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3	4	ROOT XYLEM CHARACTERISTICS AND HYDRAULIC STRATEGIES OF SPECIES CO-
4	5	OCCURRING IN SEMI-ARID AUSTRALIA
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ABSTRACT

Xylem traits such as xylem vessel size can influence the efficiency and safety of water transport and thus plant growth and survival. Root xylem traits are much less frequently examined than those of branches despite such studies being critical to our understanding of plant hydraulics. In this study, we investigated primary lateral and sinker roots of six co-occurring species of semi-arid Australia. Two species are restricted to a floodplain, two were sampled only from the adjacent sand plain, and two species co-occur in both habitats. We assessed root wood density, xylem traits (i.e., vessel diameter, fibre and vessel wall thickness), outer pit aperture diameter and calculated theoretical hydraulic conductivity and vessel implosion resistance. We hypothesized that (1) roots have larger xylem vessel diameters and lower wood density than branches of the same species and that (2) there is an inverse correlation between theoretical sapwood hydraulic conductivity and vessel implosion resistance for roots.

Variation in root wood density was explained by variations in xylem vessel lumen area across the different species ($r^2=0.73$, p=0.03), as hypothesized. We rejected our second hypothesis, finding instead that the relationship between theoretical hydraulic conductivity and vessel implosion resistance was not maintained in roots of all of our studied species, in contrast to our previous study of branches from the same species. Xylem traits were found to depend upon habitat and eco-hydrological niche, with the groupings including (i) arid-adapted shrubs and trees with shallow lateral roots (Acacia aneura and Psydrax latifolia), (ii) trees restricted to the floodplain habitat, both evergreen (Eucalyptus camaldulensis) and deciduous (Erythrina vespertilio) and (iii) evergreen trees co-occurring in both floodplain and adjacent sand plain habitats (Corymbia opaca and Hakea sp.).

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INTRODUCTION

Xylem traits play an essential role in the efficiency and safety of water transport through the plant, influencing plant growth and survival across a range of environments (Smith & Sperry 2014). Xylem vessels of large diameter (e.g., > 500 µm) can transport water more efficiently and with lower resistance to flow than small xylem vessels (e.g., $<20 \mu m$) as a result of the fourth-power relationship between radius and flow through a capillary tube (i.e. Hagen-Poiseuille law; Tyree & Zimmerman 2002). However, the architecture of the pits between large xylem vessels can also increase the risk of xylem cavitation events (e.g. Hargrave et al. 1994; Lo Gullo et al. 1995; Cai et al. 2010; Christman et al. 2012). Pits are the structures that interconnect vessels and can influence the efficiency and safety of plant water transport, determining as much as 50% of the xylem hydraulic resistance in species with small pit apertures (Sperry *et al.* 2005; Wheeler et al. 2005; Hacke et al. 2006). Recent studies also suggest that bubbles known as nanobubbles can move through pit membranes due to their smaller size than that of pit 28 104 membrane pores (2 to 200 nm) (Schenk et al. 2015, Schenk et al. 2016). Nanobubbles, different from larger bubbles, are safe from expanding and can dissolve or remain in the pit border (a confined space) until temperatures decrease (at night), thereby increasing gas solubility. Additionally, once formed, nano-bubbles can be stabilized by amphiphilic surfactants present in the xylem sap which can transform surfaces that are hydrophobic into hydrophilic ones (Schenk et al. 2015).

Xylem traits of roots are much less frequently studied than those of branches or stems, in part due to difficulty in accessing this plant organ. However, roots are the primary path for water uptake by plants; therefore, knowledge of xylem structure in roots can provide critical information on overall plant water supply (Canadell et al. 1996; Martínez-Vilalta et al. 2002; 46 114 McElrone et al. 2004; Johnson et al. 2014) and are likely to define key hydraulic traits of the entire plant. Previous studies have found that at a species level, root xylem has a larger risk of cavitation (but higher efficiency of water transport) than branch, stem and leaf xylem because of larger vessel diameters in roots (Sperry & Saliendra 1994; Martínez-Vilalta et al. 2002; 53 118 Anfodillo et al. 2006).

Our previous work with species from the Ti Tree Basin arid zone, in central Australia (Santini et al. 2016) indicates that branch xylem traits can contribute to co-occurrence within close 59 121 proximity of species in this arid habitat, where water resources are heterogeneously distributed

in space and time (O'Grady et al. 2009). For example, species with shallow lateral roots such as Acacia spp. that rely on intermittent water resources such as rainwater (Cleverly et al. 2016a) had smaller xylem vessels in their stems than that of phreatophytic species with deep tap or sinker roots and thus access to groundwater (e.g. *Eucalyptus* and *Corymbia*). We also found in stem xylem that there was a significant linear regression between lower theoretical hydraulic conductivities (K_s, kg mm⁻¹ MPa⁻¹ s⁻¹) and higher xylem vessel implosion resistance ($[t/b]^2$), the ratio of vessel double-wall thickness and vessel lumen diameter), concluding that xylem vessels of species with higher theoretical hydraulic conductivities (i.e. *Erythrina vespertilio, Corymbia* opaca and Eucalyptus camaldulensis) were less protected from implosion than xylem vessels of species like Acacia spp., Hakea sp. and Psydrax latifolia which have lower theoretical hydraulic conductivities (Santini *et al.* 2016). Vessel implosion resistance may be an indicative of vessel wall reinforcement, but in roots it might not be indicative of safety as tensile forces in the water column are low (Hacke et al. 2001; Pratt et al. 2007). We retain the nomenclature of "vessel implosion resistance" merely because it has a history within the literature, whilst recognizing that it may not be a relevant index to assess root xylem vessel safety (Pratt et al. 2007).

In the present work, we studied six co-occurring species in close proximity within the Ti Tree Basin. These species can be classified into three functional groups with respect to their leaf longevities and root deployment strategies: 1) deciduous trees (*Erythrina vespertilio*), 2) evergreen shrubs and trees (Acacia aneura, Hakea sp. and Psydrax latifolia) and 3) evergreen phreatophytes (Eucalyptus camaldulensis and Corymbia opaca). There are ca. 150 species across Australia in the genus *Hakea*. Although a positive identification could not be made in this case, Hakea sp. in this study is likely to be Hakea arborescens or Hakea macrocarpa. In general, drought-deciduous trees avoid transpiration during the dry season by dropping their leaves for at least one and as many as seven months of the year. Alternatively, evergreen trees retain *ca*. 90 % of their full canopy throughout the year (Eamus 1999). Psydrax latifolia forms an association with Acacia aneura, which has a relatively shallow root system and does not access 46 149 the groundwater of the site (Cleverly et al. 2016a; Cleverly et al. 2016b). Phreatophytes like those in the Ti Tree basin do access groundwater by having a taproot and deep sinker roots, thereby allowing retention of their canopy through the dry season (Dawson & Pate 1996; Roberts et al. 2016).

We measured wood density, xylem traits (i.e., vessel diameter, fibre and vessel wall thickness)
 and outer pit aperture diameter, and then we calculated theoretical hydraulic conductivity and
 vessel implosion resistance in a primary sinker root (in the evergreen phreatophytes) or
 shallow lateral roots (in the remainder of the species). Vessel implosion resistance has only

been observed in abnormal wood, but it is a measure of vessel wall reinforcement (Baas & Wheeler 2015). We hypothesized that: (1) roots have larger xylem vessel diameters and lower wood density than branches of the same species, placing roots at more risk of cavitation events than stems and (2) there is an inverse correlation between theoretical sapwood hydraulic conductivity and vessel implosion resistance for roots, similar to what we have observed in branches (Santini et al. 2016).

MATERIALS AND METHODS

Site description and sample collection

Our study site is located in central Australia in the Ti Tree Basin (22° 7' 48.56" S, 133° 24' 57. 67" E), approximately 180 km north of Alice Springs, Northern Territory, Australia. The Ti Tree Basin is an arid catchment that covers *ca*. 5500 km² and contains the ephemeral Woodforde River and Allungra Creek. Mean annual rainfall at the nearest meteorological station is 320 mm year⁻¹ (Australian Bureau of Meteorology 2014, BoM station 15643, Territory Grape Farm), and most of this rainfall occurs in monsoonal summer (Dec - Feb) storms (Cleverly et al. 2016b). The mean minimum and maximum daily temperature is 15 and 31 °C, respectively, while the maximum air temperature at the site reaches 46 °C in the summer months (December to February) (Harrington et al. 2002).

Woody plants were sampled along a 6.5-km transect from the banks of the Woodforde River and its floodplain (*Erythrina vespertilio n=5* and *Eucalyptus camaldulensis n=5*) into the adjacent sand plain (Acacia aneura n=5, and Psydrax latifolia n=5), and two tree species were sampled 40 178 from both habitats (*Corymbia opaca* n=3 and *Hakea* sp. n=5). All of these woody species can be found to co-occur within tens of metres of each other in this semi-arid climate, but they inhabit different eco-hydrological niches (Nolan et al. In review). Two trees are groundwater dependent in the Ti Tree Basin: one is restricted to riparian corridors and paleochannels (Eucalyptus camaldulensis); and the other (Corymbia opaca) is widespread across the basin (O'Grady et al. 2009). Acacia aneura is also found in both floodplain and sand plain habitats of the Ti Tree 52 185 Basin (although not on this transect), but these arid-adapted trees are found on siliceous hardpan which provides a near-surface reservoir of soil water storage within their shallow root zone (Cleverly et al. 2016a). Plants grow as trees, except for Psydrax latifolia, which grows as a shrub (up to 3 m tall).

Root material was collected from three to five roots in each of six woody species (one root per

190 individual plant) of the Ti Tree Basin. Samples were obtained during the austral winter and dry 191 season, June – August 2015 (Figure 1, 2, Table 1). For each species, trees and shrubs were 192 chosen which had roots of a given diameter before sinking below a practical collection depth in 193 the hardpan soil. Roots were sought in a 360° circle around the main trunk and were collected 194 from whichever direction they were found. We collected roots by digging from the main stem to 195 a point where the target primary lateral or sinker root had a diameter of *ca.* 15 mm, at a 196 distance of *ca.* 0.5–6 m from the main trunk (Table 1).

7 Wood density

Root material collected for assessment of wood density was stored in paper bags and kept cool
for laboratory analysis. Wood density of roots was measured in 4 cm lengths of sapwood.
Sample numbers of individual roots (i.e. one root sample per plant) for assessment of wood
density were as follows: *Erythrina vespertilio* (n=5), *Acacia aneura* (n=5), *Hakea* sp. (n=5), *Psydrax latifolia* (n=5), *Corymbia opaca* (n=3) and *Eucalyptus camaldulensis* (n=5). Root
sapwood segments were weighed after drying to a constant mass. Wood density was calculated
as the ratio of dry weight (5 days at 60 °C) to volume of water displaced by the fully hydrated
sapwood segments (i.e., after they were submerged in water for two days; see Santini *et al.*2016).

Assessment of xylem traits

Preparation of tissues and measurements of xylem traits were performed following Santini et al. (2016). Root samples were cut into segments of two centimetres in length for analysis of cross sections. Each root segment was fixed in formalin-acetic acid-alcohol (FAA) for ~10 days and then placed in 70% ethanol for 2 days. After this washing process, samples were dehydrated for one day each in increasing concentrations of ethanol: 70%, 96% and 100%. From the ~15 mm-diameter samples, we prepared 10–20 µm-thick cross sections using a sledge Leica SM2010R microtome. Number of samples (each representing one individual tree or shrub) for measuring xylem traits were as follows: Erythrina vespertilio n=5, Acacia aneura n=3, Hakea sp. n=4, Psydrax latifolia n=5, Corymbia opaca n=3 and Eucalyptus camaldulensis n=5. Next, Erythrina vespertilio, Acacia aneura, Hakea sp. and Psydrax latifolia were double-stained with safranin and alcian blue (1:2). Corymbia opaca and Eucalyptus camaldulensis were assessed with no-staining due to the presence of resins that complicated the assessment of wood xylem traits after staining. Sections were photographed for measurements using a Leica DM750 microscope. Vessel diameters (minor and major axes), fibre wall thickness and vessel wall thickness were measured in ImageJ v1.48 (National Institutes of Health, USA). The cross-sectional vessel lumen

area (VA) was determined as an ellipse (i.e. as a product of π and the semi-minor and semimajor radii), which was then multiplied by vessel density (VD, number of vessels per unit crosssectional area) to determine conductive lumen fraction (F, named CA in Santini *et al.* 2016). We determined fibre wall thickness in 30 - 50 adjoining fibres (Santini *et al.* 2013). Additionally, xylem vessel wall thickness was measured at three positions (i.e. pie-shaped sectors of 20 - 50 vessels) in the cross section (Santini *et al.* 2016).

9 Scanning Electron Microscopy (SEM) preparation and imaging

We measured the diameter of pit apertures in three individual roots per species (one sample per individual plant) through the analysis of SEM imagery. Samples of root tissue (which corresponded to similar positions in which we assessed the xylem traits) were hand-sectioned longitudinally to a thickness of 2 - 4 mm. The sections were treated with a series of ethanol and hexamethydisilizane as detailed in Johnson *et al.* (2014). After treatment, samples were placed into the stage of an EVO LS15 SEM (Zeiss, Germany) and were visualized in variable pressure mode. We used an accelerated voltage of 15 kV and took images with magnifications of 250X, 500X and 2kX for all our samples. Diameters of pit apertures were measured as recommended by Scholz *et al.* (2013) by assessing the widest part of the opening of a minimum of 50 pits per species with the software Image J 1.48v (National Institutes of Health, USA).

Theoretical hydraulic conductivity and resistance to vessel implosion

Theoretical hydraulic conductivity (K_s) was determined as a function of (i) conductive lumen fraction (F, equal to VA·VD) and (ii) the ratio of vessel size to vessel density (S, VA/VD; Zanne *et al.* 2010) (Equation 1).

 $K_{\rm S} \propto F^{1.5} S^{0.5}$

(Equation 1)

Vessel implosion resistance was estimated as the ratio of the double-wall thickness (t, the total
vessel wall thickness between adjacent cells) to the mean vessel diameter (Hacke *et al.* 2001;
Zanne *et al.* 2010). Mean hydraulic diameter was defined as DH=(ΣD⁴/N)^{1/4} and represents the
mean diameter that all of the vessels in a root would have in order to correspond to the same
overall conductivity for the numbers of conduits (Tyree & Zimmerman 2002).

0 Data analyses

We used linear regression analyses to test relationships between univariate pairs: wood density
and vessel lumen area, log (vessel density) and vessel lumen area, outer pit aperture and log
(vessel diameter), theoretical hydraulic conductivity (K_s) and log (xylem vessel implosion

resistance), and vessel wall thickness and vessel diameter. Significant differences among species
in fibre wall thickness, vessel wall thickness, vessel diameter, outer pit aperture, the conductive
lumen fraction and wood density were assessed with a one-way ANOVA. Statistical analyses
were performed with the software Prism version 7.0c (GraphPad Software, La Jolla, CA, USA).

We also analyzed data obtained from the same population of trees or shrubs used in this study (Santini *et al.* 2016). Santini *et al.* (2016) sampled three replicate branches in three individual trees/shrubs per species ($n = 3 \times 3$ per species); in contrast, we sampled one root from each of three to five individuals per species (n = 3 to 5 per species). We tested differences in means (\pm standard errors) between roots and branches in wood density, xylem traits (i.e. vessel diameter, fibre and vessel wall thickness and conductive lumen fraction) and theoretical hydraulic conductivity and vessel implosion resistance by using a t-test with the Holm-Sidak approach. This method assumes that means are sampled from populations with the same standard deviations.

RESULTS

In the root samples, wood density values ranged from 0.21 ± 0.009 g cm⁻³ in *Erythrina vespertilio* to 0.82 ± 0.023 g cm⁻³ in *Hakea* sp. (Table 2). Vessel diameter values were different across species and varied from 24.4 ± 0.29 µm in *Psydrax latifolia* to 188 ± 6.15 µm in *Erythrina vespertilio*. Fibre walls were thinnest in *Acacia aneura* $(3.9 \pm 0.10$ µm) and thickest in *Corymbia opaca, Hakea* sp. and *Eucalyptus camaldulensis* $(5.7 \pm 0.15$ µm, 5.4 ± 0.12 µm, 5.3 ± 0.08 µm, respectively). Vessel walls were thinnest in *Psydrax latifolia* $(4.12 \pm 0.05$ µm) and thickest in *Corymbia opaca* $(11.5 \pm 0.29$ µm). Outer pit apertures were smallest in *Psydrax latifolia* $(3.73 \pm 0.05$ µm) and largest in *Erythrina vespertilio* $(7.13 \pm 0.11$ µm) (Table 2). Conductive lumen fraction ranged from 0.05 ± 0.003 in *Erythrina vespertilio* to 0.37 ± 0.008 in *Psydrax latifolia* (Table 2).

279 Root wood density was negatively correlated with root xylem vessel lumen area (i.e. wood 280 density was smallest in roots with large xylem vessels; $r^2=0.73$, p=0.03, Figure 3). Roots of 50 281 Hakea sp. fell outside of this relationship with a much higher root wood density for its root 5₂ 282 vessel lumen area than in the other five species. We also found strong linear regressions 283 between log (root vessel density) versus root vessel lumen area (r²=0.72, p=0.03, Figure 4) and 55 284 outer pit aperture in root xylem *versus* log (root vessel diameter) (r²=0.92, p=0.002, Figure 5). 57 285 Outer pit apertures and vessel diameters were smallest in *Psydrax latifolia* $(3.73 \pm 0.05 \,\mu\text{m}$ and 286 24.4 \pm 0.29 µm) and largest in *Erythrina vespertilio* (7.13 \pm 0.11 µm and 188 \pm 6.15 µm) (Table

287 2, Figure 5). The two species which were collected from only the sand plain (*Psydrax latifolia*288 and *Acacia aneura*) had the highest vessel density and lowest vessel lumen area, vessel
289 diameter and outer pit aperture (Figures 3 - 5). By contrast, the two riparian trees (*Eucalyptus*290 *camaldulensis* and *Erythrina vespertilio*) had root xylem with the largest vessel lumen area,
291 vessel diameter and theoretical hydraulic conductivity, and the smallest theoretical vessel
292 implosion resistance, among the six-woody species (Figures 3, 5 and 6).

In root xylem, we found a strong effect of habitat (i.e., obligate riparian *versus* widespread or
arid-adapted species) on the relationship between theoretical hydraulic conductivity and log
xylem vessel implosion resistance (Figure 6). Theoretical hydraulic conductivity was smallest,
and theoretical implosion resistance largest, in the group of species which are not restricted to
riparian corridors (*Acacia aneura*, *Psydrax latifolia*, *Corymbia opaca* and *Hakea* sp.). The largest
theoretical hydraulic conductivity was found in *Eucalyptus camaldulensis* and was lowest in *Psydrax latifolia*, whereas vessel implosion resistance followed a gradient across species: *Acacia aneura > Psydrax latifolia > Hakea* sp. > *Corymbia opaca > Eucalyptus camaldulensis > Erythrina*vespertilio (Table 3, Figure 6).

We also found a significant regression between vessel diameter and vessel wall thickness for root xylem in the four species which are not restricted to riparian habitats ($r^2=0.99$, p=0.004, Figure 7). The small number of points in the regression reduces statistical power, but the regression was not significant when *Eucalyptus camaldulensis* and *Erythrina vespertilio* were included in the analysis ($r^2=0.02$, p=0.70). It was furthermore exceptional that these fourspecies fit so tightly to a single regression line, suggesting that those species which are not restricted to riparian habitats have tight co-constraints of root vessel diameter and wall thickness. Clear species pair-groupings by habitat and eco-hydrological niche were found in vessel diameter and wall thickness: arid-adapted Acacia aneura and Psydrax latifolia, obligate riparian Eucalyptus camaldulensis and Erythrina vespertilio, and Corymbia opaca and Hakea sp. which occurred in both habitats (Figure 7).

Wood density was significantly larger in branches than in roots (Table 2). Our t-test 49 314 comparisons of fibre wall thickness between roots and branches indicated that Hakea sp., 51 315 Corymbia opaca and Eucalyptus camaldulensis exhibited thicker fibre walls in roots than in branches. Fibre wall thickness in Acacia aneura was similar for roots and branches, whereas Psydrax latifolia had thinner fibre walls in roots than in branches. Vessel wall thickness was 56 318 larger in roots than in branches of Erythrina vespertilio, Acacia aneura, Hakea sp., Corymbia opaca and Eucalyptus camaldulensis, but was similar in roots and branches of Psydrax latifolia. Vessel diameter was larger in roots than in branches for *Erythrina vespertilio*, *Hakea sp., Psydrax*

latifolia, Corymbia opaca, and Eucalyptus camaldulensis but was similar in roots and branches of Acacia aneura. Conductive lumen fraction in Erythrina vespertilio and Corymbia opaca was smaller in roots than in branches, whereas it was larger in *Psydrax latifolia* roots compared to branches. Conductive lumen fraction was similar in roots and branches of Acacia aneura, Hakea sp. and *Eucalyptus camaldulensis* (Table 2). Theoretical hydraulic conductivity was consistently larger in roots than in branches; but differences were not significant for all species (Table 3). Vessel implosion resistance was larger in roots than in branches of Acacia aneura, Hakea sp. and *Corymbia opaca* but was lower in roots of *Psydrax latifolia* than in branches. Vessel implosion resistance was similar in roots and branches of Erythrina vespertilio and Eucalyptus camaldulensis (Table 3). Finally, we also observed tyloses or vascular occlusion by parenchyma cells in some of the vessels of *Eucalyptus camaldulensis* and *Corymbia opaca*.

DISCUSSION

In this study, wood density of roots was significantly smaller than that of branches of the same species (cf. Table 2, O' Grady et al. 2009; Santini et al. 2016). Our linear regression analysis indicated that variation in root wood density was explained by variations in xylem vessel lumen area across the different species (r²=0.73, p=0.03, Figure 3). Thus, our first hypothesis, that roots within a given species have larger xylem vessel diameters (and consequently larger vessel lumen areas) and lower wood density than branches was supported. Numerous studies have found that xylem vessel size is larger in roots than in branches (and that wood density is lower in roots than in branches of the same species), and these studies also suggest that roots exhibit larger hydraulic conductivities and are at more risk of cavitation events (Pate et al. 1998; Martínez-Vilalta et al. 2002). The larger vessel size observed in roots compared to branches has been suggested to be a continuation of the hydraulic tapering observed in aboveground xylem with increasing tree height (Petit et al. 2008; Petit et al. 2010; Anfodillo et al. 2013). Whereas differences in vessel diameter among the tallest trees (10 – 15 m; Corymbia opaca, Eucalyptus camaldulensis and Erythrina vespertilio) were not determined by tree height, which was similar across these species, tapering cannot be discarded as a possible explanation to smaller vessel diameter in the shorter trees and shrubs.

We can confirm that for our studied species, theoretical hydraulic conductivity (K_s) was larger in roots than in branches, but that differences were not significant for all species. Lower wood densities in roots compared to branches can allow for shrinkage adjustments at high negative water potentials and reduce mechanical strength of this plant organ (Jacobsen *et al.* 2005; Santini *et al.* 2013). Roots are contained in a soil matrix that provides mechanical protection, indicating that safety demands may be decoupled from mechanical demands in plant roots compared to branches. The arrangement of xylem traits and the total fraction of tissues (i.e.
fibre wall fraction but also fibre lumen fraction, fraction of parenchyma) also influence wood
density (e.g. Zieminska *et al.* 2013, Lachenbrunch and McCulloh 2014), but this was not
considered in the present study.

Our previous work (Santini *et al.* 2016) found a strong negative relationship between K_s and vessel implosion resistance in stems of seven species, including all of the species from this study: i.e. Erythrina vespertilio, Acacia aneura, Hakea sp., Psydrax latifolia, Corymbia opaca and *Eucalyptus camaldulensis*. A meaningful relationship between K_s and vessel implosion resistance was not found in the current study. Instead, the arid-adapted species clustered with the species sampled in both habitats and exhibited the lowest K_s and highest vessel implosion resistance, whereas K_s and vessel implosion resistance was decoupled in the obligate riparian species (one deciduous, one evergreen). Thus, we rejected our second hypothesis, finding instead that the relationship between K_s and vessel implosion resistance was not maintained in roots of all of our studied species as it was for branches. Vessel implosion resistance is a vessel wall reinforcement metric, but in roots it might not be a relevant indicator of safety as in roots tensile forces in the water column are minimal and low wood densities in roots compared to branches may allow for adjustments at high negative water potentials (Jacobsen *et al.* 2005; Pratt et al. 2007). In the present root study, we found the sinker roots of Eucalyptus camaldulensis to have an extraordinarily high Ks, which is characteristic of this species (Roberts et al. 2016). The high K_s in *Eucalyptus camaldulensis* roots was the result of larger vessel lumen area and vessel density than in *Erythrina vespertilio* and *Corymbia opaca*, which have similarly large branch Ks to that of *Eucalyptus camaldulensis* branches (Santini et al. 2016). Tyloses (or vascular occlusion by parenchyma cells) are expected to reduce K_{s} , thus the large values of K_{s} in *Eucalyptus camaldulensis* and in *Corymbia opaca* suggests that flow restriction in root vessels with tyloses is compensated by large K_s in root vessels without tyloses. Tylose formation has been observed to occur in response to wounding and pathogen attack as to compartmentalize infection (Saitoh et al. 1993; Sun et al. 2008). Tylose development was observed infrequently in roots, and therefore had only minimal impact on our understanding of K_s for *Eucalyptus camaldulensis* and *Corymbia opaca*, particularly in light of the fact that the difference in K_s between these two species was more than ten-fold.

We found a strong relationship between log (vessel density) and vessel lumen area in roots
 (r²=0.72, p=0.03, Figure 4). Thus, *Erythrina vespertilio, Corymbia opaca* and *Eucalyptus camaldulensis* had the largest xylem vessels but also had low density of xylem vessels per
 sapwood area. This relationship between vessel density and vessel lumen area has been widely

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observed across plant species (Preston et al. 2006). In addition, there was a strong relationship 391 392 between outer pit aperture and log (vessel diameter) ($r^2=0.92$, p=0.002, Figure 5), which is 393 consistent with previous findings that pit anatomy scales with lumen diameter (Lazzarin et al. 394 2016). Li *et al.* (2016) found that samples stored in ethanol (such as samples from the current 395 study) exhibited shrinkage of pit membranes. We cannot confirm shrinking in our study, 396 nonetheless our results are comparable with our previous study (Santini et al. 2016). Further measurements are required to assess relationships among pit membrane pores, nanobubble size, outer pit aperture size, vessel size and vulnerability to cavitation events. However, 399 cavitation events can occur when the pressure difference between adjacent air and water filled vessels becomes large enough to pull the air-water interface through the nanopores of the pit membranes (Hacke et al. 2001; Martínez-Vilalta et al. 2002; McElrone et al. 2004; Schenk et al. 402 2015).

403 This study advances our understanding of root anatomy and hydraulic strategies of Australian 23 404 semi-arid species and confirms that roots exhibit larger xylem vessel diameters and lower wood $_{25}\ 405$ density than branches of the same species. In terms of functional grouping, Erythrina vespertilio, 406 the deciduous species exhibited high K_s (and thus a relatively efficient water transport system) 407 and low xylem vessel implosion resistance relative to Corymbia opaca and the evergreen 30 408 species. Thus, theoretically, *Erythrina vespertilio* can maintain relatively high transpiration rates 409 and in turn metabolic carbon uptake during the wet season (Enquist *et al.* 1999). At the same 410 time, *Erythrina vespertilio* can survive the dry season by reducing its canopy area to zero. 35 411 *Erythrina vespertilio* can also store large amounts of water and sugars in its parenchyma tissue 37 412 in order to support root and shoot respiration during the dry season when the canopy is absent 413 and carbon capture does not occur (Carlquist 2001). The phreatophytic riparian species 40 414 *Eucalyptus camaldulensis* exhibited the largest values of K_s and the lowest values of xylem vessel 42 415 implosion resistance. By having deep sinker roots and sustained access to the shallow 416 groundwater reservoir at *ca*. 3 m depth all year, *Eucalyptus camaldulensis* is at low risk of 417 cavitation during the dry season. *Corymbia opaca*, the second phreatophyte, exhibited much 47 418 smaller K_s values than *Eucalyptus camaldulensis*. In turn, *Corymbia opaca* roots exhibited similar 49 419 K_s and vessel implosion resistance values to the evergreen species with lateral shallow roots. 420 Relatively low K_s in *Corymbia opaca* can be explained as this species deploys deeper sinker roots 52 421 than *Eucalyptus camaldulensis* (up to 10 m depth) and because groundwater use in *Corymbia* 54 422 opaca is limited by high temperature and vapour pressure deficit (Cleverly et al. 2016b). 423 Alternatively, low K_s in the evergreen species: *Acacia aneura*, *Hakea* sp. and *Psydrax latifolia* that 424 deploy lateral shallow roots and rely on intermittent water resources such as rainwater can be 59 425 explained as a strategy to survive or tolerate drought.

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CONCLUSIONS

2 427 Deep sinker roots of *Eucalyptus camaldulensis* exhibited a highly efficient water transport 3 4 428 system with the largest theoretical hydraulic conductivity (K_s) of all species as a result of its 5 6 429 riparian habit and continuous access to shallow groundwater at *ca.* 3 m depth. Despite also 7 430 exhibiting deep sinker roots (to 8 m) and groundwater use, Corymbia opaca had root xylem 8 9 431 traits similar to those of the other evergreen trees found in both floodplain and sand plain 10 11 432 habitats: Hakea sp. Root xylem anatomy in the six-woody species of this study tended to cluster 12 13 433 by habitat or eco-hydrological niche. At one extreme are species with shallow lateral roots 14 434 which occupy the extraordinarily dry hardpans of the Ti Tree Basin and which access 15 16 435 intermittent water resources (i.e. rainwater) (i.e. arid-adapted Acacia aneura and Psydrax 17 18 436 latifolia). Roots of these arid-adapted species have the smallest vessel diameter, vessel lumen 19 20 437 area (and thus the largest vessel density) and outer pit aperture. At the other extreme are the 21 438 species which are restricted to riparian corridors, paleochannels and topographic depressions 22 23 439 in the floodplain habitat (Eucalyptus camaldulensis and Erythrina vespertilio), which exhibited 24 25 440 the largest root K_s, vessel diameter and vessel lumen area, with the smallest root wood density 26 441 and theoretical implosion resistance, indicating low overall investment in root xylem. Vessel 27 28 442 implosion resistance as a concept can and has been used as an indicator of vessel wall 29 30 443 reinforcement. Whilst we do not suggest that vessels do implode within roots (because tensile 31 444 forces in the water column are low) the metric can be better viewed as a measure of investment 32 33 445 in cell walls. The very low density of xylem vessels in roots of *Erythrina vespertilio* was due to a 34 35 446 large fraction of parenchyma that is likely to store large amounts of water. Thus, Erythrina 36 37 447 *vespertilio* may be able to support root and shoot respiration during the dry season when the 38 448 canopy is absent and carbon capture does not occur (Carlquist 2001). Xylem traits were found 39 40 449 to depend upon habitat and eco-hydrological niche, with the groupings including (i) arid-41 42 450 adapted shrubs and trees with shallow roots (Acacia aneura and Psydrax latifolia), (ii) trees 43 451 restricted to the floodplain habitat, both evergreen (Eucalyptus camaldulensis) and deciduous 44 45 452 (Erythrina vespertilio) and (iii) evergreen trees co-occurring in both floodplain and adjacent 46 47 453 sand plain habitats (Corymbia opaca and Hakea sp.). 48

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607 **Figures** 608

609 Figure 1. (A – D) Photographs of roots of four of our studied species, *Erythrina vespertilio*, 610 Acacia aneura and Psydrax latifolia exhibit shallow lateral roots while Corymbia opaca exhibits 611 deep sinker roots.

7 612 8 613 Figure 2. (A – F) Photographs of root wood micro-sections of the studied species from the Ti 9 614 Tree Basin, Northern Territory. Photographs illustrate xylem vessels (V), vessel walls (VW) and 10 615 fibre walls (FW). (A – D) Species were double-stained with safranin and alcian blue (1:2). (F - E) 616 *Corymbia opaca* and *Eucalyptus camaldulensis* were not stained due to the presence of resins. 617 Tyloses (Ty) were observed in some of the vessels.

14⁻⁰ 618 $_{15}$ 619 Figure 3. Relationship between root wood density and root vessel lumen area, the regression was: Y = $-14.78 \times + 0.67$ (r²=0.73, p=0.03). Sample numbers of individual primary roots (one 16 620 17 621 root per individual plant) for assessment of wood density were as follows: *Erythrina vespertilio* 18 622 (n=5), Acacia aneura (n=5), Hakea sp. (n=5), Psydrax latifolia (n=5), Corymbia opaca (n=3), 19 623 *Eucalyptus camaldulensis* (n=5). Sample numbers of individual primary roots (one root per 20 624 individual plant) for assessment of root vessel lumen area were as follows: Erythrina vespertilio 21 ⁻⁻₂₂ 625 (n=5), Acacia aneura (n=3), Hakea sp. (n=4), Psydrax latifolia (n=5), Corymbia opaca (n=3), 23 626 *Eucalyptus camaldulensis* (n=5). Symbols represent mean values \pm standard errors for different species: *Erythrina vespertilio* (crossed-circle), *Acacia aneura* (triangle), *Hakea* sp. 24 627 25 628 (square), Psydrax latifolia (diamond), Corymbia opaca (upside-down triangle), Eucalyptus 26 629 camaldulensis (circle). Different coloured symbols indicate functional groups (Yellow: 27 630 deciduous, green: evergreen and blue: evergreen phreatophytes). 28

29 631 Figure 4. Relationship between log (root vessel density) and root vessel lumen area, the 30 31 632 regression was: Log (Y) = $-67.69 \text{ X} + 2.17 \text{ (r}^2=0.72, p=0.03)$. Sample numbers of individual 32 633 primary roots (one root per individual plant) for assessment of root vessel density and root 33 634 vessel lumen area were as follows: *Erythrina vespertilio* (n=5), *Acacia aneura* (n=3), *Hakea* 34 635 sp. (n=4), *Psydrax latifolia* (n=5), *Corymbia opaca* (n=3), *Eucalyptus camaldulensis* (n=5). 35 636 Symbols represent mean values \pm standard errors for different species: *Erythrina vespertilio* 36 637 (crossed-circle), Acacia aneura (triangle), Hakea sp. (square), Psydrax latifolia (diamond), 37 38 638 Corymbia opaca (upside-down triangle), Eucalyptus camaldulensis (circle). Different coloured 39 639 symbols indicate functional groups (Yellow: deciduous, green: evergreen and blue: evergreen 40 640 phreatophytes). 41

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43 642 Figure 5. Relationship between root outer pit aperture and the logarithm (Log) of root vessel 44 643 diameter, the regression was: Y = 1.56 X - 0.93 (r²=0.92, p=0.002). Sample numbers of individual 45 46 644 primary roots (one root per individual plant) for assessment of pit apertures were n = 3 per 47 645 species. Sample numbers of individual primary roots (one root per individual plant) for 48 646 assessment of root vessel diameter were as follows: Erythrina vespertilio (n=5), Acacia aneura 49 647 (n = 3), Hakea sp. (n=4), Psydrax latifolia (n=5), Corymbia opaca (n=3), Eucalyptus 50 648 *camaldulensis* (n=5). Symbols represent mean values \pm standard errors for different species: 51 649 Erythrina vespertilio (crossed-circle), Acacia aneura (triangle), Hakea sp. (square), Psydrax 52 650 latifolia (diamond), Corymbia opaca (upside-down triangle), Eucalyptus camaldulensis (circle). 53 Different coloured symbols indicate functional groups (Yellow: deciduous, green: evergreen and 651 54 55 652 blue: evergreen phreatophytes).

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58 654 **Figure 6.** Relationship between root theoretical hydraulic conductivity (K_s) and log root xylem 655 vessel implosion resistance $(t/b)^2$. Sample numbers of individual primary roots (one root per

1 2 3 4 5 6 7 8	656 657 658 659 660 661 662	individual plant) for assessment of root K _s and log root $(t/b)^2$ were as follows: Acacia <i>aneura</i> (n = 3), <i>Hakea</i> sp. (n=4), <i>Psydrax latifolia</i> (n=5), <i>Corymbia opaca</i> (n=3), <i>Eucalyptus camaldulensis</i> (n=5) and <i>Erythrina vespertilio</i> (n=5). Symbols represent mean values \pm standard errors for different species: <i>Erythrina vespertilio</i> (crossed-circle), <i>Acacia aneura</i> (triangle), <i>Hakea</i> sp. (square), <i>Psydrax latifolia</i> (diamond), <i>Corymbia opaca</i> (upside-down triangle), <i>Eucalyptus camaldulensis</i> (circle). Different coloured symbols indicate functional groups (Yellow: deciduous, green: evergreen and blue: evergreen phreatophytes).
9 10 11 12 13 14 15 16 17 18 19 20 21 22 23	 663 664 665 666 667 668 669 670 671 672 673 674 	Figure 7. Relationship between root vessel diameter and root vessel wall thickness, the regression was: Y = 0.13 X + 1.18 (r^2 =0.99, p=0.004). Sample numbers of individual primary roots (one root per individual plant) for assessment of root vessel diameter and root vessel wall thickness were as follows: <i>Erythrina vespertilio</i> (n=5), <i>Acacia aneura</i> (n = 3), <i>Hakea</i> sp. (n=4), <i>Psydrax latifolia</i> (n=5), <i>Corymbia opaca</i> (n=3). <i>Erythrina vespertilio</i> (n=5) and <i>Eucalyptus camaldulensis</i> (n=5) were not included in the regression. Symbols represent mean values \pm standard errors for different species: <i>Erythrina vespertilio</i> (crossed-circle), <i>Acacia aneura</i> (triangle), <i>Hakea</i> sp. (square), <i>Psydrax latifolia</i> (diamond), <i>Corymbia opaca</i> (upside-down triangle), <i>Eucalyptus camaldulensis</i> (circle). Different coloured symbols indicate functional groups (Yellow: deciduous, green: evergreen and blue: evergreen phreatophytes).
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Tables

Table 1. Characteristics of the studied species and sampling areas at the Ti Tree Basin, Northern Territory. Xylem traits were vessel diameter, fibre and vessel wall thickness.

Functional group	Species	Growt h form	Area utilized for assessment of xylem traits (mm ²)*	Number of assessed xylem vessels (range)	Type of collected primary roots	Distance of collected primary lateral/sinker root from the main stem, range (m)	Tree height (m)
Deciduous	Erythrina vespertilio	Tree	~ 8	7 - 12	Lateral / Shallow	~ 2 - 6	10 – 15
Evergreen	Acacia aneura	Tree	~ 0.3	37 - 55	Lateral / Shallow	~ 0.5 - 1	5 - 10
	Hakea sp.	Tree	~ 1.3	39 - 41	Lateral / Shallow	~ 1 - 2	5 - 10
	Psydrax latifolia	Shrub	~ 0.08	60 - 85	Lateral / Shallow	~ 0.5 - 2	2 - 3
Evergreen	Corymbia opaca	Tree	~ 1.3	16 - 24	Sinker / Deep	~ 1-6	10 - 15
phreatophytes	Eucalyptus camaldulensis	Tree	~ 1.3	15 - 35	Sinker / Deep	~ 2-3	10 - 15

* Areas are approximate values per image.

Table 2. Xylem characteristics and wood density of tap roots of our collected species from the Ti Tree Basin, Northern Territory. NA indicates not assessed. Data from branches were provided by Santini *et al.* (2016). Values are means and standard errors. Values with the same letter within a column were not significantly different (p > 0.05) as tested with a one-way Analysis of Variance. A paired t-test was used to compare mean and standard errors of xylem characteristics and wood density between roots and branches, (*) indicates p < 0.05 and ns indicates non-significant differences. Sample number of individual tap roots for assessment of xylem traits (i.e. vessel diameter, fibre and vessel wall thickness) were as follows: *Erythrina vespertilio* (n=5), *Acacia aneura* (n = 3), *Hakea* sp. (n=4), *Psydrax latifolia* (n=5), *Corymbia opaca* (n=3), *Eucalyptus camaldulensis* (n=5). Samples number of individual tap roots for assessment of pit apertures were n = 3 per species. Sample number of individual tap roots for assessment of wood density were as follows: *Erythrina vespertilio* (n=5), *Acacia aneura* (n = 5), *Hakea* sp. (n=5), *Psydrax latifolia* (n=5), *Corymbia opaca* (n=3), *Eucalyptus camaldulensis* (n=5).

Species	Fibre wall thickness (µm)	Vessel wall thickness (µm)	Vessel diameter (µm)	Outer pit aperture (µm)	Conductive lumen fraction (mm ² mm ⁻ ²)	Wood density (g cm ⁻³)
Erythrina vespertilio Roots	NA	6.97 ± 0.21 ^{AD}	$188 \pm 6.15^{\text{A}}$	7.13±0.11 ^A	0.05±0.003 ^A	0.21 ± 0.009 ^₄
Branches	2.80 ± 0.11	1.98 ± 0.09	40.44 ± 1.634	NA	0.249 ± 0.020	0.40 ± 0.016
Paired <i>t</i> test (t;df)	NA	(21.59; 72)*	(29.75; 12)*		(4.97; 72)*	(3.73; 72)*
Acacia aneura Roots	$3.9 \pm 0.10^{\circ}$	$6.07 \pm 0.19^{\text{D}}$	$34.8 \pm 1.08^{\text{E}}$	$4.62 \pm 0.05^{\text{D}}$	$0.17 \pm 0.014^{\text{D}}$	$0.61 \pm 0.021^{\text{D}}$
Branches	4.01 ± 0.18	4.27 ± 0.19	35.14 ± 1.61	NA	0.098 ± 0.008	0.95 ± 0.07
Paired <i>t</i> test (t;df)	ns	(7.78; 72)*	ns		ns	(6.69; 72)*
<i>Hakea sp.</i> Roots	5.4 ± 0.12 ^A	9.48±0.15 E	60.3 ± 1.91 ^F	$6.16 \pm 0.15^{\text{B}}$	$0.12\pm0.009^{\text{de}}$	$0.82 \pm 0.023^{\text{E}}$
Branches	4.26 ± 0.15	2.71 ± 0.13	27.01 ± 2.18	NA	0.123 ± 0.023	0.712 ± 0.03
Paired <i>t</i> test (t;df)	(3.47; 60)*	(29.29; 72)*	(10.14; 12)*		ns	(2.12; 72)*
<i>Psydrax latifolia</i> Roots	$4.5\pm0.07^{\text{B}}$	$4.12 \pm 0.05^{\circ}$	$24.4 \pm 0.29^{\text{D}}$	$3.73 \pm 0.05^{\circ}$	0.37±0.008 ^c	0.58 ± 0.01 ^{CD}
Branches	6.79 ± 0.42	3.78 ± 0.13	20.19 ± 0.58	NA	0.133 ± 0.013	$0.73~\pm~0.017$
Paired <i>t</i> test (t;df)	(6.99; 60)*	ns	(5.13; 12)*		(5.92; 72)*	(2.95; 72)*
Corymbia opaca Roots	5.7±0.15 ^A	$11.5 \pm 0.29^{\text{B}}$	79.4 ± 3.12°	$5.88\pm0.16^{\scriptscriptstyle B}$	$0.08\pm0.006^{\rm AE}$	$0.49\pm0.002^{\text{BC}}$
Branches	3.36 ± 0.16	2.88 ± 0.17	39.41 ± 1.57	NA	0.258 ± 0.032	0.68 ± 0.022
Paired <i>t</i> test (t;df)	(7.14; 60)*	(37.30; 72)*	(12.87; 12)*		(4.45; 72)*	(3.73; 72)*
Eucalyptus camaldulensis Roots	5.3 ± 0.08 ^A	7.25±0.12 ^A	131±3.42 ^B	$6.49\pm0.14^{\scriptscriptstyle B}$	0.25 ± 0.011 B	$0.44 \pm 0.03^{\text{B}}$
Branches	3.31 ± 0.12	2.61 ± 0.09	36.67 ± 1.38	NA	0.258 ± 0.053	0.65 ± 0.03
Paired <i>t</i> test (t;df)	(6.07; 60)*	(20; 72)*	(30.41; 12)*		ns	(4.13; 72)*

Table 3. Calculated means and standard errors of theoretical hydraulic conductivity (K_s) and vessel implosion resistance (t/ b)², where t is the double-wall thickness (in μ m) and b is the hydraulic mean vessel diameter (in μ m). Different letters among columns indicate means were significantly different p < 0.05 as tested with a one- way Analysis of Variance.

Species	K _s (kg mm ⁻¹ MPa ⁻¹ s ⁻¹)	n	(t/b) ²	n
Erythrina vespertilio Roots	1.86 ± 0.1984	5	0.0068 ± 0.00037	5
Branches	0.383 ± 0.047	9	0.0096 ± 0.00006	3
Paired t test (t;df)	(6.48; 72)*		ns	
Acacia aneura Roots	0.4192 ± 0.1166	3	0.1179 ± 0.01017	3
Branches	0.109 ± 0.017	9	0.059 ± 0.00009	3
Paired t test (t;df)	ns		(10.86; 72)*	
Hakea sp. Roots	0.8140 ± 0.1511	4	0.0995 ± 0.0040	4
Branches	0.08 ± 0.019	9	0.0404 ± 0.00154	3
Paired <i>t</i> test (t;df)	(3.22; 72)*		(10.90; 72)*	
<i>Psydrax latifolia</i> Roots	0.2103 ± 0.0090	5	0.1094 ± 0.012	5
Branches	0.047 ± 0.006	9	0.14 ± 0.00092	3
Paired <i>t</i> test (t;df)	ns		(5.64; 72)*	
Corymbia opaca Roots	0.6654 ± 0.14	3	0.08536 ± 0.0079	3
Branches	0.356 ± 0.058	9	0.0213 ± 0.00047	3
Paired t test (t;df)	ns		(11.81; 72)*	
Eucalyptus camaldulensis Roots	6.837 ± 0.677	5	0.01617 ± 0.0013	5
Branches	0.328 ± 0.082	9	0.0237 ± 0.00001	3
Paired <i>t</i> test (t;df)	(28.57; 72)*		ns	

















