standard error of the mean
SP:	Scrambled peptide
TBS:	Tris-buffered saline
UNSW:	University of New South Wales
UTS:	University of Technology Sydney

#### **Publications Arising from PhD Research**

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TONKIN, R. S., **MAO, Y.,** O'CARROLL, S. J., NICHOLSON, L. F., GREEN, C. R., GORRIE, C. A. & MOALEM-TAYLOR, G. 2014. Gap junction proteins and their role in spinal cord injury. *Frontiers in Molecular Neuroscience* 7.

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CHEN, H., Chan, Y.L., NGUYEN, L.T., **MAO, Y.,** de ROSA, A., Beh, I.T., CHEE, C., OLIVER, B., HEROK, G., SAAD, S. & GORRIE, C., 2016. Moderate traumatic brain injury is linked to acute behaviour deficits and long term mitochondrial alterations. *Clinical and Experimental Pharmacology and Physiology* 43, 1107-1114.

**MAO, Y.,** NGUYEN, T., SUTHERLAND, T. & GORRIE, C. A. 2016. Endogenous neural progenitor cells in the repair of the injured spinal cord. *Neural Regeneration Research*, 11, 1075-1076.

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**MAO, Y.,** TONKIN, R. S., NGUYEN, T., O'CARROLL, S. J., NICHOLSON, L. F., GREEN, C. R., MOALEM-TAYLOR, G. & GORRIE, C. A., 2016. Systemic administration of a Connexin43 mimetic peptide in rats improves functional recovery after spinal cord injury. *Australasian Neuroscience Society Annual Conference*, Hobart, TAS: Australasian Neuroscience Society.

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**MAO, Y.,** TONKIN, R. S., NGUYEN, T., O'CARROLL, S. J., NICHOLSON, L. F., GREEN, C. R., MOALEM-TAYLOR, G. & GORRIE, C. A., 2016. Systemic delivery of a Connexin43 hemichannel blocking mimetic peptide improves functional recovery following spinal cord injury in rats. *Neuroscience 2016*, San Diego, California: Society for Neuroscience.

**MAO, Y.,** TONKIN, R. S., NGUYEN, T., O'CARROLL, S. J., NICHOLSON, L. F., GREEN, C. R., MOALEM-TAYLOR, G. & GORRIE, C. A., 2015. Systemic delivery of a Connexin43 mimetic peptide in rats improves hindlimb locomotor function following spinal cord injury. *New Horizons Combined Health Science Conference*, Sydney, NSW: NSW Health Government.

MAO, Y., TONKIN, R. S., NGUYEN, T., O'CARROLL, S. J., NICHOLSON, L. F., GREEN, C. R., MOALEM-TAYLOR, G. & GORRIE, C. A., 2015. Systemic delivery of a connexin43 hemichannel blocking mimetic peptide in rats improves hindlimb locomotor function following spinal cord injury. *Inter-university Neuroscience and Mental Health Conference*. Sydney, NSW: Brain Sciences UNSW.

**MAO, Y.,** TONKIN, R. S., NGUYEN, T., O'CARROLL, S. J., NICHOLSON, L. F., GREEN, C. R., MOALEM-TAYLOR, G. & GORRIE, C. A., 2015. Systemic delivery of a mimetic peptide against connexin43 gap junction protein in rats following spinal cord injury. *Australasian Neuroscience Society Annual Conference*. Cairns, QLD: Australasian Neuroscience Society.

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#### Abstract

Spinal cord injury (SCI) is a major cause of morbidity, and leads to significant physical, emotional and financial burdens for individuals and their families. There is no known cure to date and current treatment options are limited. A cascade of secondary tissue damage starts within minutes to hours of an initial traumatic SCI, worsening the cellular and functional recovery. The time course of secondary damage does, however, offer a potential window of opportunity for intervention. It may be possible to administer new therapeutics during this early period, to prevent the extensive spread of damage to adjacent healthy tissue.

Over the past two decades, regulation of Connexin43 hemichannels has provided a novel avenue of research into limiting the secondary damage of SCI. Peptide5, a Connexin43 mimetic peptide, is a promising candidate that shows efficacy in blocking Connexin43 hemichannels, thus preventing the propagation of neurotoxic molecules from the injured cells to the nearby intact tissue in a number of neural injury models. Peptide5, in particular, has been shown to reduce secondary tissue damage and to improve functional recovery when delivered <u>directly</u> to the site of SCI. However, direct application of peptides to a spinal cord lesion is likely to be impractical in a clinical setting, and might result in delays to the initiation of treatment. Therefore, the current research project aims to demonstrate that the <u>systemic</u> administration of Peptide5 is a clinically relevant treatment option for traumatic SCI by (1) optimising the systemic administration protocol of Peptide5; (2) investigating the cellular and function improvements in SCI rats from the Peptide5 systemic administration; and (3) examining the safety profile of Peptide5 systemic administration protocol.

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Mass spectrometry and electrophoresis were used to determine the *in vitro* degradation curves in normal rat and human sera. A rat model of contusive SCI was then employed to optimise the dosing regimen and investigate the efficacy of Peptide5 in limiting secondary tissue damage and improving functional recovery. The lesion size was assessed on Haematoxylin and Eosin stained histological sections, and fluorescent immunohistochemistry was utilised to examine the Connexin43 protein and hemichannels, as well as cellular responses. The behavioural improvements were evaluated from motor (open field and error ladder tests) and sensory (mechanical pain hypersensitivity) functions. Fluorescent-labelled Peptide5 was delivered to the SCI and normal rats systemically in order to determine the *in vivo* distribution.

It was found that the *in vitro* half-life of Peptide5 was 2 hours in normal rat serum and that the most effective dosing regimen was three injections of Peptide5 at 0 (10 mg/kg), 2 (5 mg/kg) and 4 (2.5 mg/kg) hours post-injury. Compared with controls, Peptide5 treated rats showed significant improvements in hind limb locomotor function and mechanical pain hypersensitivity. Peptide5 treatment led to a reduction in Connexin43 protein level, hemichannel opening, lesion size, astrogliosis, macrophage and/or microglial activation, and neuronal cell death. There was no evidence to suggest that Peptide5 had any adverse systemic effects in normal and SCI animals, or influenced other connexins in the spinal tissue.

The current research project demonstrated that systemic administration of Peptide5 is a feasible, neuroprotective and safe treatment option for ameliorating secondary damage and improving functional recovery in a contusive SCI model of rats. These findings provide strong support for the clinical use of Peptide5 in the near future.

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