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Design and Synthesis of Short Amphiphilic Cationic Peptidomimetics Based on Biphenyl Backbone as Antibacterial Agents

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ABSTRACT

Antimicrobial peptides (AMPs) and their synthetic mimics have received recent interest as new alternatives to traditional antibiotics in attempts to overcome the rise of antibiotic resistance in many microbes. AMPs are part of the natural defenses of most living organisms and also they have a unique mechanism of action against bacteria. Herein, a new series of short amphiphilic cationic peptidomimetics were synthesized by incorporating the 3'-amino-[1,1'-biphenyl]-3-carboxylic acid backbone to mimic the essential properties of natural AMPs. By altering hydrophobicity and charge, we identified the most potent analogue **25g** that was active against both Gram-positive *Staphylococcus aureus* (MIC = 15.6 μ M) and Gram-negative *Escherichia coli* (MIC = 7.8 μ M) bacteria. Cytoplasmic permeability assay results revealed that **25g** acts primarily by depolarization of lipids in cytoplasmic membranes. The active compounds were also investigated for their cytotoxicity to human cells, lysis of lipid bilayers using tethered bilayer lipid membranes (tBLMs) and their activity against pre-established biofilms of *S. aureus* and *E. coli*.

Keywords: antimicrobial peptide, peptidomimetics, membrane disruption, antibiofilm activity.

1. Introduction

Antibiotic resistance in bacteria is a major concern facing global public health. Multidrug resistant strains of Gram-positive bacteria such as methicillin-resistant *Staphylococcus aureus* (MRSA), methicillin-resistant *Staphylococcus epidermidis* (MRSE), and vancomycin-resistant *Enterococci faecalis* (VRE), and Gram-negative bacteria such as *Escherichia coli*, *Klebsiella pneumoniae*, and *Pseudomonas aeruginosa*, have emerged as major causes of hospital and community-acquired infections [1]. Particularly, Gram-positive bacteria such as MRSA account for a high percentage of hospital-acquired infections [1, 2]. Due to the increasing resistance of bacterial strains against conventional antibiotics, efforts have been made to investigate naturally-occurring antimicrobial peptides (AMPs) and their derivatives as alternative antimicrobial agents [3-5]. AMPs such as PMX30063 and LTX109 are currently in clinical trials [6-8]. Although some promising AMPs are in the pipeline, there is still an urgent need for the development of new antibiotic scaffolds.

AMPs are widespread in nature and serve as the first-line of defense against microbial attack in insects, plants, amphibians, and mammals [9-12]. Unlike conventional antibiotics, AMPs act via non-receptor interactions, which make it difficult for bacteria to develop resistance to AMPs. Most cationic antimicrobial peptides possess a rigid secondary structure and adopt an amphipathic conformation such that their positively-charged face interacts electrostatically with the negatively-charged membrane surface, while their hydrophobic face inserts into the lipophilic interior of the membrane [13, 14]. The broad-spectrum antimicrobial peptide pexiganan, which acts via disruption of bacterial cell membranes, has reached phase III clinical trials [15]. Only a few naturally-occurring AMPs have been used clinically, including polymyxin B and colistin (polymyxin E), due to their *in vivo* toxicity, susceptibility to proteolytic degradation, poor activity in the presence of salts and cytotoxicity to the host cells [16]. Produced through solid-phase synthesis, AMPs also have high manufacturing costs [17].

The drawbacks of conventional AMPs have stimulated the development of peptidomimetics [18], which are synthetic nonpeptidic molecules designed to mimic the properties of peptides. The various kinds of peptidomimetics include α -peptides [19], β -peptides [20], peptoids [21-23], β -turn mimetics [24], cationic β^{3R3} -peptides [25] and lipopeptides [26]. In particular, a number of structurally simple, cationic peptidomimetics possessing natural or unnatural amino acids and with amphipathic character have been investigated as antibacterial agents [27, 28]. Svendsen and co-workers have synthesized a range of peptides of variable length utilizing arginine and tryptophan aminoacids, and showed good minimal inhibitory concentration (MIC) of 2.5 μ g mL⁻¹ against *Staphylococcus aureus* and 5 μ g mL⁻¹ against *Escherichia coli* [29]. An ultra-short pyrazole-based peptidomimetics showed a MIC of 4 μ g mL⁻¹ against MRSA and was four times more potent than melittin [30]. Pyne and co-workers have developed C2-symmetric binaphthyl-containing peptidomimetics that showed excellent antimicrobial activity against both Gram-positive and Gram-negative bacterial pathogens [31]. Halder and co-workers designed aryl-alkyl-lysine-based peptide mimics that mimicked the membrane-active properties of natural AMPs [32]. Their group also investigated cationic small molecules with spatial control of hydrophobicity to minimize toxicity against human erythrocytes while still maintaining antibacterial activity [33].

Our research group has recently synthesized short glyoxamide-based peptidomimetics via the ring-opening reaction of N-naphthoylisatins with amines and amino acids [34, 35]. As part of an ongoing program to develop short antimicrobial peptidomimetics, we were interested in utilizing the biphenyl backbone due to its frequent presence in medicinal chemistry [36, 37]. In an analysis of scaffolds of pharmacologically active molecules, biphenyl was found to be present in 2.1% of reference drug molecules [38, 39]. Furthermore, the importance of the biphenyl unit is shown by their presence in several natural products [38-

40], such as the antibacterial compounds MC21-A (**1**) and MC21-B (**2**) isolated from marine bacterium *Pseudoalteromonas phenolica* [41, 42]. The biphenyl-containing antibacterial compounds biphenomycin A (**3**) and B (**4**) were isolated from the cultured broth of *Streptomyces griseorubiginosus* 43608 [43-45]. Antibiotics such as arylomycin A2 (**5**), arylomycin B2 (**6**) and vancomycin also contain biphenyl moieties.

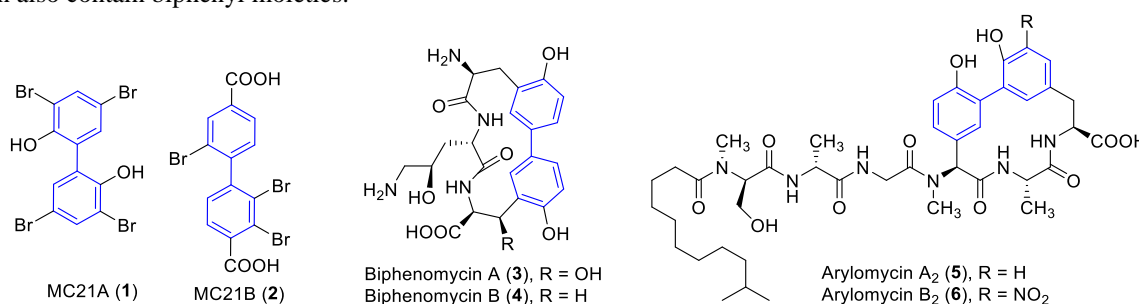
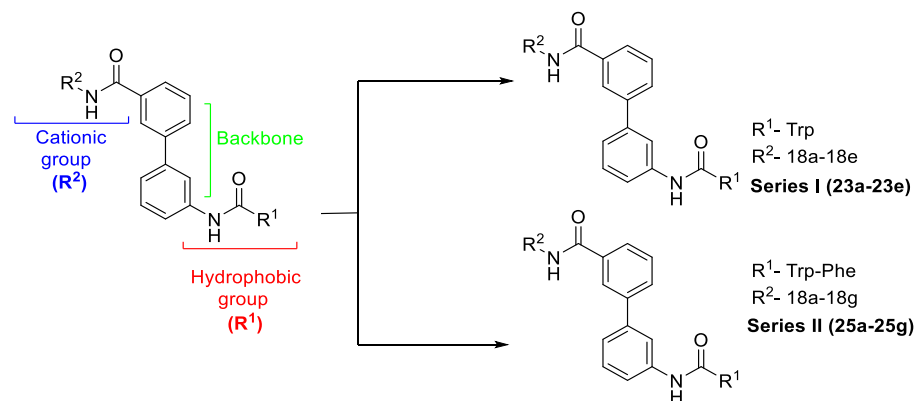


Fig. 1 Biphenyl-containing antibacterial compounds, with the biphenyl motif highlighted in blue.

In this work, we designed a unique scaffold for developing short antimicrobial peptidomimetics by utilizing a 3, 3'-substituted biphenyl unit as the key hydrophobic backbone to mimic the structural and biological properties of many AMPs. The segregation of hydrophobic (R^1) and cationic (R^2) groups via the biphenyl core confers amphipathic character to the entire molecule (**Fig. 2**). Importantly, the modular design of this scaffold allows for ready optimization of the biphenyl-based peptidomimetics simply by varying the nature of the hydrophobic (R^1) and cationic (R^2) groups.



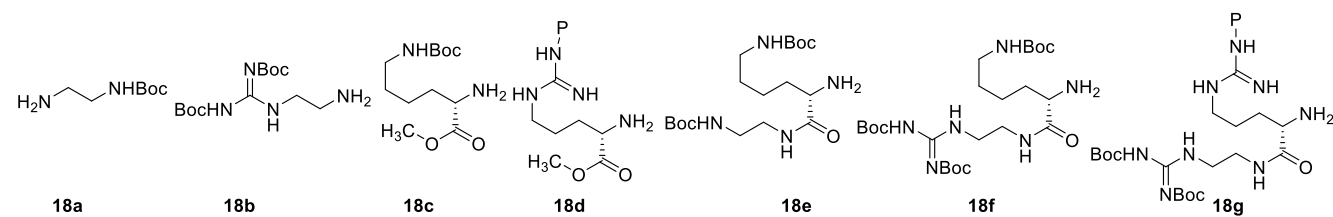
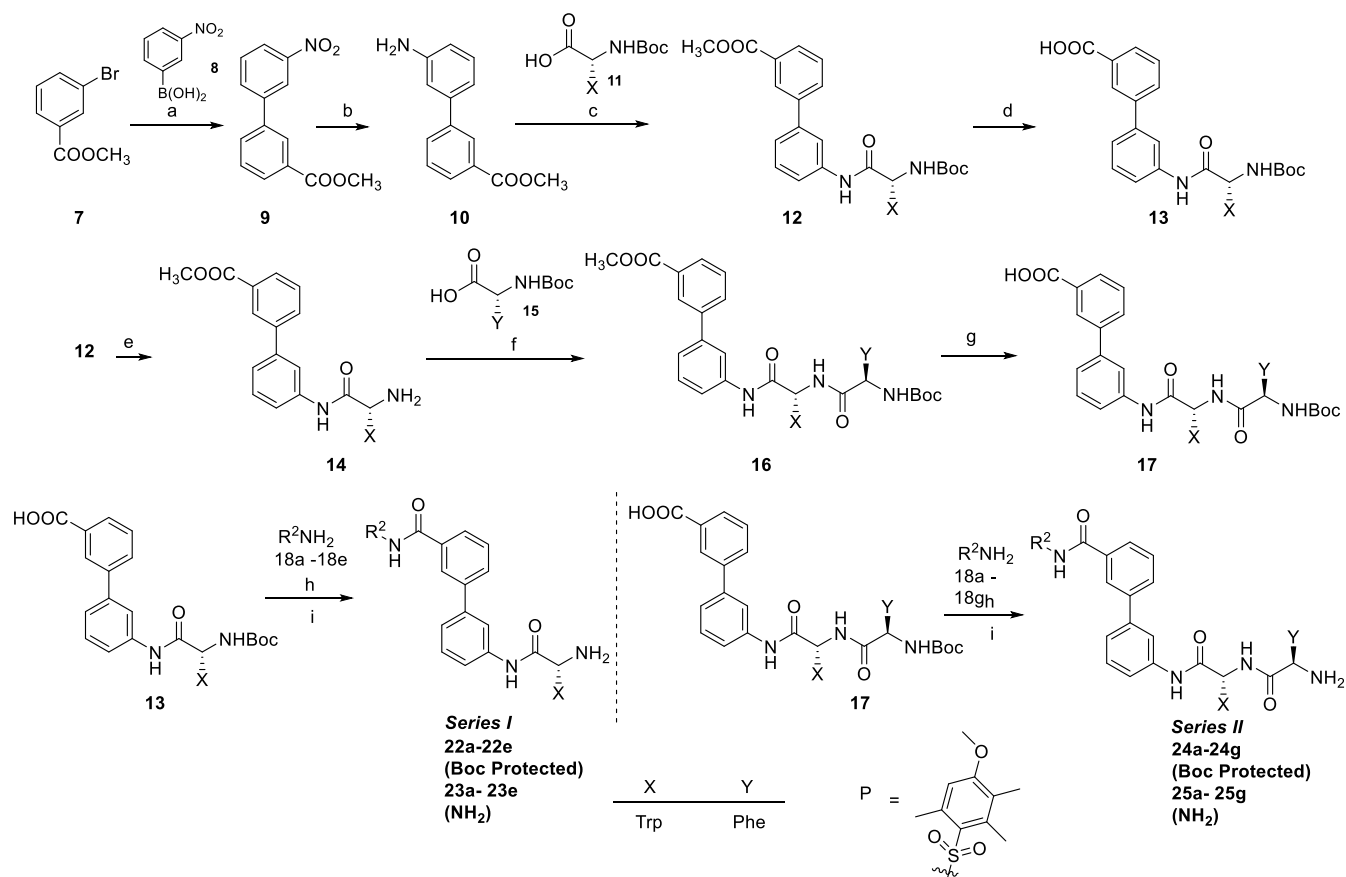
The biphenyl-based peptidomimetics synthesized in this work were classified into two series based on the length of the hydrophobic group (R^1) attached to the molecule. Series I (**23a-23e**) incorporated a single tryptophan (Trp) group at the 3-position of biphenyl, whereas series II (**25a-25g**) incorporated a tryptophan-phenylalanine (Trp-Phe) dipeptide at the same position. The selection of Trp was based on its ability to interact with the interfacial region of the bacterial membranes, thereby anchoring the biphenyl derivatives to lipid bilayers [46]. The cationic groups (R^2) were incorporated at the 3'-position of biphenyl, and they included 1,2-diaminoethane, 1-(2-aminoethyl)guanidine, and amino acids such as arginine and lysine. This demonstrated the importance of short non-natural amine and guanidine groups over cationic amino acids. Arginine and 1-(2-aminoethyl)guanidine were chosen because the guanidine group exhibits a stronger electrostatic interaction and more extensive hydrogen bonding with the negatively-charged phospholipids of the bacterial cell membrane [47].

The synthesized compounds were evaluated for biological activity against Gram-positive and Gram-negative bacterial strains. In addition, cytotoxicity was also investigated for selected potent antimicrobial compounds. To study how the active compounds interacted with lipid membranes, tethered bilayer lipid membranes (tBLMs) in association with electrical impedance spectroscopy was employed. In addition, the ability of the compounds to alter cytoplasmic permeability was assessed by using the membrane potential-sensitive cyanine dye diSC3-5. Finally, the ability of the compounds to inhibit the biofilm formation of *S. aureus* and *E. coli* was also evaluated.

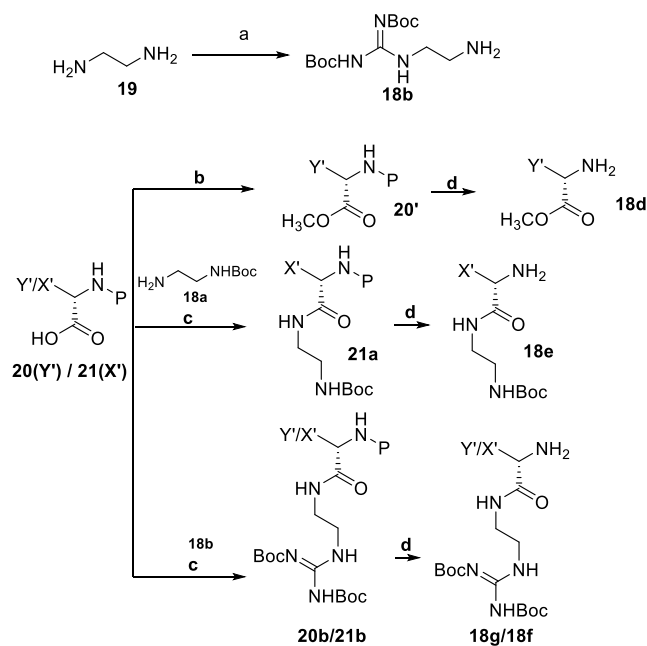
2. Results and discussion

2.1 Design and synthesis of biphenyl derived peptidomimetics

The acid **17** was synthesized by deprotecting the Boc group of **12** with TFA, followed by amide formation with **15** and subsequent ester hydrolysis using aqueous sodium hydroxide. On coupling **17** with different amines (**18a-18g**) respective amides **24a-24g** were yielded and the deprotection of **24a-24g** gave the series II compounds (**25a-25g**). The synthesis of amines **18b**, and **18d-18g** were shown in scheme2. The guanylation reaction using *N,N*-Di-Boc-*1H*-pyrazole-1-carboxamide provides the corresponding amine **18b**. The amino ester **18d** was prepared from NH_2 -Arg(Mtr)-OH using thionyl chloride and methanol. The acids **21(X')**, **21(X'')**, **20(Y')** were used to synthesize the amides **21a**, **20b**, **21b** using PyBOP/ CH_2Cl_2 coupling agent, and subsequent Fmoc cleavage using piperidine/DMF to afford **18e-18g**.



Scheme 1. Synthesis of compounds 13 and 17. Reagents and conditions: a) 8 (1.1 equiv), Pd(PPh₃)₄ (0.03 equiv), aq. 2M Na₂CO₃ (3.0 equiv), 1,4-dioxane, reflux, overnight, 65%; b) 10% Pd/C, H₂ balloon, THF, rt, overnight, 90%; c) EDCI (1.2 equiv), HOBT (1.0 equiv), DIEA (2.5 equiv), DMF, rt, 8 h, 63%; d) aq. 1N NaOH (2.0 equiv), THF, MeOH, rt, overnight, 90%; e) TFA, CH₂Cl₂, RT, 4h, 89%; f) EDCI (1.2 equiv), HOBT (1.0 equiv), DIEA (2.5 equiv), DMF, rt, 8h, 54%; g) aq. 1N NaOH (2.0 equiv), THF, MeOH, rt, overnight, 89%; h) EDCI (1.2 equiv), HOBT (1.0 equiv), DIEA (2.5 equiv), DMF, rt, 8 h-overnight, 52-84%; i) TFA, CH₂Cl₂, RT, 4 h-overnight, 49-89%.



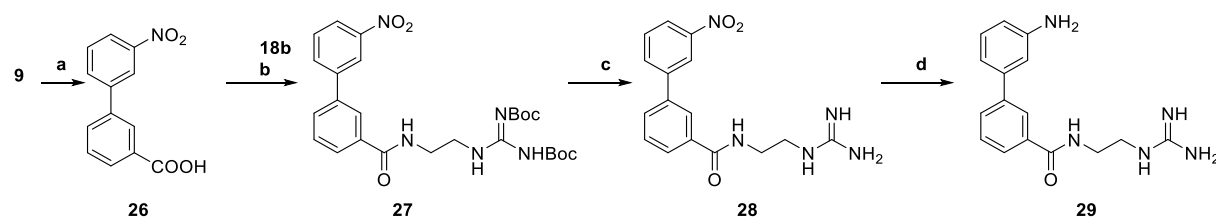
X'	Y'	P	% Yield
20'	Arg	Fmoc	90
21a	Lys	Fmoc	54
20b	Arg	Fmoc	34
21b	Lys	Fmoc	41

X'	Y'	% Yield
18d	Arg	85
18e	Lys	89
18g	Arg	78
18f	Lys	79

Scheme 2. Synthesis of 18b, 18d-18g. Reagents and conditions: a) *N,N'*-Di-Boc-1*H*-pyrazole-1-carboxamidide (0.1 equiv), THF, rt, 30 min, crude; b) SOCl₂ (1.5 equiv), MeOH, rt, overnight; c) PyBOP (1.1 equiv), HOBT (1.0 equiv), DIEA (3.0 equiv), CH₂Cl₂, rt, overnight; d) Piperidine (2.0 equiv), DMF, RT, overnight.

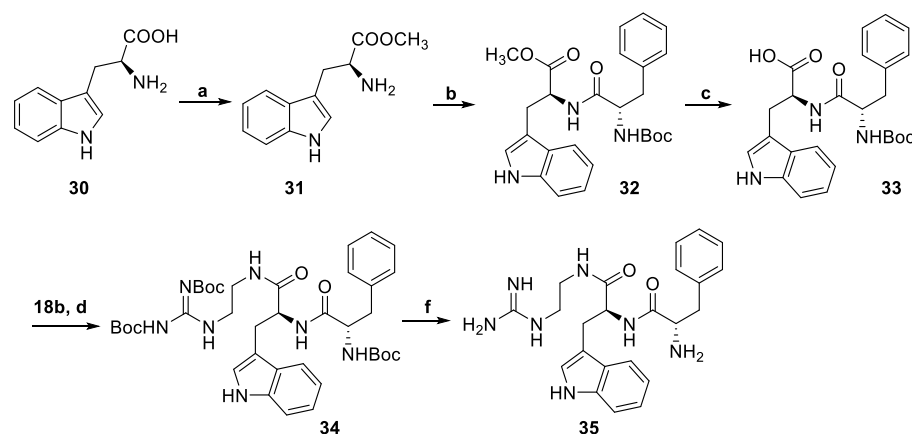
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The biphenyl derivative **29** attached with guanidine without any hydrophobic group was synthesized as shown in scheme 3. The biphenyl ester **9** was hydrolyzed to acid **26** and coupling with **18b** to afford the amide **27** and, subsequent deprotection with TFA gave **29**. Finally, **35** without the biphenyl backbone but intact with hydrophobic and cationic group was synthesized from L-Tryptophan **30**. The acid **33** was obtained by the sequential formation of dipeptide **32** from **30**, followed by ester hydrolysis. The amide **34** was obtained on coupling **33** with **18b**, and it was deprotected with TFA to yield **35** (Scheme 4).



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Scheme 3. Synthesis of compound **29**. Reagents and conditions: a) aq. 1N NaOH (2.0 equiv), THF, MeOH, rt, overnight, 90%; b) EDCI (1.2 equiv), HOBT (1.0 equiv), DIEA (2.5 equiv), DMF, rt, overnight, 80%; c) TFA, CH₂Cl₂, rt, 4h-overnight, 90%; d) *N,N'*-Di-Boc-1*H*-pyrazole-1-carboxamide (1.0 equiv), Et₃N (2.0 equiv), THF, RT, 6 h, 85% e) 10% Pd/C, H₂ balloon, rt, 5 h, 40%.



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Scheme 4. Synthesis of compound **35**. Reagents and conditions: a) SOCl₂ (1.5 equiv), MeOH, RT, overnight, 90%; b) EDCI (1.2 equiv), HOBT (1.0 equiv), DIEA (2.5 equiv), DMF, rt, overnight, 65%; c) 1N NaOH_(aq) (2.0 equiv), THF, MeOH, rt, overnight, 91%; d) EDCI (1.2 equiv), HOBT (1.0 equiv), DIEA (2.5 equiv), DMF, rt, overnight, 45%; e) TFA, CH₂Cl₂, RT, overnight, 47%.

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2.2 Antimicrobial activity study

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The antimicrobial activities of the newly synthesized peptidomimetics were assessed against the Gram-positive bacterium *Staphylococcus aureus* [SA38], and two Gram-negative bacteria, *Pseudomonas aeruginosa* [PA01], and *Escherichia coli* [K12] are summarized in table 1.

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The series I compounds **23a-23e** were tested against Gram-positive bacterium *Staphylococcus aureus* [SA38]. The compound **23b** (MIC = 62.5 μM) in which the biphenyl group segregated with tryptophan hydrophobic group and simple guanidine cationic group showed more activity compared to simple amine cationic group **23a** (MIC = 125 μM). The compound **23b** containing simple guanidium cationic moiety which mimic the arginine amino acid found in natural AMPs displayed the good activity. The MIC values of **23c** (31.2 μM) and **23d** (62.5 μM) attached with the Lys-ester and Arg-ester cationic groups are compared with **23a** and **23b** by maintaining the total net charge and hydrophobicity of the compounds. We noticed that the amine containing cationic group of lysine methyl ester **23d** showed four-fold increase in antibacterial activity compared to the **23a**. Interestingly, the similar antibacterial activity of **23b** and **23d** revealed that the simple guanidine cationic group is enough to mimic the arginine. The antibacterial activity of compound **23e** (MIC = 31.2 μM), and **23c** remains same even though the net cationic charge is increased. This could be due to the imbalance of the peptide hydrophobicity and charge distribution. Although, the series I compounds **23a-23e** found to be active against *S. aureus*, these compounds did not have significant activity against the two Gram-negative bacteria, *Pseudomonas aeruginosa* [PA01], and *Escherichia coli* [K12]. Based on the results from series I, increase in the net cationic charge did not show any profound antibacterial effect against Gram-positive and Gram-negative bacterial strains.

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The series II compounds **25a-25g** were made on hypothesis that the increase in hydrophobicity will enhance the antibacterial activity and to understand the effect of hydrophobicity along with the cationic character against the Gram-negative pathogens. All the compounds **25a-25g** were tested against the Gram-positive bacterium *Staphylococcus aureus* [SA38]. As expected **25a** (MIC = 62.5 μM) and **25b** (MIC = 15.6 μM) containing excess hydrophobic phenylalanine group showed excellent activity compared to less hydrophobic compounds **23a** (MIC = 125 μM) and **23b** (MIC = 62.5 μM). The increase in hydrophobicity of **23c** (31.2 μM) did not improve the MIC of **25c**. Whereas, in compound **25d** (31.2 μM) the activity increased two-fold compared to **23d** (62.5 μM). This may be resulted due to the increase in hydrophobic bulkiness along with the extensive hydrogen bonding of guanidine group. **25e** (MIC = 31.2 μM) retained the same activity compared to **23e** (MIC = 31.2 μM) though the hydrophobicity is increased may be due to the net positive charge equals with the number of hydrophobic group. We hypothesized that if the net positive charge is important in the activity of **25e** and if the amine groups are replaced with guanidine groups, the activity should increase.

223 **25f** (31.2 μM) showed same activity after replacing with one guanidine of **25e**. As anticipated the compound **25g** (MIC = 15.6
224 μM) replaced with the two-guanidine by attaching arginine in place of lysine group in **25f** showed excellent activity compared to
225 **25e** (MIC = 31.2 μM) probably due to the strong hydrogen bonding with the negatively charged phospholipids of bacterial cell
226 membrane.

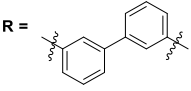
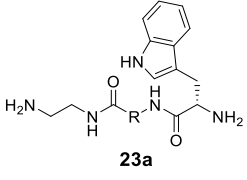
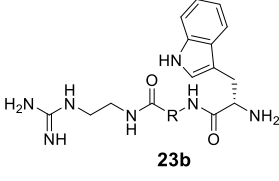
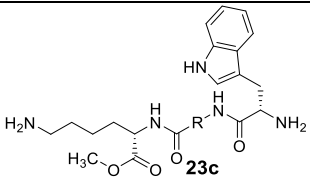
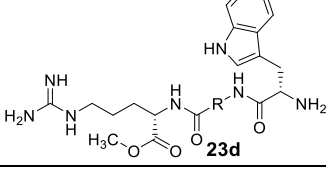
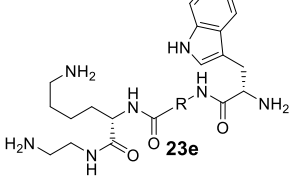
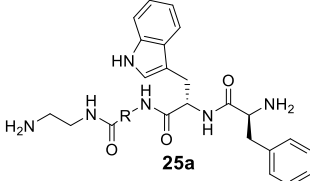
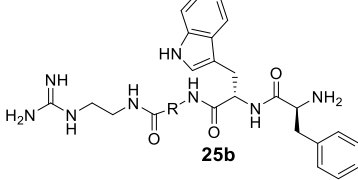
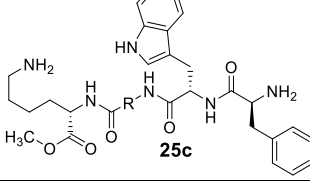
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228 **25a-25g** were also tested against Gram-negative bacteria *Escherichia coli* [K12]. Compounds **25a**, **25c**, **25d** did not show notable
229 antibacterial activity. The compound **25b** (MIC = 31.2 μM), with the increased hydrophobicity compared to **23e** (MIC = >125
230 μM) showed dramatically four-fold improvement against *E. coli*. The antibacterial activity of cationic peptides on Gram-negative
231 bacteria is influenced by two steps to overcome the outer-membrane barrier. The high membrane bound concentration of the
232 cationic peptides facilitates the outer-membrane permeabilization and in the final step the inner membrane damage causes the cell
233 lysis [48]. In this step, the hydrophobic interactions become dominant. This clearly suggests the importance of activity of **25b**
234 compared to **23e**. The compounds **25f** (MIC = 15.6 μM), and **25g** (MIC = 7.8 μM) containing guanidine cationic groups displayed
235 good activity against *E. coli* compared to **25c** (MIC = 125 μM) and **25e** (MIC = 31.2 μM) containing the amine cationic groups.
236 These results evidently showed that the electrostatic attraction played the decisive role for the excellent activity against the *E. coli*.
237 The compound **29** (62.5 μM) retains the biphenyl backbone showed the similar activity of **23b** against *S. aureus* and *E. coli* even
238 though the hydrophobic tryptophan group was removed. This could be due to the hydrophobicity of the biphenyl backbone.

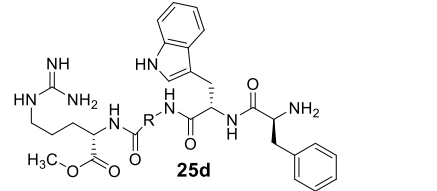
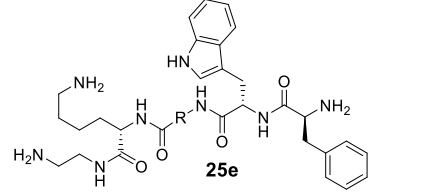
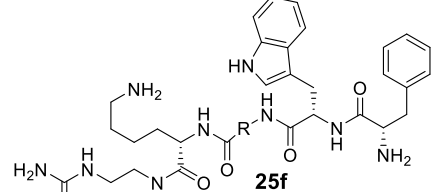
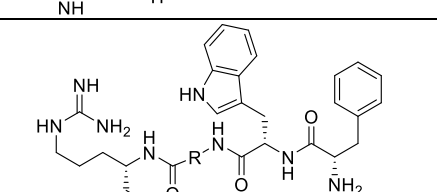
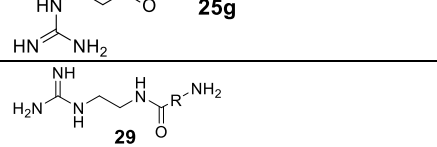
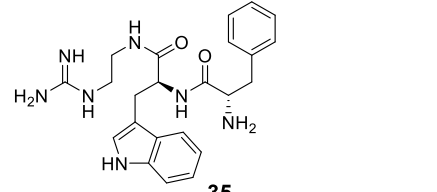
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240 Finally, the MIC results of **35** without the biphenyl backbone (>250 μM) compared with **23b** (62.5 μM) against *Staphylococcus*
241 *aureus* [SA38]. The antibacterial activity is totally lost after the removal of biphenyl backbone from the active compound **23b**.
242 The MIC results confirms the biphenyl backbone plays a major role in balancing the hydrophobicity and amphipathicity which are
243 considered as the key parameters for the antibacterial activity against Gram-positive and Gram-negative bacterial strains.

244
245 Overall, the structure activity relationship (SAR) draw the inference regarding the antibacterial activity of the biphenyl backbone
246 peptidomimetic derivatives against *S. aureus* and *E. coli*. Firstly, the series II compounds with excess hydrophobic group showed
247 good to excellent antimicrobial activity against *S. aureus* and *E. coli*. Secondly, the amphipathic nature played a crucial role in
248 determining the antimicrobial activity. The antibacterial activity of compounds **25c-25f** remain intact against *S. aureus* even after
249 increasing the net positive charge. Among all the synthesized biphenyl derivatives, **25b** and **25g** displayed the most potent
250 antibacterial activity against *S. aureus* (15.6 μM). Finally, **25g** revealed that an increase in cationic charge by utilizing guanidine
251 groups along with increase in hydrophobicity produced excellent activity against Gram-negative *E. coli* (7.6 μM) compared to the
252 corresponding amine compound.

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254 The active peptidomimetic compound **25g** which contains less number of amino-acids is comparable with the MIC of MSI-78.
255 MSI-78 (also known as pexiganan) is a chimera of magainin and melittin peptides [49]. This peptide was chosen as a reference
256 compound as it is currently in Phase-III clinical trials and one of the most well-studied amphipathic antimicrobial peptides.
257 Resistant strains of bacteria cannot be generated against MSI-78 even after repeated exposure to sub-inhibitory concentrations
258 [15], and many studies have shown that the antimicrobial action of the peptide involves disruption of bacterial membranes [50,
259 51].
260

Table 1. Antibacterial activities of biphenyl peptidomimetic derivatives.

 R =	MIC (μM)	
	Gram +ve <i>S.</i> <i>aureus</i>	Gram -ve <i>P.</i> <i>aeruginosa</i> / <i>E. coli</i>
 23a	125	>250 / >125
 23b	62.5	>250 / >125
 23c	31.2	>250 / >125
 23d	62.5	>250 / >125
 23e	31.2	>250 / >125
 25a	62.5	250 / >125
 25b	15.6	250 / 31.2
 25c	31.2	250 / 125

	31.2	250 / 125
	31.2	250 / 31.2
	31.2	NT / 15.6
	15.6	NT / 7.8
	62.5	>250 / >125
	>250	NT / >125
MSI-78 [15, 52, 53]	3.2 – 6.45 (NT)	*3.2 – 6.45 (NT)
(NT) – Not tested; MSI-78 [15, 52, 53] – Literature Value * <i>E. coli</i>		

2.3 Toxicity against human cells

To further evaluate the utility of these biphenyl derivatives as antimicrobial agents, their specificity for bacterial cells over human cells was determined. Therefore, the in vitro toxicity of the most active compounds (**23b**, **23d**, **25b-25g**) was assessed against MRC-5 normal human lung fibroblasts using the Alamar Blue (Resazurin) assay [54]. A dose-response curve was generated for each compound (shown in [supplementary material Fig. S2](#)) at concentrations ranging from 1-350 μM and their IC_{50} values were determined (shown in [supplementary material Fig. S3](#)). All of the tested compounds displayed very low toxicity ($\text{IC}_{50} > 300 \mu\text{M}$) towards human cells. Although amphipathic antibacterial agents are often cytotoxic, our compounds did not show noticeable cytotoxicity up to the concentration of 350 μM .

2.4 Lipid bilayer membrane interactions

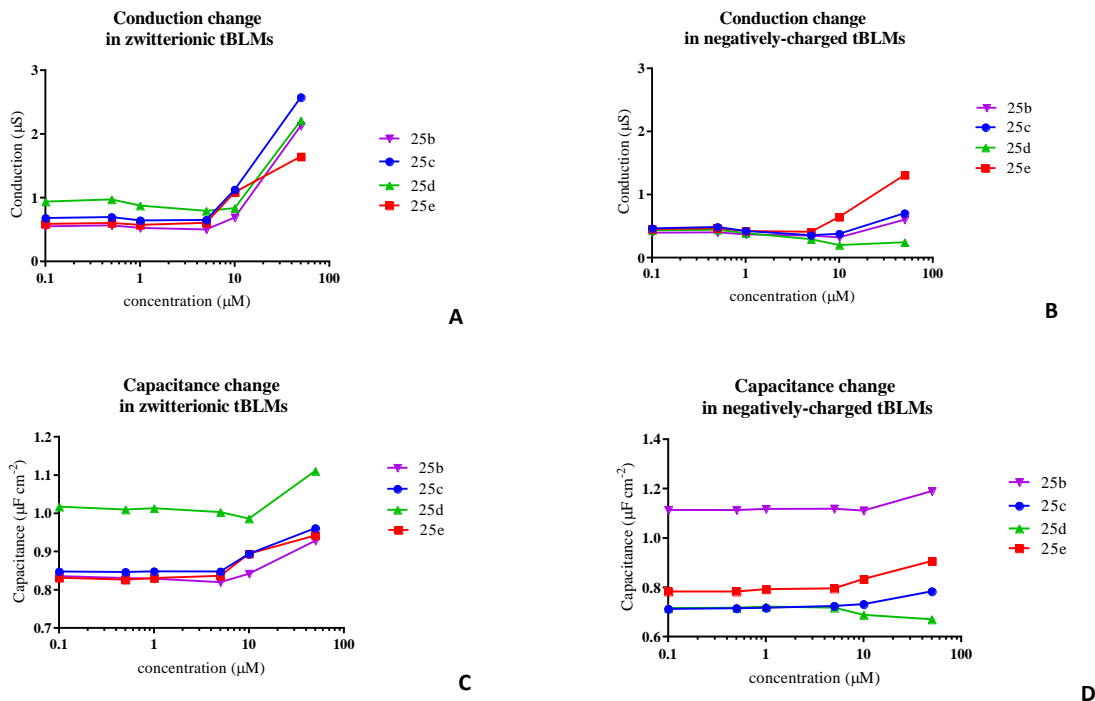
The compounds ability to interact with lipid bilayers was assessed using tethered bilayer lipid membranes (tBLMs) in association with AC electrical impedance spectroscopy.[55, 56]. Compounds which are shown membrane conduction at concentration of 10 μM are only discussed. Membrane conduction responses of a zwitterionic phospholipid membrane to compounds **25b-25e** increased with increasing concentration of compounds ([Fig. 3A](#)). Significant conduction changes are evident at concentrations equal to or higher than 10 μM . Interestingly, these responses are more muted in negatively-charged phospholipid membranes which mimic the negatively charged bacterial cell membranes ([Fig. 3B](#)).

The changes in membrane conduction are modest when compared to that produced by α -hemolysin or gramicidin-A [56, 57] which produce ion channels in the membranes. This would indicate that it is unlikely that these compounds form membrane

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spanning ion channel-like pores. Alternatively, the changes in membrane conduction could be the result of the formation of toroidal pore-like structures in the membrane. Either the compounds form a part of the lipidic toroidal pore themselves, or they independently induce phospholipidic toroidal pore formation by altering the critical packing parameter of the membrane[58].

The membrane capacitance changes increased with compound concentration in both zwitterionic (Fig. 3C) and negatively-charged (Fig. 3D) tBLMs, with the exception of compound 25d which exhibited a slight decrease in capacitance at high concentrations in negatively-charged membranes. An increase in capacitance is indicative of a thinning of the lipid membrane and/or an increase in membrane dielectric caused by the presence of water molecules. This indicates these compounds may follow mechanism of pore formation like AMPs by membrane thinning effect [59].



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Fig. 3. Change in conduction in (A) zwitterionic and (B) negatively-charged tBLMs in response to increasing concentrations of compounds 25b-25e. Change in membrane capacitance in (C) zwitterionic and (D) negatively-charged tBLMs in response to increasing concentrations of compounds 25b-25e.

Notably, the change in membrane conduction at 10 μM was correlated with antimicrobial activity, with compounds having lower MIC values (compounds 25c-25e) showing greater positive changes in membrane conduction in zwitterionic lipids than compounds with higher MIC values 25b (Fig. 4A). In negatively-charged lipids, however, the results were mixed and there was no clear relationship between MIC and conductance change (Fig. 4B). This would suggest that the effectiveness of these compounds is not so much related to their overall net cationic charge, rather their ability to insert into the bilayer proper. To test the hypothesis that the tryptophan moiety plays a significant role in membrane disruption, the change of conductance induced by compound 29, which lacks any Trp residue, was tested. As expected, compound 29 exhibited no change in membrane conduction in either zwitterionic or negatively-charged lipids, demonstrating the importance of the Trp residue.

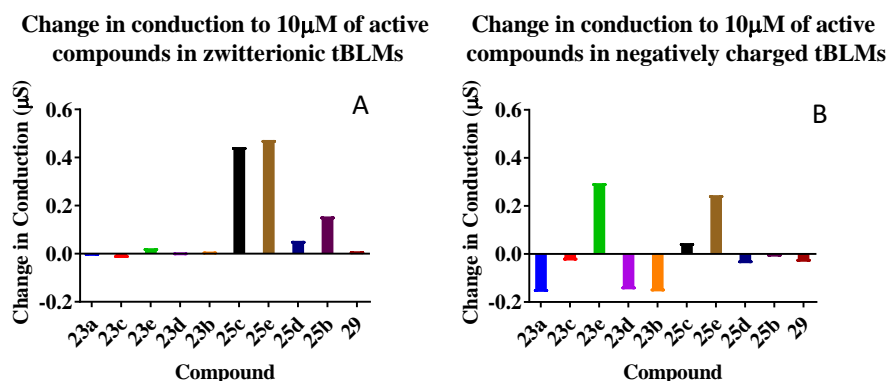


Fig. 4. The change in tBLM conduction of the various antimicrobial compounds at 10 μM in (A) zwitterionic and (B) negatively-charged membranes.

2.5 Cytoplasmic membrane depolarization

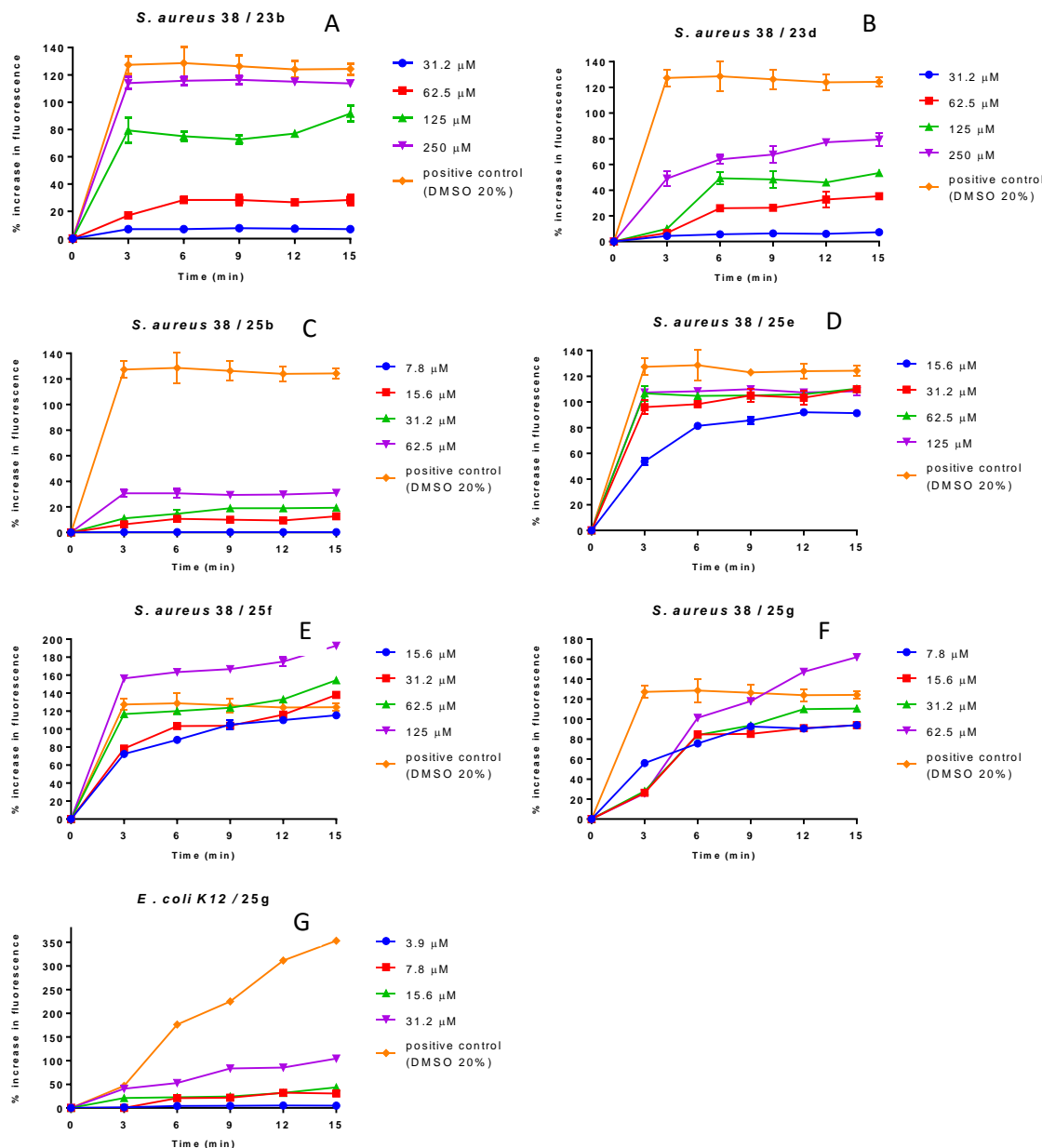
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We evaluated the disruption effect of the biphenyl derivatives on the bacterial cytoplasmic membrane using the membrane potential-sensitive dye diSC3-5 (3,3'-dipropylthiadicarbocyanine iodide). The distribution of diSC3-5 between the cell membrane and periphery medium is dependent on the cytoplasmic membrane potential gradient. The dye readily partitions into the bacterial cell membrane and aggregates within the membrane, causing self-quenching. If the antimicrobial compounds perturb the cell membrane, it can lead to the loss of the membrane potential gradient, causing the dye to be released into the medium. As a result,

314 the fluorescence intensity of the dye increases. As shown in Fig. 5, compounds 23b, 23d, 25b, 25e, 25f, and 25g (added at 0.5×,
 315 1×, 2× or 4× MIC) induced disruption of the cytoplasmic membrane of *S. aureus* in a time and concentration-dependent manner.
 316 Interestingly, the most active compound 25g showed an increase in fluorescence intensity even at sub-MIC levels within 3 min.
 317 Similarly, increases in fluorescence intensity of 25e and 25f also occurred at 1× MIC within 3 minutes.
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319 We also conducted the cytoplasmic permeability assay on Gram-negative bacterium *E. coli* (K12) with the most active compound
 320 25g. 25g also perturbs the cell membrane led to similar disruption of diSC3-5 fluorescence intensity (Fig. 5G), indicating that the
 321 cell membrane of both Gram-positive and Gram-negative bacteria can be disrupted. Taken together, these observations indicate
 322 that the biphenyl amphiphilic compounds can readily permeabilize the bacterial membrane, thus resulting in bacterial death.

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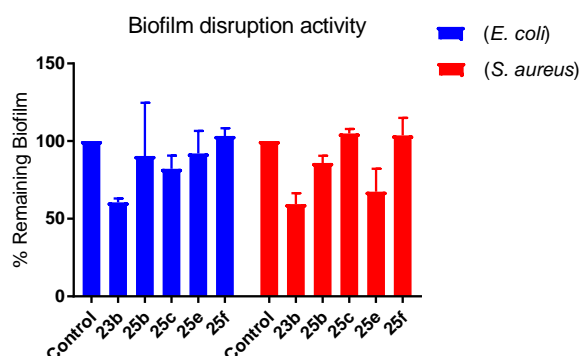
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Fig. 5. (A-F) *S. aureus* cytoplasmic membrane disruption promoted by 23b, 23d, 25b, 25e, 25f, 25g. (G) *E. coli* cytoplasmic membrane disruption promoted by 25g. Error bars represent the standard error of triplicates (n = 3).

2.6 Antibiofilm activity

The ability of the amphiphilic biphenyl derivatives to inhibit established *S. aureus* or *E. coli* biofilms was measured at 250 μM (Fig. 6). In general, the compounds showed low level of inhibition of biofilm formation. Compound 23b showed the highest level of disruption against *S. aureus* and *E. coli* of 41% and 39% respectively at 250 μM. Meanwhile, compound 25e displayed 30% inhibition against *S. aureus* biofilms but only 8% activity against biofilms of *E. coli*. The pre-established biofilms are harder to eradicate and have limited data available on novel AMPs with anti-biofilm properties [60]. Compound 23b containing less hydrophobic and cationic groups moderately disrupted both the *S. aureus* or *E. coli* biofilms compared to active antibacterial compounds. These results revealed that the increase in cationic charge and hydrophobicity elevates the activity against planktonic cells, but they are unable to disrupt the large aggregates of bacteria surrounded by an extracellular matrix. Previous studies have used a similar method for measuring the antibiofilm properties of cationic peptides, that is allowing the biofilm to form in the presence of the peptide [61]. Cirioni et al.[61] found that at a concentration of half the MIC, MSI-78 (pexiganan) was able to prevent biofilm formation by *P. aeruginosa* by 27.5%. This is higher than the activity of the compounds in the current study

343 which showed 39% inhibition of biofilm production by another Gram-negative bacterium *E. coli* but at a concentration of 31 times
344 the MIC. In an alternative biofilm study, where biofilms were formed prior to the addition of MSI-78 (pexiganan) was unable to
345 minimize the amount of biofilm produced by with *P. aeruginosa*, *E. coli* or *S. aureus* even at a concentration of 128 times its
346 MIC. However, in a similar preformed biofilm assay, another naturally occurring peptide, LL-37 which is also a membrane
347 disrupting peptide [62], reduced the biofilm mass by approximately 40% at 4 times its MIC [63]. Thus, the current compounds
348 show similar antibiofilm capabilities to cationic antimicrobial peptides, but future studies should examine whether the current
349 compounds are able to disrupt preformed biofilms.



350 **Fig. 6.** Percentage of remaining biofilm of *S. aureus* and *E. coli* after 24 h treatment with the synthesized compounds at 250 μM . The control represents the pre-
351 established biofilms without any compounds. Error bars represent the standard error of triplicates ($n = 3$).
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353 3. Conclusion

354 In conclusion, we have developed novel peptidomimetics based on the 3,3'-biphenyl structural scaffold. The systematic tuning of
355 hydrophobicity and cationic charge of the peptidomimetics resulted in moderate to excellent antibacterial activities. **25g**, the
356 amphipathic peptidomimetic compound in which the hydrophobic Trp-Phe and cationic Arg-aminoethylguanidine segregated by
357 biphenyl backbone showed excellent antibacterial activity against *S. aureus* (**15.6 μM**) and *E. coli* (**7.8 μM**) without cytotoxicity
358 against mammalian cells. Based on the results of cytoplasmic permeability assay and tBLMs-AC impedance spectroscopy, we
359 propose that the biphenyl peptidomimetics exhibit bacterial cell membrane disruption mechanism similar to most AMPs. The
360 importance of tryptophan in mechanism of action was also revealed by compound **29**. In addition, **23b** biphenyl attached with
361 hydrophobic Trp and cationic aminoethyl guanidine could disrupt biofilms of *S. aureus* and *E. coli* at 250 μM . Collectively, our
362 results suggest biphenyl is a versatile core about which hydrophobic and cationic amino acids can be arranged in order to generate
363 short cationic amphipathic peptidomimetics that mimic natural AMPs.
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367 4. Experimental section

368 4.1 General notes

369 All chemical reagents were purchased from commercial sources (Combi-Blocks, Chem-Impex and Sigma Aldrich) and used
370 without further purification. Solvents were commercial and used as obtained. Reactions were performed using oven-dried
371 glassware under an atmosphere of nitrogen and in anhydrous conditions (as required). Room temperature refers to the ambient
372 temperature. Yields refer to chromatographically and spectroscopically pure compounds unless otherwise stated. Reactions were
373 monitored by thin layer chromatography (TLC) plates pre-coated with Merck silica gel 60 F254. Visualization was accomplished
374 with UV light, a ninhydrin staining solution in n-butanol. Flash chromatography and silica pipette plugs were performed under
375 positive air pressure using Silica Gel 60 of 230–400 mesh (40–63 μm) and also using Grace Davison LC60A 6- μm for reverse
376 phase chromatography. Infrared spectra were recorded using a Cary 630 ATR spectrophotometer. **Melting points were obtained**
377 **using a OptiMelt melting point apparatus and are uncorrected.** High-resolution mass spectrometry was performed by the
378 Bioanalytical Mass Spectrometry facility, UNSW. Proton and Carbon NMR spectra were recorded in the solvents specified using
379 a Bruker DPX 300 or a Bruker Avance 400 or 600 MHz spectrometer as designated. Chemical shifts (δ) are quoted in parts per
380 million (ppm), to the nearest 0.01 ppm and internally referenced relative to the solvent nuclei. ¹HNMR spectral data are reported
381 as follows [chemical shift in ppm; multiplicity in br, broad; s, singlet; d, doublet; t, triplet; q, quartet; quint, quintet; sext, sextet;
382 sept, septet; m, multiplet; or as a combination of these (*e.g.* dd, dt *etc.*)]; coupling constant (*J*) in hertz, integration, proton count
383 and assignment.
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387 4.2 General methods

388 4.2.1 Procedure 1: peptide formation method a

389 To a stirred solution of an acid (1 equiv), amine (1.0 equiv), HOBt (1.0 equiv), DIEA (2.5 equiv) in DMF (3 – 5 mL) EDCI (1.2
390 equiv) was added portion-wise. The reaction was stirred overnight before the solvent was removed under reduced pressure and the
391 resultant residue subjected to flash chromatography (2-5% MeOH/CH₂Cl₂ as the eluent) to afford the desired compound.
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395 4.2.1 Procedure 2: peptide formation method b

To a stirred solution of an acid (1 equiv), amine (1.0 equiv), HOBt (1.0 equiv), PyBOP (1.1 equiv) in CH₂Cl₂ (3 – 5 mL) DIEA (2.0 equiv) was added dropwise. The reaction was stirred overnight before the solvent was removed under reduced pressure and the resultant residue subjected to flash chromatography (2-5% MeOH/CH₂Cl₂ as the eluent) to afford the desired compound.

4.2.2 Procedure 3: *N*-fmoc deprotection

To a stirred solution of the Fmoc-protected peptide in DMF (3.0 – 5.0 mL) was added piperidine (2.0 equiv). The resultant solution was then stirred at rt for overnight. The solvent was then removed under reduced pressure and purified using flash chromatography (5% of 100:10:1; CHCl₃:MeOH: aqueous NH₃ and 95% of CH₂Cl₂) as eluent. The resultant compounds isolated as TFA salt.

4.2.3 Procedure 4: *N*-boc and *Pbf* deprotection

To a stirred solution of the boc/Pbf protected peptide in CH₂Cl₂ (3.0 mL) was added TFA (3.0 mL). The reaction mixture was stirred at room temperature overnight before the solvent was removed under reduced pressure. After triturating with diethyl ether, the residue was concentrated to dryness. For Pbf and deboc of guanidine the residue was dissolved in CH₂Cl₂ and CH₃CN and precipitated by addition of diethyl ether and filtered and dried. Some of the compounds were purified using reverse phase chromatography with 40% CH₃CN/ H₂O as eluent utilizing GRACE instrument.

4.3 Preparation of derivatives

4.3.1. ¹H NMR and ¹³C NMR spectra of Methyl 3'-nitro-[1,1'-biphenyl]-3-carboxylate (9)

1-Bromo-3-nitrobenzene **7** (5.0 g, 24.7 mmol), (3-(Methoxycarbonyl)phenyl)boronic acid **8** (5.34 g, 29.7 mmol), tetrakis(triphenyl)phosphinepalladium(0) (1.40 g, 1.23 mmol), sodium carbonate (2 M) (37 mL, 74.1 mmol), and 1,4-dioxane (120 mL) were combined and the reaction mixture was heated at reflux under a nitrogen atmosphere for 16 h. The reaction mixture was allowed to cool at room temperature and partitioned between ethyl acetate and water. The organic phase was separated, dried over magnesium sulfate, filtered, and the filtrate was concentrated to give a liquid. The crude product was purified by flash chromatography over silica with a hexanes:ethyl acetate gradient (100:0 to 90:10) to give (4.12 g, 65%) the title compound **9** as a yellow solid; **m.p.**: 96.5–96.6 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.45 (t, *J* = 4.00 Hz, 1H), 8.27-8.24 (m, 2H), 8.19 (dd, *J* = 4.00, 8.00 Hz, 1H), 8.08-8.01 (m, 2H), 7.79 (t, *J* = 8.00 Hz, 1H), 7.68 (t, *J* = 8.00 Hz, 1H), 3.90 (s, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 166.4, 148.9, 141.0, 138.8, 133.9, 132.3, 131.1, 131.0, 130.2, 129.6, 127.9, 123.1, 121.8, 52.8; IR (ATR): ν_{max} 3312, 2953, 2342, 1722, 1520, 1429, 1348, 1301, 1236, 1191, 1104, 963, 883, 839, 801, 690; HRMS (ESI): *m/z* calcd for C₁₄H₁₁NO₄ Na [M + Na]⁺: 280.0586; found: 280.0579.

4.3.2 ¹H NMR and ¹³C NMR spectra of Methyl 3'-amino-[1,1'-biphenyl]-3-carboxylate (10)

To a stirred solution of 3'-nitro-[1,1'-biphenyl]-3-carboxylic acid methyl ester **9** (4.0 g, 15.5 mmol) in anhydrous THF (100 mL) under nitrogen atmosphere 10% palladium on activated charcoal (1.0 g) was added. The reaction was evacuated and placed under a hydrogen atmosphere and stirred overnight. The reaction mixture was filtered through Celite, and the solvent was removed under reduced pressure to yield gray oil. The residue was chromatographed on silica, eluting with 3:1 hexane/EtOAc. Concentration of the appropriate fractions provided the product as an off-white solid (3.17 g, 90% yield); **m.p.**: 84.9–85.1 °C; ¹H NMR (400 MHz, CDCl₃): δ 8.28 (t, *J* = 4.00 Hz, 1H), 8.03 (dt, *J* = 2.00, 5.00 Hz, 1H), 7.79-7.76 (m, 1H), 7.51 (t, *J* = 12.00 Hz, 1H), 7.27 (q, *J* = 4.00 Hz, 1H), 7.06-7.03 (m, 1H), 6.97 (t, *J* = 4.00 Hz, 1H), 6.75-6.73 (m, 1H), 3.96 (s, 5H); ¹³C NMR (100 MHz, CDCl₃): δ 167.1, 146.6, 141.6, 141.3, 131.5, 130.6, 129.9, 128.7, 128.3, 128.2, 117.7, 114.6, 113.9, 52.2; HRMS (ESI): *m/z* calcd for C₁₄H₁₃NO₂ Na [M + Na]⁺: 250.0844; found: 250.0838.

4.3.3. ¹H NMR and ¹³C NMR spectra of Methyl (*S*)-3'-(2-((*tert*-butoxycarbonyl)amino)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxylate (12)

The title compound was prepared *via* protocol 1, using **10** (1.0 g, 4.4 mmol) and (*tert*-butoxycarbonyl)-*L*-tryptophan (1.33 g, 4.4 mmol) to afford the coupled product **12** as an off-white solid (1.42 g, 63%); **m.p.**: 107.3–108.0 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.82 (s, 1H), 10.17 (s, 1H), 8.17 (s, 1H), 7.98-7.91 (m, 3H), 7.69-7.62 (m, 3H), 7.45-7.38 (m, 2H), 7.32 (d, *J* = 8.00 Hz, 1H), 7.19 (s, 1H), 7.05 (t, *J* = 8.00 Hz, 1H), 6.99-6.97 (m, 2H), 4.41-4.39 (m, 1H), 3.90 (s, 3H), 3.18-2.99 (m, 2H), 1.34 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 171.8, 166.5, 155.8, 140.9, 140.2, 139.9, 136.5, 131.9, 130.8, 130.8, 128.7, 127.7, 127.4, 124.2, 122.1, 121.3, 119.4, 118.6, 118.1, 111.8, 111.8, 110.4, 78.6, 56.4, 52.8, 28.6, 28.3; IR (ATR): ν_{max} 3304, 2330, 2099, 1665, 1605, 1528, 1491, 1423, 1303, 1250, 1160, 1111, 1082, 1010, 856, 733, 690; HRMS (ESI): *m/z* calcd for C₃₀H₃₁N₃O₅ Na [M + Na]⁺: 536.2161; found: 536.2153.

4.3.4. ¹H NMR and ¹³C NMR spectra of (*S*)-3'-(2-((*tert*-butoxycarbonyl)amino)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxylic acid (13)

To a solution of **12** (1.2 g, 1.81 mmol) in THF (10.0 mL) and MeOH (10.0 mL), was added a 1N NaOH_(aq) (3.63 mL, 3.63 mmol) and stirred at room temperature for 16 h. Ethyl acetate was added and the layers were separated. The aqueous layer was then

acidified with 1N HCl and then extracted with CH₂Cl₂ (2 X 100 mL) and then the solvent was removed under reduced pressure to yield **13** (1.00 g, 85%) as an off-white solid; **m.p.:** 140–140.3 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 13.10 (s, 1H), 10.82 (s, 1H), 10.17 (s, 1H), 8.18 (t, *J* = 4.00 Hz, 1H), 7.97–7.95 (m, 2H), 7.90–7.87 (m, 1H), 7.69–7.66 (m, 2H), 7.61 (t, *J* = 8.00 Hz, 1H), 7.45–7.38 (m, 2H), 7.32 (d, *J* = 8.00 Hz, 1H), 7.19 (s, 1H), 7.06 (t, *J* = 4.00 Hz, 1H), 6.98 (t, *J* = 8.00 Hz, 2H), 4.41 (q, *J* = 8.00 Hz, 1H), 3.19–3.14 (m, 1H), 3.05–3.00 (m, 1H), 1.34 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 171.7, 167.6, 155.8, 140.7, 140.2, 140.1, 136.5, 132.0, 129.9, 128.8, 127.8, 127.6, 124.3, 122.1, 121.4, 119.3, 119.1, 118.7, 118.1, 111.8, 110.4, 79.6, 78.6, 56.3, 28.6, 28.3; IR (ATR).vmax 3278, 3051, 2946, 2342, 2117, 1917, 1712, 1661, 1589, 1524, 1432, 1310, 1252, 1109, 973, 876, 790, 690; HRMS (ESI): *m/z* calcd for C₂₉H₂₉N₃O₅ Na [M + Na]⁺ 522.2005; found: 522.1996.

4.3.5. ¹H NMR and ¹³C NMR spectra of Methyl (*S*)-3'-(2-amino-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxylate (**14**)

The title compound **14** was prepared *via* protocol 4 using **12** (1.4 g, 2.72 mmol) to yield an off-white solid (1.00 g, 89%); **m.p.:** 86.4–86.7 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 11.06 (s, 1H), 10.73 (s, 1H), 8.33 (br s, 2H), 8.16–8.15 (m, 1H), 8.00–7.97 (m, 1H), 7.92–7.89 (m, 1H), 7.86–7.85 (m, 1H), 7.68–7.60 (m, 3H), 7.48–7.46 (m, 2H), 7.36 (d, *J* = 8.00 Hz, 1H), 7.26 (d, *J* = 4.00 Hz, 1H), 7.09–7.05 (m, 1H), 6.98–6.94 (m, 1H), 4.21 (br s, 1H), 3.89 (s, 3H), 3.40–3.25 (m, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 167.9, 166.6, 140.7, 140.1, 139.2, 136.7, 131.9, 130.9, 130.3, 130.1, 128.8, 127.5, 127.4, 125.4, 123.0, 121.7, 119.7, 119.0, 118.9, 118.4, 111.9, 107.1, 54.1, 52.8, 27.8; IR (ATR): vmax 3282, 3054, 2947, 2341, 2106, 1911, 1713, 1663, 1589, 1525, 1433, 1310, 1253, 1110, 974, 877, 791, 691; HRMS (ESI): *m/z* calcd for C₂₅H₂₃N₃O₃ [M + H]⁺: 413.1739; found 414.1810.

4.3.6. ¹H NMR and ¹³C NMR spectra of Methyl 3'-((*S*)-2-((*S*)-2-((*tert*-butoxycarbonyl)amino)-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxylate (**16**)

The title compound was prepared *via* protocol 1 using **14** and **15** (1.0 g, 3.76 mmol) to afford the coupled product **16** as an off-white solid; **m.p.:** 116.3–117.2 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.85 (s, 1H), 10.20 (s, 1H), 8.16–8.14 (m, 2H), 7.98–7.95 (m, 1H), 7.90–7.88 (m, 2H), 7.66–7.62 (m, 3H), 7.44–7.38 (m, 2H), 7.32 (d, *J* = 8.00 Hz, 1H), 7.21–7.11 (m, 6H), 7.07–7.03 (m, 1H), 6.95 (dd, *J* = 8.00, 14.00 Hz, 2H), 4.75 (q, *J* = 8.00 Hz, 1H), 4.22–4.16 (m, 1H), 3.89 (s, 3H), 3.24 (dd, *J* = 8.00, 16.00 Hz, 1H), 3.12 (dd, *J* = 8.00, 14.00 Hz, 1H), 2.93 (dd, *J* = 4.00, 12.00 Hz, 1H), 2.75–2.69 (m, 1H), 1.28 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 171.9, 170.9, 166.6, 155.7, 140.9, 139.9, 138.4, 136.5, 131.8, 130.8, 130.1, 130.0, 129.7, 128.7, 128.4, 127.8, 127.4, 126.6, 124.1, 122.3, 121.4, 119.4, 119.0, 118.7, 118.1, 111.7, 110.0, 78.7, 56.4, 54.7, 52.8, 37.9, 28.6, 28.4; IR (ATR).vmax 3293, 3055, 2320, 2096, 1649, 1492, 1433, 1310, 1250, 1259, 1110, 1012, 882; HRMS (ESI): *m/z* calcd for C₃₉H₄₀N₄O₆ [M + H]⁺: 683.2846; found: 683.2841.

4.3.7. ¹H NMR and ¹³C NMR spectra of 3'-((*S*)-2-((*S*)-2-((*tert*-butoxycarbonyl)amino)-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxylic acid (**17**)

To a solution of **16** (1.2 g, 2.33 mmol) in THF (10.0 mL) and MeOH (10.0 mL), was added a 1N NaOH_(aq) (4.66 mL, 4.66 mmol) and stirred at room temperature for 16 h. Ethyl acetate was added and the layers were separated. The aqueous layer was then acidified with 1N HCl and then extracted with CH₂Cl₂ (2 X 100 mL) and then the solvent was removed under reduced pressure to yield **17** (1.05 g, 90%) as an off-white solid; **m.p.:** 172.6–173.6 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 13.08 (br s, 1H), 10.84 (s, 1H), 10.20 (s, 1H), 8.16–8.14 (m, 2H), 7.96–7.85 (m, 3H), 7.69–7.58 (m, 3H), 7.44–7.38 (m, 2H), 7.32 (d, *J* = 4.00 Hz, 1H), 7.21–7.10 (m, 6H), 7.08–7.03 (m, 1H), 6.96 (dd, *J* = 8.00, 12.00 Hz, 2H), 4.75 (q, *J* = 8.00 Hz, 1H), 4.21–4.16 (m, 1H), 3.26–3.21 (m, 1H), 3.14–3.10 (m, 1H), 2.95–2.90 (m, 1H), 2.75–2.69 (m, 1H), 1.28 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 171.9, 170.8, 167.7, 155.7, 140.7, 140.1, 140.0, 138.4, 136.5, 132.0, 131.4, 130.0, 138.4, 136.5, 132.0, 131.3, 130.0, 129.8, 129.6, 128.8, 128.4, 127.8, 127.6, 126.6, 126.6, 124.1, 122.2, 121.4, 119.3, 119.0, 118.7, 118.1, 111.7, 110.0, 78.7, 56.4, 54.7, 37.9, 28.6, 28.4; IR (ATR).vmax 2048, 1653, 1492, 1434, 1391, 1228, 1158, 1058, 849; HRMS (ESI): *m/z* calcd for C₃₈H₃₈N₄O₆Na [M + Na]⁺: 669.2689; found: 669.2689.

4.3.8. ¹H NMR and ¹³C NMR spectra of 1-(2-aminoethyl)2,3-Bis(*tert*-butoxycarbonyl)guanidine (**18b**)

The title compound **18b** was prepared by adding the solution of N,N'-bis-(Boc)-1*H*-Pyrazole-1-carboxamide (2 g, 6.45 mmol) in THF (50 ml) dropwise to ethylene diamine (4 ml, 59.6 mmol) in THF (100 ml). After 30 min of mixing at room temperature solvent was evaporated then toluene (100 ml) was added and evaporated in order to remove remaining traces of the ethylene diamine. The crude reaction mixture taken for next step due to the instability of the compound. ¹H NMR (400 MHz, CDCl₃): δ 11.50 (br s, 1H), 8.63 (br s, 1H), 3.47 (q, *J* = 4.00 Hz, 2H), 2.87 (t, *J* = 4.00 Hz, 2H), 1.49 (s, 9H), 1.48 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 163.6, 156.5, 153.2, 83.1, 79.3, 43.4, 41.0, 28.3, 28.1; IR (ATR).vmax 3385, 3068, 2972, 2887, 2807, 1613, 1519, 1463, 1347, 1311, 1249, 1090, 1161, 884, 799.

4.3.9. ¹H NMR and ¹³C NMR spectra of Methyl N^w-((4-methoxy-2,3,6-trimethylphenyl)sulfonyl)-L-argininate (**18d**)

The title compound **18d** was prepared *via* protocol 3 using **20a** (0.9 g, 1.44 mmol) to yield the desired product as an off-white solid (450 mg, 77%); **m.p.:** 136.3–137.3 °C; ¹H NMR (400 MHz, CDCl₃): δ 6.50 (s, 1H), 6.46 (br s, 1H), 6.38 (br s, 2H), 3.80 (s, 3H), 3.67 (s, 3H), 3.47–3.45 (m, 1H), 3.15 (br s, 2H), 2.65 (s, 3H), 2.97 (s, 3H), 2.10 (s, 3H), 1.74–1.55 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ 175.7, 158.4, 156.4, 138.4, 136.4, 133.5, 124.8, 111.7, 55.5, 53.7, 52.2, 40.7, 31.2, 25.5, 24.1, 18.3, 11.9; IR

(ATR).vmax 3428, 3329, 2934, 1733, 1541, 1459, 1398, 1246, 1303, 1098, 1016, 911, 802; HRMS (ESI): m/z calcd for C₁₇H₂₈N₄O₅S [M + H]⁺: 401.1780; found: 401.1851.

4.3.10. ¹H NMR and ¹³C NMR spectra of tert-butyl (S)-(5-amino-6-((2-((tert-butoxycarbonyl)amino)ethyl)amino)-6-oxohexyl)carbamate (18e)

The title compound was prepared via protocol 3, using **20a** (630 mg, 1.03 mmol) to yield the desired product **18e** as an off-white solid (0.32 g, 80%); m.p.: 206.2–207.5 °C; ¹H NMR (600 MHz, CDCl₃): δ 7.63 (s, 1H), 5.09 (s, 1H), 4.67 (s, 1H), 3.34–3.08 (m, 6H), 1.84–1.76 (m, 1H), 1.50–1.46 (m, 2H), 1.42 (s, 18H), 1.37–1.36 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 175.6, 156.5, 156.1, 79.4, 79.1, 55.0, 40.6, 40.1, 39.7, 34.4, 29.8, 28.4, 28.4, 22.8; IR (ATR).vmax 3313, 2930, 1683, 1518, 1452, 1246, 1162, 994, 859, 779; HRMS (ESI): m/z calcd for C₁₈H₃₆N₄O₅ [M + H]⁺: 389.2686; found: 389.2759.

4.3.11. ¹H NMR and ¹³C NMR spectra of tert-butyl (S)-(5-amino-6-((2-(2,3-dibocguanidino)ethyl)amino)-6-oxohexyl)carbamate (18f)

The title compound was prepared via protocol 3, using **20b** (850 mg, 1.07 mmol) to yield the desired product **18f** as an off-white solid (0.45 g, 79%); m.p.: 184.4–185.6 °C; ¹H NMR (400 MHz, CDCl₃): δ 11.43 (s, 1H), 8.56 (t, J = 4.00 Hz, 1H), 7.91 (t, J = 8.00 Hz, 1H), 4.59 (br s, 1H), 3.59–3.55 (m, 2H), 3.45–3.41 (m, 2H), 3.33–3.30 (m, 1H), 3.13–3.08 (m, 2H), 1.82–1.77 (m, 1H), 1.52–1.51 (m, 1H), 1.49 (s, 18H), 1.49–1.45 (m, 4H), 1.43 (s, 9H), 1.39–1.37 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 175.6, 163.2, 157.1, 156.0, 153.1, 83.4, 79.5, 55.4, 40.2, 40.1, 40.0, 34.8, 30.0, 34.8, 30.0, 28.4, 28.3, 28.1, 23.0; IR (ATR).vmax 3326, 2931, 1714, 1612, 1521, 1409, 1362, 1324, 1246, 1128, 1047, 854; HRMS (ESI): m/z calcd for C₂₄H₄₆N₆O₇ [M + H]⁺: 531.3428; found: 531.3500.

4.3.12. ¹H NMR and ¹³C NMR spectra of (S)-2-amino-N-(2-(2,3-dibocguanidino)ethyl)-5-(3-((4-methoxy-2,3,6-trimethylphenyl)sulfonyl)guanidino)pentanamide (18g)

The title compound was prepared via protocol 3, using **21b** (730 mg, 0.82 mmol) to yield the desired product **18g** as an off-white solid (0.48 g, 87%); m.p.: 102.2–103.6 °C; ¹H NMR (400 MHz, CDCl₃): δ 11.39 (br s, 1H), 8.56 (br s, 1H), 8.04 (t, J = 4.00 Hz, 1H), 6.51 (s, 1H), 6.26 (br s, 3H), 3.81 (s, 3H), 3.54–3.39 (m, 5H), 3.24–3.20 (m, 2H), 2.67 (s, 3H), 2.60 (s, 3H), 2.11 (s, 3H), 1.76–1.61 (m, 4H), 1.46 (s, 9H), 1.45 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 175.1, 163.0, 158.4, 157.1, 156.4, 153.0, 138.5, 136.6, 133.6, 124.7, 111.7, 83.6, 79.8, 55.4, 54.3, 40.1, 31.7, 28.3, 28.0, 25.2, 24.1, 18.3, 11.9; IR (ATR).vmax 3326, 1719, 1611, 1543, 1439, 1409, 1326, 1248, 1115, 1046, 839, 803; HRMS (ESI): m/z calcd for C₂₉H₅₀N₈O₈S [M + H]⁺: 671.3540; found: 671.3540.

4.3.13. ¹H NMR and ¹³C NMR spectra of Methyl N²-(((9H-fluoren-9-yl)methoxy)carbonyl)-N^ω-((4-methoxy-2,3,6-trimethylphenyl)sulfonyl)-L-argininate (20')

The title compound **20'** was prepared by adding thionyl chloride (0.23 mL, 3.28 mmol) dropwise to N_α-Fmoc-N_ω-MTR-L-arginine (1.0 g, 1.64 mmol) in methanol (20.0 mL) at 0° C and then stirred at room temperature for 16 h. The solvents were removed and reduced pressure and the residue was diluted with ethylacetate (100.0 mL) and washed with saturated NaHCO₃ (40 mL), saturated brine (40 mL), then dried (Na₂SO₄) and concentrated under reduced pressure to afford a white solid (0.95 g, 93%); m.p.: 94.0–94.4 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.76–7.35 (m, 7H), 7.30–7.28 (m, 2H), 6.50 (s, 1H), 6.20 (br s, 2H), 5.74–5.72 (br d, J = 7.2 Hz), 4.38–4.29 (m, 2H), 4.19–4.14 (m, 2H), 3.79 (s, 3H), 3.71 (s, 3H), 3.22–3.21 (m, 2H), 2.69 (s, 3H), 2.62 (s, 3H), 2.11 (s, 3H), 1.70–1.57 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ 172.7, 158.5, 156.3, 143.7, 143.6, 141.2, 138.5, 136.5, 133.5, 127.8, 127.1, 125.1, 124.8, 120.0, 111.7, 67.13, 55.4, 52.6, 47.1, 40.7, 30.1, 25.2, 24.1, 18.3, 17.9, 11.9; IR (ATR).vmax 2946, 1717, 1542, 1616, 1445, 1244, 1169, 1102; HRMS (ESI): m/z calcd for C₃₂H₃₈N₄O₇SNa [M + Na]⁺: 645.2359; found: 645.2356.

4.3.14. ¹H NMR and ¹³C NMR spectra of (9H-fluoren-9-yl)methyl (S)-1-((2-(2,3-diboc guanidino)ethyl)amino)-5-(3-((4-methoxy-2,3,6-trimethylphenyl)sulfonyl)guanidino)-1-oxopentan-2-yl)carbamate (20b)

The title compound **20b** was prepared from compound **18b** (0.5 g, 1.64 mmol) and N_α-Fmoc-N_ω-MTR-L-arginine (1.0 g, 1.64 mmol) according to the protocol 2. Off-white solid (0.73 g, 50%); m.p.: 148.0–148.6 °C; ¹H NMR (400 MHz, CDCl₃): δ 11.40 (br s, 1H), 8.61 (br s, 1H), 7.98 (br s, 1H), 7.73 (d, J = 8.00 Hz, 2H), 7.57 (d, J = 8.00 Hz, 2H), 7.36 (t, J = 8.00 Hz, 2H), 7.30–7.24 (m, 2H), 6.50 (br s, 1H), 6.23 (br s, 2H), 5.99 (d, J = 8.00 Hz, 1H), 4.34–4.22 (m, 2H), 4.18–4.14 (m, 2H), 3.79 (s, 3H), 3.54–3.15 (m, 7H), 2.67 (s, 3H), 2.60 (s, 3H), 2.10 (s, 3H), 1.84–1.57 (m, 4H), 1.45 (s, 18H); ¹³C NMR (100 MHz, CDCl₃): δ 163.3, 162.8, 159.7, 158.5, 157.2, 156.4, 156.3, 153.0, 143.8, 143.8, 141.3, 138.6, 136.6, 133.4, 127.7, 127.1, 125.2, 124.8, 120.0, 83.6, 80.4, 80.1, 67.1, 55.4, 47.1, 42.1, 40.1, 28.2, 28.2, 28.0, 25.2, 24.1, 18.4, 12.0; IR (ATR).vmax 3324, 2976, 2064, 1717, 1615, 1543, 1446, 1304, 1248, 1117, 840; HRMS (ESI): m/z calcd for C₄₄H₆₀N₈O₁₀S [M + H]⁺: 893.4153; found: 893.4224.

4.3.15. ¹H NMR and ¹³C NMR spectra of (9H-fluoren-9-yl)methyl tert-butyl (6-((2-((tert-butoxycarbonyl)amino)ethyl)amino)-6-oxohexane-1,5-diyl)(S)-dicarbamate (21a)

The title compound **21a** was prepared from compound **18a** (0.34 g, 2.13 mmol) and N_α-Fmoc-N_ε-Boc-L-lysine (1.0 g, 2.13 mmol) according to the protocol 2. Off-white solid (0.65 g, 50%); m.p.: 173.7–175.2 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 7.90–7.87 (m, 3H), 7.72 (t, J = 6.00 Hz, 2H), 7.42–7.38 (m, 3H), 7.32 (t, J = 6.00 Hz, 2H), 6.73 (d, J = 6.00 Hz, 2H), 4.28–4.27 (m, 1H),

4.24-4.20 (m, 2H), 3.88-3.87 (m, 1H), 3.12-3.00 (m, 2H), 2.99-2.96 (m, 2H), 2.90-2.86 (m, 2H), 1.74-1.46 (m, 3H), 1.36 (s, 18H), 1.28-1.19 (m, 3H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.5, 156.4, 156.1, 156.0, 144.4, 144.2, 141.2, 128.1, 127.5, 125.7, 120.6, 79.6, 79.4, 79.2, 79.2, 78.2, 77.8, 66.1, 55.2, 47.1, 46.3, 46.3, 32.1, 29.7, 28.7, 28.6, 26.4, 26.3, 23.3; IR (ATR).vmax 3356, 2943, 2265, 2106, 1526, 1447, 1389, 1526, 1240, 1167, 1025, 836, 735; HRMS (ESI): m/z calcd for C₃₃H₄₆N₄O₇Na [M + Na]⁺: 633.3264; found: 633.3261.

4.3.16. ¹H NMR and ¹³C NMR spectra of (9H-fluoren-9-yl)methyl tert-butyl (6-((2-(2,3-dibocguanidino)ethyl)amino)-6-oxohexane-1,5-diyl)(*S*)-dicarbamate (21b)

The title compound **21b** was prepared from compound **18b** (0.643 g, 2.13 mmol) and *N*_α-Fmoc-*N*_ε-Boc-*L*-lysine (1.0 g, 2.13 mmol) according to the protocol 2. Off-white solid (0.89 g, 55%); m.p.: 102.2–103.6 °C; ¹H NMR (400 MHz, CDCl₃): δ 11.43 (s, 1H), 8.65 (br s, 1H), 8.07 (br s, 1H), 7.75 (d, *J* = 8.00 Hz, 2H), 7.59 (d, *J* = 8.00 Hz, 2H), 7.39 (t, *J* = 8.00 Hz, 2H), 7.33-7.26 (m, 2H), 5.65 (d, *J* = 4.00 Hz, 1H), 4.59 (br s, 1H), 4.38-4.36 (m, 1H), 4.22-4.17 (m, 1H), 3.57-3.53 (m, 2H), 3.45-3.42 (m, 2H), 3.10-3.08 (m, 2H), 1.87-1.80 (m, 1H), 1.70-1.66 (m, 1H), 1.50 (s, 9H), 1.48 (s, 9H), 1.42 (s, 9H), 1.48-1.32 (m, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 171.6, 162.8, 157.6, 156.0, 155.9, 153.0, 143.9, 141.3, 127.7, 127.0, 125.2, 120.0, 83.7, 80.0, 79.1, 66.9, 54.8, 47.2, 41.5, 40.3, 32.9, 29.8, 28.4, 28.3, 28.0, 22.3; IR (ATR).vmax 3316, 2973, 1713, 1612, 1516, 1448, 1410, 1325, 1244, 1129, 1046, 857; HRMS (ESI): m/z calcd for C₃₉H₅₆N₆O₉Na [M + Na]⁺: 775.4006; found: 775.4006.

4.3.17. ¹H NMR and ¹³C NMR spectra of tert-butyl (*S*)-(1-((3'-((2-(tert-butoxycarbonyl)amino)ethyl)carbamoyle)-[1,1'-biphenyl]-3-yl)amino)-3-(1H-indol-3-yl)-1-oxopropan-2-yl)carbamate (22a)

The title compound was prepared from compound **13** (100 mg, 0.20 mmol) and **18a** (32 mg, 0.20 mmol) following the general protocol 1 to afford **22a** as an off-white solid (83 mg, 65%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.82 (br s, 1H), 10.20 (br s, 1H), 8.60 (t, *J* = 4.00 Hz, 1H), 8.10 (s, 1H), 7.90 (br s, 1H), 7.85 (d, *J* = 4.00 Hz, 1H), 7.75 (d, *J* = 8.00 Hz, 1H), 7.67 (d, *J* = 8.00 Hz, 2H), 7.57 (t, *J* = 8.00 Hz, 1H), 7.46-7.40 (m, 2H), 7.33 (d, *J* = 4.00 Hz, 1H), 7.20 (s, 1H), 7.08-6.92 (m, 4H), 4.44-4.40 (m, 1H), 3.18-2.99 (m, 6H), 1.37 (s, 18H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 166.7, 156.2, 140.6, 140.6, 136.5, 135.7, 129.8, 129.4, 127.8, 126.9, 125.9, 124.3, 122.3, 121.3, 119.2, 119.1, 118.7, 118.3, 111.8, 110.4, 79.6, 78.6, 78.2, 28.7, 28.6, 28.3; IR (ATR).vmax 3298, 2974, 1669, 1492, 1453, 1247, 1158, 1010, 853; HRMS (ESI): m/z calcd for C₃₆H₄₃N₅O₆Na [M + Na]⁺: 664.3111; found: 664.3102.

4.3.18. ¹H NMR and ¹³C NMR spectra of tert-butyl (*S*)-(1-((3'-((2-(2,3-dibocguanidino)ethyl)carbamoyle)-[1,1'-biphenyl]-3-yl)amino)-3-(1H-indol-3-yl)-1-oxopropan-2-yl)carbamate (22b)

The title compound was prepared from compound **13** (100 mg, 0.20 mmol) and **18b** (61 mg, 0.2 mmol) following the general protocol 1 to afford **22b** as an off-white solid (94 mg, 60%); m.p.: 127.2–127.6 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.49 (br s, 1H), 10.81 (br s, 1H), 10.15 (br s, 1H), 8.67 (t, *J* = 6.00 Hz, 1H), 8.47 (t, *J* = 6.00 Hz, 1H), 8.08 (br s, 1H), 7.91 (s, 1H), 7.82 (d, *J* = 12.00 Hz, 1H), 7.75 (d, *J* = 12.00 Hz, 1H), 7.67-7.66 (m, 2H), 7.56 (t, *J* = 6.00 Hz, 1H), 7.32 (d, *J* = 6.00 Hz, 1H), 7.19 (br s, 1H), 7.05 (t, *J* = 6.00 Hz, 1H), 6.99-6.95 (m, 2H), 4.40 (q, *J* = 6.00 Hz, 1H), 3.52 (q, *J* = 6.00 Hz, 2H), 3.46 (q, *J* = 6.00 Hz, 2H), 3.17-3.13 (m, 1H), 3.04-3.00 (m, 1H), 1.43 (s, 9H), 1.37 (s, 9H), 1.33 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 171.1, 166.9, 163.7, 163.5, 156.2, 155.7, 152.3, 140.6, 140.5, 140.1, 136.5, 135.6, 129.8, 129.7, 129.4, 129.4, 127.7, 126.7, 125.9, 124.2, 122.2, 121.3, 119.2, 119.0, 118.6, 118.2, 111.7, 110.4, 83.3, 78.6, 78.5, 67.4, 56.3, 40.5, 39.2, 31.7, 28.7, 28.6, 28.4, 28.2, 28; IR (ATR).vmax 2977, 1612, 1535, 1364, 1326, 1229, 1133, 875; HRMS (ESI): m/z calcd for C₄₂H₅₃N₇O₈ [M + H]⁺: 784.3956; found: 784.4031.

4.3.19. ¹H NMR and ¹³C NMR spectra of methyl *N*⁶-(tert-butoxycarbonyl)-*N*²-(3'-((*S*)-2-((tert-butoxycarbonyl)amino)-3-(1H-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carbonyl)-*L*-lysinate (22c)

The title compound was prepared from compound **13** (100 mg, 0.20 mmol) and **18c** (52 mg, 0.2 mmol) following the general protocol 1 to afford **22c** as an off-white solid (109 mg, 74%); m.p.: 235.5–236.1 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.82 (s, 1H), 10.17 (s, 1H), 8.84 (d, *J* = 8.00 Hz, 1H), 8.14 (s, 1H), 7.91-7.88 (m, 2H), 7.79 (d, *J* = 8.00 Hz, 1H), 7.68 (t, *J* = 4.00 Hz, 1H), 7.59 (t, *J* = 8.00 Hz, 1H), 7.47-7.41 (m, 2H), 7.32 (d, *J* = 8.00 Hz, 1H), 7.20 (s, 1H), 7.06 (t, *J* = 8.00 Hz, 1H), 7.00-6.97 (m, 2H), 6.78 (t, *J* = 8.00 Hz, 1H), 4.47-4.38 (m, 2H), 3.66 (s, 3H), 3.17-2.89 (m, 4H), 1.81 (t, *J* = 8.00 Hz, 1H), 1.41-1.37 (m, 2H), 1.34 (s, 18H), 1.24-1.18 (m, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 173.2, 166.9, 156.0, 155.7, 140.7, 140.5, 140.1, 136.5, 134.8, 130.0, 129.8, 129.4, 127.7, 127.1, 126.1, 124.2, 122.3, 121.3, 119.0, 118.6, 118.2, 111.7, 110.4, 79.6, 78.5, 77.8, 53.2, 52.3, 30.7, 29.5, 28.7, 28.6, 28.2, 23.5; IR (ATR).vmax 2929, 2312, 1992, 1671, 1521, 1433, 1363, 1228, 1160, 1010, 860; HRMS (ESI): m/z calcd for C₄₁H₅₁N₅O₈Na [M + Na]⁺: 764.3635; found: 764.3630.

4.3.20. ¹H NMR and ¹³C NMR spectra of Methyl *N*²-(3'-((*S*)-2-((tert-butoxycarbonyl)amino)-3-(1H-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carbonyl)-*N*^w-((4-methoxy-2,3,6-trimethylphenyl)sulfonyl)-*L*-argininate (22d)

The title compound was prepared from compound **13** (100 mg, 0.20 mmol) and **18d** (80 mg, 0.2 mmol) following the general protocol 1 to afford **22d** as an off-white solid (104 mg, 59%); m.p.: 172.2–173.1 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.82 (s, 1H), 10.17 (s, 1H), 8.88 (d, *J* = 12.00 Hz, 1H), 8.13 (br s, 1H), 7.92-7.88 (m, 2H), 7.79 (d, *J* = 12.00 Hz, 1H), 7.68-7.66 (m, 2H), 7.60 (t, *J* = 6.00 Hz, 1H), 7.47-7.41 (m, 2H), 7.33 (d, *J* = 12.00 Hz, 1H), 7.19 (br s, 1H), 7.08-7.04 (m, 1H), 7.00-6.96 (m, 2H), 6.74-6.66 (m, 2H), 6.48-6.26 (m, 2H), 4.48-4.40 (m, 2H), 3.77 (s, 3H), 3.65 (s, 3H), 3.15-2.99 (m, 4H), 2.59 (s, 3H), 2.51 (s, 3H),

2.02 (s, 3H), 1.82-1.74 (m, 2H), 1.52-1.49 (m, 2H), 1.34 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 173.0, 171.8, 166.9, 157.9, 156.6, 155.7, 140.7, 140.5, 140.1, 138.1, 136.5, 134.8, 130.1, 129.8, 129.5, 127.7, 127.1, 126.1, 124.3, 122.3, 121.3, 119.2, 119.1, 118.6, 118.3, 112.2, 111.8, 110.4, 79.6, 78.6, 56.3, 55.9, 53.0, 52.3, 28.6, 28.4, 24.1, 18.5, 12.2; IR (ATR).vmax 3305, 2932, 1669, 1541, 1456, 1364, 1303, 1227, 1159, 1114, 1013; HRMS (ESI): m/z calcd for C₄₆H₅₅N₇O₉S [M + H]⁺: 882.3782; found: 882.3854.

4.3.21. ¹H NMR and ¹³C NMR spectra of tert-butyl ((*S*)-5-(3'-((*S*)-2-((tert-butoxycarbonyl)amino)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxamido)-6-((2-((tert-butoxycarbonyl)amino)ethyl)amino)-6-oxohexyl)carbamate (**22e**)

The title compound was prepared from compound **13** (100 mg, 0.20 mmol) and **18e** (77 mg, 0.20 mmol) following the general protocol 1 to afford **22e** as an off-white solid (139 mg, 80%); m.p.: 131.5–131.7 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.82 (s, 1H), 10.16 (s, 1H), 8.55 (d, *J* = 6.00 Hz, 1H), 8.16 (s, 1H), 8.01 (t, *J* = 6.00 Hz, 1H), 7.92-7.91 (m, 2H), 7.77-7.75 (m, 1H), 7.68-7.67 (m, 1H), 7.57 (t, *J* = 12.00 Hz, 1H), 7.45-7.43 (m, 2H), 7.32 (d, *J* = 6.00 Hz, 1H), 7.20 (s, 1H), 7.06 (t, *J* = 6.00 Hz, 1H), 6.99-6.96 (m, 2H), 6.77-6.74 (m, 2H), 4.43-4.37 (m, 2H), 3.18-2.86 (m, 8H), 1.77-1.68 (m, 2H), 1.37-1.36 (m, 2H), 1.36 (s, 9H), 1.34 (s, 18H), 1.24-1.15 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.4, 171.8, 166.7, 156.1, 156.0, 155.8, 140.6, 140.6, 136.5, 135.3, 129.8, 129.3, 127.7, 127.2, 126.3, 124.3, 121.3, 119.2, 119.1, 118.7, 118.3, 111.7, 110.4, 78.6, 78.1, 77.7, 56.3, 54.1, 40.5, 39.3, 31.8, 31.4, 29.7, 23.7, 22.5; IR (ATR).vmax 2972, 2930, 1683, 1521, 1455, 1246, 1163, 1010, 861; HRMS (ESI): m/z calcd for C₄₇H₆₃N₇O₉Na [M + Na]⁺: 892.4585; found: 892.4575.

4.3.22. ¹H NMR and ¹³C NMR spectra of (*S*)-3'-(2-amino-3-(1*H*-indol-3-yl)propanamido)-*N*-(2-aminoethyl)-[1,1'-biphenyl]-3-carboxamide (**23a**)

The title compound was prepared from compound **22a** (75 mg, 0.116 mmol) following the general protocol 4 to afford **23a** as a white solid (45 mg, 89%); m.p.: 164.5–164.7 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.06 (d, *J* = 6.00 Hz, 1H), 10.70 (s, 1H), 8.78 (t, *J* = 6.00 Hz, 1H), 8.30 (br s, 3H), 8.13 (t, *J* = 6.00 Hz, 1H), 7.90-7.87 (m, 5H), 7.79-7.77 (m, 1H), 7.68 (d, *J* = 12.00 Hz, 1H), 7.62-7.60 (m, 2H), 7.50-7.49 (m, 2H), 7.37 (d, *J* = 12.00 Hz, 1H), 7.26 (d, *J* = 6.00 Hz, 1H), 7.10-7.07 (m, 1H), 6.99-6.97 (m, 1H), 4.21-4.20 (m, 1H), 3.55 (q, *J* = 6.00 Hz, 2H), 3.39-3.35 (m, 2H), 3.28-3.25 (m, 2H), 3.03 (q, *J* = 6.00 Hz, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 167.9, 167.2, 140.6, 140.4, 139.1, 136.7, 135.3, 130.1, 129.9, 129.6, 127.5, 127.1, 126.1, 125.4, 123.1, 121.7, 119.6, 119.0, 118.9, 118.5, 111.9, 107.1, 54.1, 40.5, 39.1, 37.6, 27.8; IR (ATR).vmax 3041, 2890, 1774, 1664, 1525, 1429, 1318, 1179, 835, 795, 744; HRMS (ESI): m/z calcd for C₂₆H₂₇N₅O₂ [M + H]⁺: 442.2165; found: 442.2234.

4.3.23. ¹H NMR and ¹³C NMR spectra of (*S*)-3'-(2-amino-3-(1*H*-indol-3-yl)propanamido)-*N*-(2-guanidinoethyl)-[1,1'-biphenyl]-3-carboxamide (**23b**)

The title compound was prepared from compound **22b** (90 mg, 0.122 mmol) following the general protocol 4 to afford **23b** as a white solid (29 mg, 50%); m.p.: 236.7–238.0 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.05 (br s, 1H), 10.69 (s, 1H), 8.77 (t, *J* = 6.00 Hz, 1H), 8.28 (br s, 3H), 8.12 (t, *J* = 6.00 Hz, 1H), 7.89-7.87 (m, 2H), 7.78-7.77 (m, 1H), 7.70-7.67 (m, 2H), 7.62-7.59 (m, 2H), 7.49-7.48 (m, 3H), 7.38-7.36 (m, 2H), 7.26 (d, *J* = 6.00 Hz, 2H), 7.10-7.07 (m, 1H), 6.99-6.97 (m, 1H), 4.20 (t, *J* = 6.00 Hz, 1H), 3.45 (q, *J* = 6.00 Hz, 2H), 3.39-3.36 (m, 1H), 3.28-3.24 (m, 1H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 167.9, 167.1, 157.5, 140.6, 140.4, 139.1, 135.3, 130.1, 129.8, 129.6, 127.5, 126.9, 126.0, 125.4, 123.1, 121.7, 119.6, 119.0, 118.9, 118.5, 112.0, 107.1, 54.1, 40.8, 39.1, 27.8; IR (ATR).vmax 3178, 3080, 2340, 2110, 1657, 1542, 1430, 1316, 1200, 1148, 835, 797, 743; HRMS (ESI): m/z calcd for C₂₇H₂₉N₇O₂ [M + H]⁺: 484.2383; found: 484.2455.

4.3.24. ¹H NMR and ¹³C NMR spectra of Methyl (3'-((*S*)-2-amino-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carbonyl)-*L*-lysinate (**23c**)

The title compound was prepared from compound **22c** (50 mg, 0.067 mmol) following the general protocol 4 to afford **23c** as a gummy solid (36 mg, 70%); ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.00 (s, 1H), 10.70 (s, 1H), 8.89 (d, *J* = 6.00 Hz, 1H), 8.29 (br s, 3H), 8.14-8.13 (m, 1H), 7.93-7.91 (m, 1H), 7.87-7.86 (m, 1H), 7.80-7.75 (m, 4H), 7.68 (d, *J* = 12.00 Hz, 1H), 7.62-7.60 (m, 2H), 7.50-7.48 (m, 2H), 7.37-7.36 (m, 1H), 7.26 (d, *J* = 6.00 Hz, 1H), 7.10-7.07 (m, 1H), 6.99-6.97 (m, 1H), 4.50-4.46 (m, 1H), 4.20 (br s, 1H), 3.67 (s, 3H), 3.35-3.25 (m, 2H), 2.80-2.79 (m, 2H), 1.86-1.39 (m, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 173.1, 167.9, 166.9, 158.6, 158.4, 140.6, 140.4, 139.1, 136.7, 134.8, 130.2, 130.1, 129.6, 127.5, 127.2, 126.2, 125.4, 123.2, 121.6, 119.6, 118.95, 118.9, 118.6, 111.9, 107.1, 54.1, 53.0, 52.4, 40.5, 40.4, 39.0, 30.4, 27.8, 27.0, 23.1; IR (ATR).vmax 3251, 3044, 2932, 2098, 1666, 1524, 1432, 1338, 1178, 1118, 835, 796, 743, 719; HRMS (ESI): m/z calcd for C₃₁H₃₅N₅O₄ [M + H]⁺: 542.2689; found: 542.2157.

4.3.25. ¹H NMR and ¹³C NMR spectra of Methyl (3'-((*S*)-2-amino-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carbonyl)-*L*-argininate (**23d**)

The title compound was prepared from compound **22d** (90 mg, 0.10 mmol) following the general protocol 4 to afford **23d** as a gummy solid (50 mg, 54%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.04 (s, 1H), 10.66 (s, 1H), 8.93 (d, *J* = 6.00 Hz, 1H), 8.27 (br s, 3H), 8.13 (br s, 1H), 7.92 (d, *J* = 6.00 Hz, 2H), 7.79 (d, *J* = 12.00 Hz, 1H), 7.73-7.45 (m, 6H), 7.38-7.25 (m, 2H), 7.10-6.97 (m, 3H), 4.52-4.49 (m, 1H), 4.19-4.18 (m, 1H), 3.68 (s, 3H), 3.35-3.12 (m, 4H), 1.92-1.54 (m, 4H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.9, 167.9, 166.9, 162.2, 158.7, 158.5, 157.2, 140.6, 140.4, 139.1, 136.7, 134.8, 130.2, 130.1, 129.6, 127.5, 127.2, 126.2, 125.4, 123.2, 121.7, 119.6, 118.9, 118.9, 118.8, 118.5, 112.0, 107.1, 54.1, 52.8, 52.5, 40.7, 40.5, 28.1, 27.8, 25.8; IR (ATR).vmax

, 3192, 3065, 2118, 1660, 1539, 1434, 1315, 1227, 1159, 836, 797, 744; HRMS (ESI): m/z calcd for C₃₁H₃₅N₇O₄ [M + H]⁺: 570.3114; found: 570.2820.

4.3.26. ¹H NMR and ¹³C NMR spectra of *N*-((*S*)-6-amino-1-((2-aminoethyl)amino)-1-oxohexan-2-yl)-3'-((*S*)-2-amino-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxamide (23e)

The title compound was prepared from compound **22c** (100 mg, 0.114 mmol) following the general protocol 4 to afford **23e** as a white solid (45 mg, 70%); **m.p.**: 237.6–238.0 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.05 (d, *J* = 4.00 Hz, 1H), 10.71 (s, 1H), 8.68 (d, *J* = 4.00 Hz, 1H), 8.29–8.23 (m, 4H), 8.16 (s, 1H), 7.94 (d, *J* = 4.00 Hz, 1H), 7.85 (br s, 4H), 7.77–7.75 (m, 4H), 7.67 (d, *J* = 8.00 Hz, 1H), 7.60–7.58 (m, 2H), 7.51–7.47 (m, 2H), 7.36 (d, *J* = 8.00 Hz, 1H), 7.25 (d, *J* = 4.00 Hz, 1H), 7.08 (t, *J* = 4.00 Hz, 1H), 6.97 (t, *J* = 4.00 Hz, 1H), 4.44–4.41 (m, 1H), 4.21–4.20 (m, 1H), 3.37–3.24 (m, 4H), 2.88–2.77 (m, 4H), 1.86–1.70 (m, 2H), 1.60–1.52 (m, 2H), 1.44–1.31 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.8, 167.9, 166.8, 158.9, 158.7, 158.5, 140.7, 140.3, 139.1, 136.7, 135.2, 130.0, 129.4, 127.5, 127.3, 126.3, 125.4, 123.2, 121.7, 119.6, 118.9, 118.6, 116.7, 111.9, 107.1, 54.1, 53.8, 40.5, 39.1, 38.9, 36.9, 31.2, 27.8, 27.2, 23.2; IR (ATR).vmax 3257, 3052, 2930, 2099, 1664, 1524, 1431, 1317, 1170, 797, 744, 720; HRMS (ESI): m/z calcd for C₃₂H₃₉N₇O₃ [M + H]⁺: 570.3114; found: 570.2820.

4.3.27. ¹H NMR and ¹³C NMR spectra of *tert*-butyl ((*S*)-1-(((*S*)-1-((3'-((2-((*tert*-butoxycarbonyl)amino)ethyl)carbamoyl)-[1,1'-biphenyl]-3-yl)amino)-3-(1*H*-indol-3-yl)-1-oxopropan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate (24a)

The title compound was prepared from compound **17** (100 mg, 0.15 mmol) and **18a** (25 mg, 0.15 mmol) following the general protocol 1 to afford **24a** as an off-white solid (94 mg, 80%); **m.p.**: 126.7–128.0 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.82 (s, 1H), 10.15 (s, 1H), 8.57–8.56 (m, 1H), 8.43 (d, *J* = 8.00 Hz, 1H), 8.10 (s, 1H), 7.81–7.67 (m, 7H), 7.52 (t, *J* = 8.00 Hz, 1H), 7.30 (d, *J* = 8.00 Hz, 1H), 7.17–6.91 (m, 9H), 6.76 (d, *J* = 8.00 Hz, 1H), 4.75 (q, *J* = 8.00 Hz, 1H), 4.26–4.21 (m, 1H), 3.29–3.03 (m, 6H), 2.73–2.72 (m, 1H), 2.60–2.54 (m, 1H), 1.36 (s, 9H), 1.29 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 171.9, 166.7, 160.0, 156.2, 155.7, 155.1, 140.0, 139.8, 138.9, 138.3, 137.1, 136.5, 135.6, 134.9, 129.7, 129.3, 129.3, 128.3, 127.7, 127.5, 126.5, 125.3, 124.2, 121.4, 120.2, 119.0, 118.7, 111.7, 110.1, 78.5, 78.1, 56.0, 54.5, 37.8, 28.7, 28.5, 28.2; IR (ATR).vmax 3299, 2976, 1663, 1518, 1455, 1246, 1160, 1118, 1021, 806; HRMS (ESI): m/z calcd for C₄₅H₅₂N₆O₇Na [M + Na]⁺: 811.3795; found: 811.3789.

4.3.28. ¹H NMR and ¹³C NMR spectra of *tert*-butyl ((*S*)-1-(((*S*)-1-((3'-((2-(2,3-dibocguanidino)ethyl)carbamoyl)-[1,1'-biphenyl]-3-yl)amino)-3-(1*H*-indol-3-yl)-1-oxopropan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate (24b)

The title compound was prepared from compound **17** (100 mg, 0.15 mmol) and **18b** (46 mg, 0.15 mmol) following the general protocol 1 to afford **24b** as an off-white solid (95 mg, 68%); **m.p.**: 206.3–206.5 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.50 (s, 1H), 10.80 (s, 1H), 10.20 (s, 1H), 8.41–8.45 (m, 1H), 8.14 (br s, 1H), 8.07 (br s, 1H), 7.88 (s, 1H), 7.82 (d, *J* = 4.00 Hz, 1H), 7.73 (d, *J* = 4.00 Hz, 1H), 7.62 (t, *J* = 8.00 Hz, 2H), 7.55 (t, *J* = 4.00 Hz, 1H), 7.42–7.40 (m, 2H), 7.32–7.31 (m, 1H), 7.20–7.13 (m, 7H), 7.05 (t, *J* = 4.00 Hz, 1H), 6.97–6.92 (m, 2H), 4.75–4.74 (m, 1H), 4.18–4.17 (m, 1H), 3.50 (t, *J* = 4.00 Hz, 2H), 3.45 (q, *J* = 4.00 Hz, 2H), 3.25–3.21 (m, 1H), 3.13–3.09 (m, 1H), 2.93–2.90 (m, 1H), 2.73–2.69 (m, 1H), 1.42 (s, 9H), 1.37 (s, 9H), 1.28 (s, 9H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 171.9, 170.8, 166.9, 163.6, 156.2, 155.7, 152.3, 140.6, 140.5, 139.9, 138.4, 136.5, 135.7, 129.8, 129.6, 129.4, 128.4, 127.8, 126.7, 126.6, 125.9, 124.1, 122.4, 121.4, 119.2, 118.9, 118.7, 118.3, 111.7, 110.0, 83.3, 78.6, 56.4, 54.6, 40.5, 39.2, 37.9, 31.7, 28.7, 28.6, 28.0; IR (ATR).vmax 2974, 1641, 1539, 1364, 1326, 1250, 1133, 1049, 876; HRMS (ESI): m/z calcd for C₅₁H₆₂N₈O₉Na [M + Na]⁺: 953.4537; found: 953.4533.

4.3.29. ¹H NMR and ¹³C NMR spectra of Methyl *N*⁶-(*tert*-butoxycarbonyl)-*N*²-(3'-((*S*)-2-((*S*)-2-((*tert*-butoxycarbonyl)amino)-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carbonyl)-*L*-lysinate (24c)

The title compound was prepared from compound **17** (100 mg, 0.15 mmol) and **18c** (39 mg, 0.15 mmol) following the general protocol 1 to afford **24c** as an off-white solid (106 mg, 80%); **m.p.**: 132.5–133.9 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.85 (s, 1H), 10.20 (s, 1H), 8.83 (d, *J* = 8.00 Hz, 1H), 8.16–8.12 (m, 2H), 7.90–7.88 (m, 2H), 7.76 (d, *J* = 8.00 Hz, 1H), 7.65–7.56 (m, 3H), 7.46–7.43 (m, 2H), 7.32 (d, *J* = 8.00 Hz, 1H), 7.21–7.14 (m, 6H), 7.06 (t, *J* = 4.00 Hz, 1H), 6.99–6.93 (m, 2H), 6.78 (t, *J* = 4.00 Hz, 1H), 4.76 (q, *J* = 8.00 Hz, 1H), 4.44 (q, *J* = 8.00 Hz, 1H), 4.22–4.16 (m, 1H), 3.66 (s, 3H), 3.27–3.21 (m, 1H), 3.15–3.09 (m, 1H), 2.95–2.94 (m, 3H), 2.75–2.69 (m, 1H), 1.83–1.80 (m, 2H), 1.40–1.37 (m, 2H), 1.34 (s, 9H), 1.29 (s, 9H), 1.18–1.14 (m, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 173.2, 171.9, 170.8, 166.9, 156.0, 155.7, 140.6, 140.5, 139.8, 138.4, 136.5, 134.8, 130.0, 129.8, 129.6, 129.5, 128.4, 127.8, 127.2, 126.6, 126.1, 124.1, 122.4, 121.4, 119.2, 118.9, 118.7, 118.3, 118.7, 118.3, 111.7, 110.0, 78.7, 77.7, 56.4, 54.6, 53.2, 37.9, 31.4, 30.7, 29.6, 28.7, 28.6; IR (ATR).vmax 3305, 3047, 2925, 1652, 1509, 1453, 1363, 1245, 1161, 1012, 855; HRMS (ESI): m/z calcd for C₅₀H₆₀N₆O₉Na [M + Na]⁺: 911.4319; found: 911.4316.

4.3.30. ¹H NMR and ¹³C NMR spectra of Methyl *N*²-(3'-((*S*)-2-((*S*)-2-((*tert*-butoxycarbonyl)amino)-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carbonyl)-*N*^w-((4-methoxy-2,3,6-trimethylphenyl)sulfonyl)-*L*-argininate (24d)

The title compound was prepared from compound **17** (100 mg, 0.15 mmol) and **18d** (60 mg, 0.15 mmol) following the general protocol 1 to afford **24d** as an off-white solid (115 mg, 75%); **m.p.**: 179.5–179.7 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.85 (s, 1H), 10.20 (s, 1H), 8.87 (d, *J* = 4.00 Hz, 1H), 8.15–8.12 (m, 2H), 7.89–7.88 (m, 2H), 7.77 (d, *J* = 4.00 Hz, 1H), 7.64–7.57 (m, 3H), 7.45–7.41 (m, 2H), 7.33 (d, *J* = 4.00 Hz, 1H), 7.21–7.13 (m, 6H), 7.07–7.04 (m, 1H), 6.98–6.93 (m, 2H), 6.66 (br s, 2H), 4.75 (q, *J* = 4.00 Hz, 1H), 4.46–4.42 (m, 1H), 4.21–4.17 (m, 1H), 3.77 (s, 3H), 3.65 (s, 3H), 3.25–3.22 (m, 1H), 3.14–3.06 (m, 3H), 2.94–2.91 (m, 1H), 2.74–2.70 (m, 1H), 2.59 (s, 3H), 2.02 (s, 3H), 1.84–1.59 (m, 2H), 1.52–1.47 (m, 2H), 1.29 (s, 9H); ¹³C NMR (150 MHz,

DMSO-*d*₆): δ 172.9, 171.9, 166.9, 157.9, 155.7, 140.7, 140.5, 139.8, 138.4, 138.1, 136.5, 136.0, 134.8, 130.1, 129.8, 129.6, 129.5, 128.4, 127.8, 127.2, 126.6, 126.1, 124.1, 122.5, 121.4, 119.3, 118.9, 118.7, 118.3, 112.1, 111.7, 110.0, 79.6, 78.7, 56.4, 55.9, 52.9, 52.4, 40.5, 37.9, 28.6, 28.4, 24.1, 18.5, 12.1; IR (ATR).vmax 2931, 1649, 1541, 1454, 1363, 1304, 1227, 1158, 1114, 1013; HRMS (ESI): m/z calcd for C₅₅H₆₄N₈O₁₀S [M + H]⁺: 1029.4466; found: 1029.4540.

4.3.31. ¹H NMR and ¹³C NMR spectra of tert-butyl ((S)-1-(((S)-3-(1H-indol-3-yl)-1-oxo-1-((3'-(((S)-2,2,18,18-tetramethyl-4,9,16-trioxo-3,17-dioxo-5,8,15-triazanonadecan-10-yl)carbamoyl)-[1,1'-biphenyl]-3-yl)amino)propan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate (24e)

The title compound was prepared from compound **17** (100 mg, 0.15 mmol) and **18e** (58 mg, 0.15 mmol) following the general protocol 1 to afford **24e** as an off-white solid (106 mg, 70%); m.p.: 131.9–132.4 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.84 (s, 1H), 10.19 (s, 1H), 8.53 (d, *J* = 6.00 Hz, 1H), 8.13 (br s, 2H), 8.00 (t, *J* = 6.00 Hz, 1H), 7.91-7.88 (m, 2H), 7.73 (d, *J* = 12.00 Hz, 1H), 7.63-7.62 (m, 2H), 7.56 (t, *J* = 6.00 Hz, 1H), 7.44-7.42 (m, 2H), 7.31 (d, *J* = 12.00 Hz, 1H), 7.21-7.13 (m, 6H), 7.06-7.03 (m, 1H), 6.97-6.92 (m, 2H), 6.76-6.73 (m, 2H), 4.75 (q, *J* = 6.00 Hz, 1H), 4.39-4.37 (m, 1H), 4.20-4.16 (m, 1H), 3.25-3.05 (m, 4H), 3.00-2.86 (m, 5H), 2.73-2.69 (m, 1H), 1.75-1.67 (m, 2H), 1.34 (s, 9H), 1.33 (s, 9H), 1.28 (s, 9H), 1.16-1.13 (m, 2H), 0.96-0.94 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.4, 171.9, 170.8, 166.7, 156.1, 156.0, 155.7, 140.6, 140.5, 139.8, 138.4, 136.5, 135.3, 129.8, 129.6, 129.3, 128.4, 127.8, 127.2, 126.6, 126.2, 124.1, 122.5, 121.4, 118.9, 118.3, 111.7, 110.0, 78.7, 78.1, 77.7, 56.4, 54.6, 54.0, 56.4, 54.6, 54.0, 40.5, 39.3, 37.9, 31.8, 29.7, 28.7, 28.6, 28.5, 23.6; IR (ATR).vmax 3292, 2973, 1650, 1510, 1453, 1247, 1163, 1045, 856, 741; HRMS (ESI): m/z calcd for C₅₆H₇₂N₈O₁₀Na [M + Na]⁺: 1039.5269; found: 1039.5261.

4.3.32. ¹H NMR and ¹³C NMR spectra of tert-butyl ((S)-1-(((S)-1-((3'-(((S)-6-amino-1-((2-(2,3-dibocguanidino)ethyl)amino)-1-oxohexan-2-yl)carbamoyl)-[1,1'-biphenyl]-3-yl)amino)-3-(1H-indol-3-yl)-1-oxopropan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate (24f)

The title compound was prepared from compound **17** (100 mg, 0.15 mmol) and **18f** (80 mg, 0.15 mmol) following the general protocol 1 to afford **24f** as an off-white solid (128 mg, 74%); m.p.: 192.5–194.0 °C; ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.50 (s, 1H), 10.80 (s, 1H), 10.19 (s, 1H), 8.52 (d, *J* = 12.00 Hz, 1H), 8.35 (t, *J* = 6.00 Hz, 1H), 8.13 (br s, 3H), 7.90-7.74 (m, 2H), 7.73 (d, *J* = 12.00 Hz, 1H), 7.64-7.62 (m, 2H), 7.55 (t, *J* = 12.00 Hz, 1H), 7.44-7.41 (m, 2H), 7.31 (d, *J* = 12.00 Hz, 1H), 7.20-7.12 (m, 6H), 7.07-7.03 (m, 1H), 6.98-6.92 (m, 2H), 6.73 (t, *J* = 6.00 Hz, 1H), 4.75 (q, *J* = 6.00 Hz, 1H), 4.40 (q, *J* = 12.00 Hz, 1H), 4.20-4.15 (m, 1H), 3.34-3.36 (m, 2H), 3.30-3.10 (m, 4H), 2.94-2.86 (m, 3H), 2.74-2.71 (m, 1H), 1.75-1.68 (m, 2H), 1.43 (s, 9H), 1.38 (s, 9H), 1.37-1.34 (m, 2H), 1.28 (s, 9H), 1.32 (s, 9H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.6, 172.0, 170.8, 166.5, 163.5, 156.1, 155.7, 152.3, 151.8, 140.6, 140.5, 139.8, 136.5, 135.3, 129.8, 129.6, 129.3, 128.4, 127.8, 127.2, 126.6, 126.2, 124.1, 121.3, 119.3, 118.9, 118.7, 118.4, 111.7, 110.0, 83.3, 79.6, 78.6, 77.7, 56.4, 54.6, 54.1, 38.3, 31.9, 29.7, 28.7, 28.5, 28.4, 28.0, 23.6; IR (ATR).vmax 3284, 3066, 2973, 1635, 1540, 1363, 1322, 1247, 1131, 1022, 856, 741; HRMS (ESI): m/z calcd for C₆₂H₈₂N₁₀O₁₂Na [M + Na]⁺: 1181.6011; found: 1181.6010.

4.3.33. ¹H NMR and ¹³C NMR spectra of tert-butyl ((S)-1-(((S)-1-((3'-(((S)-1-((2-(2,3-dibocguanidino)ethyl)amino)-5-(3-(4-methoxy-2,3,6-trimethylphenyl)sulfonyl)guanidino)-1-oxopentan-2-yl)carbamoyl)-[1,1'-biphenyl]-3-yl)amino)-3-(1H-indol-3-yl)-1-oxopropan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate (24g)

The title compound was prepared from compound **17** (50 mg, 0.075 mmol) and **18g** (50 mg, 0.15 mmol) following the general protocol 1 to afford **24g** as a gummy solid (50 mg, 49%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 11.40 (s, 1H), 10.84 (s, 1H), 10.19 (s, 1H), 8.56 (d, *J* = 12.00 Hz, 1H), 8.33 (t, *J* = 6.00 Hz, 1H), 8.18-8.14 (m, 3H), 7.91-7.89 (m, 2H), 7.75-7.89 (m, 2H), 7.74 (d, *J* = 12.00 Hz, 1H), 7.69-7.62 (m, 2H), 7.56 (t, *J* = 12.00 Hz, 1H), 7.43-7.40 (m, 2H), 7.32 (d, *J* = 12.00 Hz, 1H), 7.21-7.09 (m, 6H), 7.06 (t, *J* = 6.00 Hz, 1H), 6.95 (dd, *J* = 12.00, 24.00 Hz, 2H), 6.78-6.66 (m, 2H), 6.56-6.36 (m, 2H), 4.75 (q, *J* = 6.00 Hz, 1H), 4.43 (q, *J* = 12.00 Hz, 1H), 4.21-4.15 (m, 1H), 3.75 (s, 3H), 3.27-3.05 (m, 8H), 2.94-2.90 (m, 1H), 2.74-2.68 (m, 1H), 2.57 (s, 3H), 2.49 (s, 3H), 2.00 (s, 3H), 1.80-1.53 (m, 4H), 1.41 (s, 9H), 1.38 (s, 9H), 1.28 (s, 9H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.4, 170.8, 166.6, 163.5, 157.9, 156.6, 156.0, 152.3, 140.5, 140.53, 139.8, 138.4, 138.1, 136.5, 136.0, 135.1, 129.8, 129.6, 129.3, 128.4, 127.7, 126.6, 126.2, 124.1, 121.4, 118.9, 112.1, 111.7, 110.0, 83.3, 79.6, 78.7, 56.4, 55.9, 54.6, 53.6, 40.5, 38.3, 29.5, 28.5, 28.4, 28.0, 24.0, 18.4, 12; IR (ATR).vmax 3310, 2305, 2095, 1868, 1609, 1543, 1363, 1304, 1246, 1112, 740, 683; HRMS (ESI): m/z calcd for C₆₇H₈₆N₁₂O₁₃S [M + H]⁺: 1299.6158; found: 1299.6229.

4.3.34. ¹H NMR and ¹³C NMR spectra of 3'-((S)-2-((S)-2-amino-3-phenylpropanamido)-3-(1H-indol-3-yl)propanamido)-N-(2-aminoethyl)-[1,1'-biphenyl]-3-carboxamide (25a)

The title compound was prepared from compound **24a** (80 mg, 0.101 mmol) following the general protocol 4 to afford **25a** as a white solid (38 mg, 65%); m.p.: 206.9–207.5 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.86 (d, *J* = 4.00 Hz, 1H), 10.40 (s, 1H), 8.94 (t, *J* = 4.00 Hz, 1H), 8.60 (d, *J* = 8.00 Hz, 1H), 8.27 (br s, 3H), 8.16 (br s, 1H), 7.84-7.83 (m, 2H), 7.76-7.67 (m, 5H), 7.55 (t, *J* = 16.00 Hz, 1H), 7.32 (d, *J* = 8.00 Hz, 1H), 7.20-7.05 (m, 6H), 7.02-6.98 (m, 3H), 4.77-4.76 (m, 2H), 3.70 (q, *J* = 4.00 Hz, 1H), 3.54 (q, *J* = 8.00 Hz, 1H), 3.20-3.17 (m, 1H), 3.07-2.99 (m, 3H), 2.89-2.83 (m, 1H), 2.63-2.57 (m, 1H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 172.3, 170.9, 167.2, 164.7, 140.1, 139.1, 137.6, 136.6, 135.2, 134.8, 129.8, 129.5, 129.4, 128.5, 127.7, 127.5, 126.8, 125.5, 124.4, 121.4, 120.3, 119.1, 118.7, 111.8, 109.9, 55.3, 54.6, 39.1, 37.9, 28.6; IR (ATR).vmax 2919, 2848, 2250, 1654, 1537, 1397, 1299, 1173, 1067, 743; HRMS (ESI): m/z calcd for C₃₅H₃₆N₆O₃ [M + H]⁺: 589.2849; found: 589.2911.

4.3.35 ¹H NMR and ¹³C NMR spectra of 3'-((S)-2-((S)-2-amino-3-phenylpropanamido)-3-(1H-indol-3-yl)propanamido)-N-(2-guanidinoethyl)-[1,1'-biphenyl]-3-carboxamide (25b)

The title compound was prepared from compound **24b** (80 mg, 0.08 mmol) following the general protocol 4 to afford **25b** as a white gummy solid (27 mg, 49%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.89 (br s, 1H), 10.35 (br s, 1H), 8.94 (d, *J* = 4.00 Hz, 1H), 8.75 (t, *J* = 4.00 Hz, 1H), 8.11-7.93 (m, 4H), 7.87-7.85 (m, 1H), 7.77-7.75 (m, 1H), 7.68 (d, *J* = 8.00 Hz, 1H), 7.63-7.58 (m, 4H), 7.46-7.42 (m, 4H), 7.26-7.15 (m, 6H), 7.08-6.97 (m, 4H), 4.82 (q, *J* = 4.00 Hz, 1H), 4.07 (t, *J* = 4.00 Hz, 1H), 3.43 (q, *J* = 4.00 Hz, 2H), 3.28-3.24 (m, 1H), 3.14-3.08 (m, 2H), 2.97-2.93 (m, 1H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 169.8, 166.6, 158.1, 157.9, 156.9, 140.2, 140.0, 139.3, 136.0, 134.8, 129.5, 129.4, 129.41, 129.1, 128.4, 127.2, 127.0, 126.3, 125.5, 123.7, 122.1, 121.0, 118.9, 118.4, 118.3, 117.8, 111.4, 109.2, 87.8, 54.5, 53.3, 40.3, 38.6, 37.1, 28.0; IR (ATR).vmax 3276, 3189, 3064, 1651, 1538, 1433, 1318, 1201, 1150, 836, 798, 742 HRMS (ESI): *m/z* calcd for C₃₆H₃₈N₈O₃ [M + H]⁺: 631.3067; found: 631.3141.

4.3.36. ¹H NMR and ¹³C NMR spectra of Methyl (3'-((*S*)-2-((*S*)-2-amino-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carbonyl)-L-lysinate (25c)

The title compound was prepared from compound **24c** (90 mg, 0.112 mmol) following the general protocol 4 to afford **25c** as a white gummy solid (46 mg, 60%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.89 (br s, 1H), 10.35 (s, 1H), 8.94 (d, *J* = 4.00 Hz, 1H), 8.87 (d, *J* = 8.00 Hz, 1H), 8.13-7.90 (m, 5H), 7.79-7.68 (m, 5H), 7.63-7.59 (m, 2H), 7.46-7.44 (m, 2H), 7.33 (d, *J* = 4.00 Hz, 1H), 7.26-7.15 (m, 6H), 7.07-7.05 (m, 1H), 6.99-6.97 (m, 1H), 4.81 (q, *J* = 4.00 Hz, 1H), 4.49-4.45 (m, 1H), 4.07 (t, *J* = 4.00 Hz, 1H), 3.66 (s, 3H), 3.28-3.24 (m, 1H), 3.14-3.09 (m, 2H), 2.99-2.94 (m, 1H), 2.79-2.77 (m, 2H), 1.85-1.80 (m, 2H), 1.60-1.53 (m, 2H), 1.47-1.38 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 173.1, 170.3, 168.6, 167.0, 158.5, 158.3, 140.6, 140.5, 139.9, 136.6, 135.2, 134.8, 130.0, 130.0, 129.5, 128.9, 127.7, 127.5, 127.1, 126.2, 124.2, 122.6, 121.5, 119.4, 118.9, 118.8, 118.3, 111.8, 109.7, 55.0, 53.7, 52.9, 52.4, 39.1, 37.6, 30.4, 28.5, 27.0, 23.1; IR (ATR).vmax 3276, 3195, 3081, 2109, 1540, 1433, 1318, 1198, 798, 743; HRMS (ESI): *m/z* calcd for C₄₀H₄₄N₆O₅ [M + H]⁺: 689.3373; found: 689.3448.

4.3.37. ¹H NMR and ¹³C NMR spectra of Methyl (3'-((*S*)-2-((*S*)-2-amino-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carbonyl)-L-argininate (25d)

The title compound was prepared from compound **24d** (90 mg, 0.087 mmol) following the general protocol 4 to afford **25d** as a brown gummy solid (25 mg, 40%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.89 (s, 1H), 10.35 (s, 1H), 8.94 (dd, *J* = 4.00, 12.00 Hz, 2H), 8.13-8.12 (m, 4H), 7.93-7.90 (m, 2H), 7.79-7.78 (m, 1H), 7.69 (d, *J* = 8.00 Hz, 1H), 7.64-7.60 (m, 3H), 7.48-7.33 (m, 4H), 7.27-7.16 (m, 7H), 7.08-7.06 (m, 1H), 7.00-6.98 (m, 1H), 4.82 (q, *J* = -4.00 Hz, 1H), 4.52-4.49 (m, 1H), 4.10 (br s, 1H), 3.67 (s, 3H), 3.29-3.25 (m, 1H), 3.15-3.09 (m, 4H), 2.99-2.95 (m, 1H), 1.91-1.54 (m, 4H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.9, 170.3, 168.4, 166.9, 158.2, 157.2, 140.6, 140.4, 139.8, 136.6, 135.1, 134.8, 130.1, 130.0, 129.9, 129.5, 128.9, 129.6, 128.9, 127.7, 126.1, 124.2, 122.6, 121.5, 119.4, 118.9, 118.9, 118.3, 111.8, 109.7, 55.0, 53.7, 52.8, 52.4, 40.7, 40.5, 37.5, 28.5, 28.1, 25.8; IR (ATR).vmax 3272, 3191, 3063, 2103, 1529, 1434, 1340, 1029, 835, 798, 744, 720; HRMS (ESI): *m/z* calcd for C₄₀H₄₄N₈O₅ [M + H]⁺: 717.3435; found: 717.3501.

4.3.38. ¹H NMR and ¹³C NMR spectra of *N*-((*S*)-6-amino-1-((2-aminoethyl)amino)-1-oxohexan-2-yl)-3'-((*S*)-2-((*S*)-2-amino-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxamide (25e)

The title compound was prepared from compound **24e** (90 mg, 0.088 mmol) following the general protocol 4 to afford **25e** as a gummy solid (45 mg, 49%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.91 (s, 1H), 10.36 (s, 1H), 8.97 (d, *J* = 6.00 Hz, 1H), 8.68 (d, *J* = 6.00 Hz, 1H), 8.24 (t, *J* = 6.00 Hz, 1H), 8.16-7.67 (m, 12H), 7.68 (d, *J* = 6.00 Hz, 1H), 7.62-7.58 (m, 2H), 7.46-7.45 (m, 2H), 7.33 (d, *J* = 12.00 Hz, 1H), 7.26-7.15 (m, 6H), 7.06 (t, *J* = 6.00 Hz, 1H), 6.98 (t, *J* = 6.00 Hz, 1H), 4.81 (q, *J* = 6.00 Hz, 1H), 4.44-4.41 (m, 1H), 4.09 (t, *J* = 6.00 Hz, 1H), 3.31-3.30 (m, 2H), 3.27 (dd, *J* = 6.00, 12.00 Hz, 1H), 3.15-3.09 (m, 2H), 2.97 (dd, *J* = 12.00, 15.00 Hz, 1H), 2.87 (t, *J* = 6.00 Hz, 2H), 2.77 (br s, 2H), 1.84-1.81 (m, 1H), 1.76-1.72 (m, 1H), 1.59-1.52 (m, 2H), 1.42-1.32 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.8, 170.3, 168.5, 166.9, 158.5, 158.3, 140.5, 139.8, 136.6, 135.1, 135.12, 130.0, 129.9, 129.4, 128.8, 127.7, 127.5, 127.2, 126.2, 124.2, 122.6, 121.5, 119.3, 118.9, 118.7, 118.3, 118.8, 109.7, 55.03, 53.8, 53.7, 40.5, 39.5, 39.2, 39.9, 37.4, 36.9, 31.3, 28.4, 27.1, 23.2; IR (ATR).vmax 3274, 3056, 2926, 2119, 1661, 1526, 1431, 1200, 1121, 836, 797, 743, 721; HRMS (ESI): *m/z* calcd for C₄₁H₄₈N₈O₄ [M + H]⁺: 717.3435; found: 717.3501.

4.3.39. ¹H NMR and ¹³C NMR spectra of *N*-((*S*)-6-amino-1-((2-guanidinoethyl)amino)-1-oxohexan-2-yl)-3'-((*S*)-2-((*S*)-2-amino-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-[1,1'-biphenyl]-3-carboxamide (25f)

The title compound was prepared from compound **24f** (100 mg, 0.086 mmol) following the general protocol 4 to afford **25f** as a gummy solid (41 mg, 40%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.90 (d, *J* = 6.00 Hz, 1H), 10.35 (s, 1H), 8.93 (br s, 1H), 8.65 (d, *J* = 6.00 Hz, 1H), 8.20-8.13 (m, 4H), 7.96-7.91 (m, 2H), 7.84-7.53 (m, 7H), 7.51-7.43 (m, 2H), 7.37-7.10 (m, 8H), 7.10-7.04 (m, 1H), 7.00-6.97 (m, 1H), 6.54 (br s, 1H), 4.82 (q, *J* = 6.00 Hz, 1H), 4.47-4.44 (m, 1H), 4.06 (br s, 1H), 3.46-3.39 (m, 2H), 3.27-2.77 (m, 8H), 1.91-1.24 (m, 6H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.9, 170.3, 166.7, 158.5, 158.3, 157.4, 140.5, 139.7, 136.5, 135.1, 130.0, 129.9, 129.4, 128.9, 127.7, 127.5, 126.3, 124.2, 123.4, 122.63, 122.6, 121.5, 119.4, 118.9, 118.7, 118.4, 118.3, 116.7, 111.8, 109.7, 54.9, 54.0, 53.7, 40.8, 40.5, 39.2, 38.4, 31.4, 28.5, 27.2, 23.2; IR (ATR).vmax 3265, 3065, 1655, 1537, 1428, 1170, 1128, 1001, 787; HRMS (ESI): *m/z* calcd for C₄₂H₅₀N₁₀O₄ [M + H]⁺: 759.4017; found: 759.4084.

4.3.40. ¹H NMR and ¹³C NMR spectra of 3'-((*S*)-2-((*S*)-2-amino-3-phenylpropanamido)-3-(1*H*-indol-3-yl)propanamido)-*N*-((*S*)-5-guanidino-1-((2-guanidinoethyl)amino)-1-oxopentan-2-yl)-[1,1'-biphenyl]-3-carboxamide (25g)

914 The title compound was prepared from compound **24g** (48 mg, 0.036 mmol) following the general protocol 4 to afford **25g** as a
915 gummy solid (20 mg, 40%). ¹H NMR (600 MHz, DMSO-*d*₆): δ 10.84 (s, 1H), 10.27 (s, 1H), 8.84 (s, 1H), 8.62 (d, *J* = 6.00 Hz,
916 1H), 8.16-8.11 (m, 2H), 7.94 (d, *J* = 6.00 Hz, 1H), 7.76 (d, *J* = 12.00 Hz, 1H), 7.67 (d, *J* = 12.00 Hz, 1H), 7.62-7.56 (m, 3H), 7.46
917 (d, *J* = 6.00 Hz, 2H), 7.34 (d, *J* = 12.00 Hz, 1H), 7.28-7.14 (m, 8H), 7.08-7.06 (m, 2H), 7.01-6.97 (m, 1H), 4.82 (q, *J* = 6.00 Hz,
918 1H), 4.51-4.46 (m, 1H), 4.07 (br s, 1H), 3.25-3.19 (m, 6H), 3.15-3.12 (m, 3H), 2.96-2.94 (m, 1H), 1.89-1.84 (m, 1H), 1.76-1.72
919 (m, 1H), 1.60-1.50 (m, 2H); ¹³C NMR (150 MHz, DMSO-*d*₆): δ 172.6, 170.3, 166.8, 158.6, 158.4, 158.2, 157.5, 157.2, 140.6,
920 140.5, 139.8, 136.6, 135.1, 130.0, 130.0, 129.8, 129.4, 128.8, 128.8, 127.7, 127.4, 127.2, 126.2, 124.1, 122.6, 121.4, 119.4, 118.9,
921 118.7, 118.4, 116.8, 111.8, 109.8, 55.0, 53.5, 40.98, 40.9, 40.6, 38.4, 29.1, 28.4, 25.8; IR (ATR).vmax 3279, 3186, 3086, 1646,
922 1529, 1431, 1175, 1125, 798, 743; HRMS (ESI): *m/z* calcd for C₄₂H₅₀N₁₂O₄ [M + H]⁺: 787.4078; found: 787.4152.

923 4.3.41. ¹H NMR and ¹³C NMR spectra of 3'-nitro-[1,1'-biphenyl]-3-carboxylic acid (26)

924 To the solution of **9** (1.2 g, 3.88 mmol) in THF (10.0 mL) and MeOH (10.0 mL), was added a 1N NaOH_(aq) (7.77 mL, 7.77 mmol)
925 and stirred at room temperature for 16 h. Ethyl acetate was added and the layers were separated. The aqueous layer was then
926 acidified with 1N HCl and then extracted with CH₂Cl₂ (2 X 100 mL) and then the solvent was removed under reduced pressure to
927 yield **26** (0.84 g, 90%) as a pale-yellow solid; **m.p.**: 209.4–210.6 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 13.19 (br s, 1H), 8.44 (s,
928 1H), 8.25 (br s, 2H), 8.18 (d, *J* = 7.60 Hz, 1H), 8.02 (t, *J* = 8.00 Hz, 1H), 7.78 (t, *J* = 8.00 Hz, 1H); ¹³C NMR (100 MHz, DMSO-
929 *d*₆): δ 167.5, 148.9, 141.3, 138.6, 133.9, 132.2, 131.9, 131.1, 130.1, 129.8, 128.1, 123.1, 121.8; IR (ATR).vmax 2965, 2840, 2541,
930 2099, 1681, 1590, 1514, 1421, 1189, 1095, 928, 853; HRMS (ESI): *m/z* calcd for C₁₃H₉NO₄Na [M + Na]⁺: 266.0429; found:
931 266.0424.

932 4.3.42. ¹H NMR and ¹³C NMR spectra of *N*-(2-(2,3-dibocguanidino)ethyl)-3'-nitro-[1,1'-biphenyl]-3-carboxamide (27)

933 The title compound **27** was prepared from compound **18b** (0.621 g, 2.00 mmol) and **33** (0.5 g, 2.00 mmol) according to the
934 protocol 2. Off-white solid (0.47 g, 45%); **m.p.**: 117.6–118.9 °C; ¹H NMR (400 MHz, CDCl₃): δ 11.51 (br s, 1H), 8.83-8.82 (m,
935 1H), 8.26-8.16 (m, 3H), 8.00-7.97 (m, 1H), 7.84-7.82 (m, 1H), 7.66-7.63 (m, 1H), 7.56 (t, *J* = 8.00 Hz, 1H), 3.80-3.77 (m, 2H),
936 3.71-3.67 (m, 2H), 1.51 (s, 9H), 1.49 (s, 9H); ¹³C NMR (100 MHz, CDCl₃): δ 167.1, 162.7, 157.9, 153.0, 148.8, 142.0, 139.1,
937 135.4, 133.2, 129.9, 129.8, 129.1, 126.9, 126.3, 122.4, 122.0, 83.8, 79.9, 79.7, 42.5, 39.7, 28.3, 28.2, 28.0; IR (ATR).vmax 3196,
938 2985, 2930, 1784, 1719, 1532, 1473, 1268, 1139, 869; HRMS (ESI): *m/z* calcd for C₄₁H₅₁N₅O₈ [M + Na]⁺: 550.2278; found:
939 550.2273.

940 4.3.43. ¹H NMR and ¹³C NMR spectra of 3'-amino-*N*-(2-guanidinoethyl)-[1,1'-biphenyl]-3-carboxamide (28)

941 The title compound **28** was prepared from **27** (0.25 g, 0.47 mmol) according to the protocol 4. Pale brown solid (139 mg, 90%
942 yield); **m.p.**: 183.8–184.5 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.93 (t, *J* = 5.20 Hz, 1H), 8.54 (t, *J* = 2.00 Hz, 1H), 8.28-8.25 (m,
943 3H), 7.98 (t, *J* = 7.60 Hz, 2H), 7.83-7.75 (m, 2H), 7.65 (t, *J* = 8.00 Hz, 1H), 7.26 (br s, 3H), 3.48-3.43 (m, 2H), 3.39-3.35 (m, 2H);
944 ¹³C NMR (100 MHz, DMSO-*d*₆): δ 166.8, 157.5, 148.9, 141.5, 138.3, 135.5, 133.9, 131.1, 130.3, 129.8, 128.0, 126.1, 123.0,
945 121.7, 40.7, 39.1; IR (ATR).vmax 3280, 3155, 2341, 2112, 1638, 1522, 1457, 1344, 1299, 1192, 1114, 996, 864, 798, 675, 730;
946 HRMS (ESI): *m/z* calcd for C₁₆H₁₇N₅O₃ [M + H]⁺: 327.1331; found: 328.1401.

947 4.3.44. ¹H NMR and ¹³C NMR spectra of 3'-amino-*N*-(2-guanidinoethyl)-[1,1'-biphenyl]-3-carboxamide (29)

948 To a stirred solution of **28** (0.1 g, 0.305 mmol) in anhydrous THF (100 mL) under nitrogen atmosphere 10% palladium on
949 activated charcoal (0.15 g) was added. The reaction was evacuated and placed under a hydrogen atmosphere and stirred overnight.
950 The reaction mixture was filtered through celite, and the solvent was removed under reduced pressure to yield the desired product
951 **29** as a brown solid (39 mg, 40% yield); **m.p.**: 233.0–233.6 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 8.76 (t, *J* = 5.20 Hz, 1H), 8.07
952 (br s, 1H), 7.83 (d, *J* = 7.60 Hz, 1H), 7.72 (d, *J* = 8.00 Hz, 1H), 7.66 (t, *J* = 5.60 Hz, 1H), 7.54 (t, *J* = 8.00 Hz, 1H), 7.14 (t, *J* =
953 7.60 Hz, 1H), 6.90-6.84 (m, 2H), 6.62-6.59 (m, 1H), 5.28 (br s, 2H), 3.44-3.37 (m, 4H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 167.2,
954 157.5, 149.5, 141.6, 140.6, 135.0, 129.9, 129.7, 129.3, 126.4, 125.7, 115.0, 114.0, 112.7, 40.8, 39.0; IR (ATR).vmax 3262, 3144,
955 2849, 2587, 2342, 1621, 1541, 1471, 1309, 1182, 1129, 807; HRMS (ESI): *m/z* calcd for C₁₆H₁₉N₅O [M + H]⁺: 298.1590; found:
956 298.1660.

957 4.3.45. ¹H NMR and ¹³C NMR spectra of Methyl *L*-tryptophanate (31)

958 The title compound **31** was prepared by adding thionyl chloride (0.79 mL, 9.80 mmol) dropwise to *L*-Tryptophan (1.0 g, 4.90
959 mmol) in methanol (20.0 mL) at 0° C and then stirred at room temperature for 16 h. The solvents were removed and reduced
960 pressure and the residue was diluted with ethylacetate (100.0 mL) and washed with saturated NaHCO₃ (40 mL), saturated brine (40
961 mL), then dried (Na₂SO₄) and concentrated under reduced pressure to afford a white solid (1.00 g, 94%); **m.p.**: 89.4–89.9 °C; ¹H
962 NMR (400 MHz, DMSO-*d*₆): δ 10.87 (s, 1H), 7.48 (d, *J* = 8.00 Hz, 1H), 7.33 (d, *J* = 8.00 Hz, 1H), 7.12 (br s, 1H), 7.05 (t, *J* =
963 8.00 Hz, 1H), 6.97 (t, *J* = 8.00 Hz, 1H), 3.63 (t, *J* = 8.00 Hz, 1H), 3.55 (s, 1H), 3.05-2.91 (m, 2H); ¹³C NMR (100 MHz, DMSO-
964 *d*₆): δ 176.1, 136.5, 127.8, 124.1, 121.3, 118.8, 118.7, 111.8, 110.3, 55.6, 51.8, 40.6, 31.1; IR (ATR).vmax 3356, 3292, 3096,
965 3051, 2916, 2871, 2338, 2095, 1727, 1567, 1448, 1351, 1290, 1222, 1104, 1012, 945, 890, 805, 737; HRMS (ESI): *m/z* calcd for
966 C₁₂H₁₄N₂O₂ [M + H]⁺: 219.1055; found: 219.1125.

967 4.3.46. ¹H NMR and ¹³C NMR spectra of Methyl (*tert*-butoxycarbonyl)-*L*-phenylalanyl-*L*-tryptophanate (32)

The title compound was prepared *via* protocol 1, using *N*-(*tert*-Butoxycarbonyl)-*L*-phenylalanine (1.21 g, 4.6 mmol) and **31** (1.00 g, 4.6 mmol) to afford the coupled product **32** as an off-white solid (1.40 g, 65%); **m.p.**: 135.3–135.7 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.89 (s, 1H), 8.29 (d, *J* = 8.00 Hz, 1H), 7.49 (d, *J* = 8.00 Hz, 1H), 7.34 (d, *J* = 8.00 Hz, 1H), 7.23–7.18 (m, 6H), 7.07 (t, *J* = 8.00 Hz, 1H), 6.99 (t, *J* = 8.00 Hz, 1H), 6.87 (d, *J* = 12.00 Hz, 1H), 4.55 (q, *J* = 8.00 Hz, 1H), 4.23–4.18 (m, 1H), 3.55 (s, 3H), 3.20–3.07 (m, 2H), 2.95–2.90 (m, 1H), 2.72–2.66 (m, 1H), 1.28 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 172.6, 172.2, 155.6, 138.5, 136.5, 129.6, 128.4, 127.5, 126.6, 124.2, 121.4, 118.9, 118.4, 111.8, 109.6, 78.5, 56.0, 53.5, 52.2, 37.8, 28.5, 28.2, 27.5; IR (ATR).vmax 3295, 3001, 2952, 1694, 1648, 1525, 1433, 1288, 1164, 1112, 922, 846; HRMS (ESI): *m/z* calcd for C₂₆H₃₁N₃O₅ [M + H]⁺: 465.23; found: 337.14340

4.3.47. ¹H NMR and ¹³C NMR spectra of (*tert*-butoxycarbonyl)-*L*-phenylalanyl-*L*-tryptophan (**33**)

To a solution of **31** (1.4 g, 3.00 mmol) in THF (10.0 mL) and MeOH (10.0 mL), was added a 1N NaOH_(aq) (6.0 mL, 6.00 mmol) and stirred at room temperature for 16 h. Ethyl acetate was added and the layers were separated. The aqueous layer was then acidified with 1N HCl and then extracted with CH₂Cl₂ (2 X 100 mL) and then the solvent was removed under reduced pressure to yield **33** (1.23 g, 91%) as an off-white solid; **m.p.**: 133.2–134.0 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.89 (s, 1H), 8.08 (d, *J* = 8.00 Hz, 1H), 7.54 (d, *J* = 8.00 Hz, 1H), 7.33 (d, *J* = 8.00 Hz, 1H), 7.24–7.17 (m, 6H), 7.08–7.04 (m, 1H), 7.00–6.96 (m, 1H), 6.87 (d, *J* = -8.00 Hz, 1H), 4.51 (q, *J* = 8.00 Hz, 1H), 4.21–4.15 (m, 1H), 3.25–3.17 (m, 1H), 3.11–3.06 (m, 1H), 2.96–2.92 (m, 1H), 2.71–2.65 (m, 1H), 1.28 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 173.6, 172.1, 155.6, 138.5, 136.5, 129.6, 128.4, 127.7, 126.5, 124.1, 121.3, 118.8, 118.6, 111.8, 110.3, 110.0, 78.5, 56.1, 53.3, 37.9, 31.4, 28.5, 27.5, 22.5, 14.4; IR (ATR).vmax 3310, 3046, 2926, 1650, 1509, 1436, 1364, 1247, 1158, 1011, 848; HRMS (ESI): *m/z* calcd for C₂₅H₂₉N₃O₅Na[M + H]⁺: 474.2005; found: 474.1998.

4.3.48. ¹H NMR and ¹³C NMR spectra of *tert*-butyl ((*S*)-1-(((*S*)-1-((2-(2,3-dibocguanidino)ethyl)amino)-3-(1*H*-indol-3-yl)-1-oxopropan-2-yl)amino)-1-oxo-3-phenylpropan-2-yl)carbamate (**34**)

The title compound **34** was prepared from compound **18b** (0.333 g, 1.1 mmol) and **33** (0.5 g, 1.1 mmol) according to the protocol 2. Off-white solid (0.36 g, 45%); **m.p.**: 180.6–181.2 °C; ¹H NMR (400 MHz, DMSO-*d*₆): δ 11.47 (br s, 1H), 10.80 (d, *J* = 4.00 Hz, 1H), 8.37 (t, *J* = 4.00 Hz, 1H), 8.13 (t, *J* = 4.00 Hz, 1H), 7.91 (d, *J* = 8.00 Hz, 1H), 7.57 (d, *J* = 8.00 Hz, 1H), 7.30 (d, *J* = 8.00 Hz, 1H), 7.20–7.06 (m, 6H), 7.07–7.02 (m, 1H), 6.98–6.94 (m, 1H), 6.88 (d, *J* = 8.00 Hz, 1H), 4.48 (q, *J* = 4.00 Hz, 1H), 4.15–4.09 (m, 1H), 3.27–2.95 (m, 6H), 2.90–2.85 (m, 1H), 2.69–2.63 (m, 1H), 1.44 (s, 9H), 1.38 (s, 9H), 1.28 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 172.0, 171.6, 163.5, 156.0, 155.6, 152.3, 138.5, 136.5, 129.6, 128.4, 127.8, 126.6, 124.0, 121.3, 118.9, 118.6, 111.7, 110.3, 83.3, 78.7, 78.6, 56.4, 53.8, 38.5, 37.9, 37.8, 28.6, 28.5, 28.4, 28.1; IR (ATR).vmax 2974, 1637, 1523, 1363, 1248, 1132, 1021; HRMS (ESI): *m/z* calcd for C₃₈H₅₃N₇O₈ [M + H]⁺: 736.3956; found: 736.4023.

4.3.49. ¹H NMR and ¹³C NMR spectra of (*S*)-2-amino-*N*-((*S*)-1-((2-guanidinoethyl)amino)-3-(1*H*-indol-3-yl)-1-oxopropan-2-yl)-3-phenylpropanamide (**35** TFA Salt)

The title compound was prepared *via* protocol 4, using **34** (0.150 g, 0.203 mmol) to yield the desired product **35** as a brown gummy solid (40 mg, 47%). ¹H NMR (400 MHz, DMSO-*d*₆): δ 10.87 (br s, 1H), 8.81 (d, *J* = 8.00 Hz, 1H), 8.30 (t, *J* = 4.00 Hz, 1H), 8.06 (br s, 3H), 7.64–7.62 (m, 2H), 7.34–7.24 (m, 9H), 7.16 (d, *J* = 4.00 Hz, 1H), 7.08–7.04 (m, 1H), 7.00–6.96 (m, 1H), 4.58–4.52 (m, 1H), 4.03 (br s, 1H), 3.26–2.88 (m, 8H); ¹³C NMR (100 MHz, DMSO-*d*₆): δ 172.0, 168.3, 157.5, 136.6, 135.2, 130.0, 129.0, 127.64, 127.6, 124.3, 121.4, 118.9, 118.7, 111.8, 110.0, 54.2, 53.6, 38.5, 37.5, 28.6; IR (ATR).vmax 3060, 2919, 1670, 1542, 1421, 1199, 1128, 1002, 825; HRMS (ESI): *m/z* calcd for C₂₃H₂₉N₇O₂ [M + H]⁺: 436.2383; found: 436.2451.

4.4 Antibacterial activity

The antimicrobial activity of compounds was evaluated through the broth micro dilution assay using the procedure described by CLSI.[64] Briefly, *Staphylococcus aureus* [SA38] were grown to mid-log phase in Muller-Hinton broth (MHB) with shaking at 120 rpm and incubated at 37 °C for 18–24 h. Following incubation, bacteria were washed three times using PBS pH 7.4 with centrifugation at 3500 g for 10 min after each wash. After washing, bacteria were diluted with fresh MHB. The turbidity of the bacterial suspensions were adjusted so that OD_{600nm} was 0.1, which gave 1×10⁸ cfu/ml, and then further diluted to achieve 1–2×10⁵ cfu/ml as a final bacterial concentration. Each compound was added at concentrations ranging from 250–3.9 μM through serial two-fold dilution. Wells in microtitre plates were loaded with 100 μl of inoculum containing 1–2×10⁵ cfu/ml bacteria. Wells containing only bacteria and without any compound were used as a negative controls (i.e. no inhibition of growth). Wells without bacteria but containing compound acted as another control. The microtitre plate was wrapped with paraffin to prevent evaporation and incubated with shaking at 120 rpm and 37 °C for 18–24 h. After incubation, spectrophotometric reading of the wells was taken at 600nm. The minimal inhibitory concentration (MIC) was defined as the lowest concentration of the biphenyl compounds that causes 100% inhibition of microbial growth. The same procedure was followed for the two Gram-negative bacteria, *Pseudomonas aeruginosa* [PA01], and *Escherichia coli* [K12]. Each experiment was performed in triplicate and was repeated in three independent experiments. The MICs of the compounds were compared to published MIC [15, 52, 53] of MSI-78. The method for determining the MIC of MSI-78 [52, 53], that recommended by the CLSI (formerly named the National Committee for Clinical Laboratory Standards (NCCLS)) was exactly the same as described herein for the current compounds.

4.5 Toxicity assay

044 Normal human lung fibroblasts MRC-5 were cultured in minimal essential medium (MEM, Invitrogen) supplemented with 10%
045 foetal calf serum (FCS), 1% L-glutamine–penicillin–streptomycin, 2% sodium bicarbonate, 1% non-essential amino acids
046 (NEAA) and 1% sodium pyruvate. The cell line was maintained at 37 °C in 5% CO₂ as an adherent monolayer and was passaged
047 upon reaching confluence by standard cell culture techniques. MRC-5 cells were seeded at 2 × 10⁴ cells per well in 96-well plates
048 to ensure full confluence (quiescence). Cells were treated 24 h after seeding with 0.1 to 1000 μM of compounds. After 72 h drug
049 incubation, the treated media was replaced with fresh media containing 10% Alamar blue and the cells were incubated for another
050 6 h. The metabolic activity was detected by spectrophotometric analysis by assessing the absorbance of Alamar blue as previously
051 described by Pasquier *et al.* [65] Cell proliferation was determined and expressed as a percentage of untreated control cells. The
052 determination of IC₅₀ values was performed using GraphPad Prism 6 (San Diego, CA, USA).]. Each experiment was performed in
053 triplicate and was repeated in three independent experiments.

054 4.6 Tethered bilayer lipid membranes

055
056 The ability of the biphenyl derivatives to interact with lipid bilayers was tested using tethered bilayer lipid membranes (tBLMs) in
057 association with AC electrical impedance spectroscopy techniques [55, 56]. Lipid bilayers were anchored to a gold substrate
058 which were created using the solvent exchange technique described previously [56, 57]. In short, tethered benzyl-disulfide (tetra-
059 ethyleneglycol) n=2 C20-phytanyl tethers: benzyl-disulfide-tetra-ethyleneglycol-OH spacers were pre-prepared in the ratio of 1:10
060 and were coated onto a gold patterned polycarbonate slide containing a large gold return electrode (SDx Tethered Membranes Pty
061 Ltd, Australia). Attached to this slide was a specialised cartridge chamber which allows for easy addition of reagent between
062 tethering and reference electrodes. A 3 mM solution of a standard mobile lipid phase [70% zwitterionic C20 Diphytanyl-Glycero-
063 Phosphatidylcholine lipid: 30% C20 Diphytanyl-diglyceride-OH ether] was added to the tethering chemistries. All lipids were
064 dissolved in 100% ethanol. Lipids were left for 2 minutes to associate with the tethering chemistries before being washed with 3 x
065 200 ml phosphate buffered saline (PBS). The membranes formed from these lipids are designated as the “zwitterionic
066 membranes”. Alternatively, negatively charged membranes, that resemble those present in many bacterial species, were produced
067 using the same mobile lipid phase which was instead supplemented with 30% palmitoyl-oleoyl-phosphatidylglycerol (POPG)
068 (Avanti Lipids, USA). The presence of a lipid bilayer was verified using AC electrical impedance spectrometry. These measures
069 were done using a TethaPod™ operated with TethaQuick™ software (SDx Tethered Membranes Pty Ltd, Australia). The signal
070 employed was 50 mV peak-to-peak AC excitation at 0.1–2000 Hz with four steps per decade. Impedance data were fitted to a
071 constant phase element (CPE) in series with a resistor/capacitor. The CPE represents the capacitance of the gold electrode
072 interface, whilst the resistor and capacitor represent the resistance and capacitance of the tethered membrane respectively. Fitting
073 was done using a proprietary adaptation of a Lev Mar fitting routine as described previously [56]. All tethered membrane assays
074 were performed in triplicate. Individual results shown are of typical responses to the antimicrobial compounds.

075 4.7 Cytoplasmic membrane permeability assay

076
077 The method was adopted from Wu *et al.* [66] with slight modification. Bacterial cytoplasmic membrane permeability was
078 determined using membrane potential sensitive dye diSC3-5 (3,3'-dipropylthiadicarbocyanine iodide) which penetrates inside
079 bacterial cells depending on the membrane potential gradient of the cytoplasmic membrane. *Staphylococcus aureus* [SA38] and
080 *Escherichia coli* [K12] were grown in MHB to mid-log phase by incubating with shaking at 37 °C for 18-24 h. Following
081 incubation bacteria were washed with 5 mM HEPES containing 20 mM glucose pH (7.2) and resuspended in the same buffer to an
082 OD₆₀₀ 0.05-0.06 which gave 1 × 10⁷ CFU/ml. diSC3-5 was added at 4 μM to the bacterial suspension. The suspensions were
083 incubated at room temperature for 1 h in the dark for maximum dye take-up by bacterial cell. Then, 100 mM KCl was added to
084 balance the K⁺ outside and inside the bacterial cell to prevent further uptake or outflow of the dye. For Gram-negative bacteria,
085 0.5 mM EDTA was used to remove the stabilizing divalent cations from the lipopolysaccharide (LPS) layer which help in dye
086 penetration without affecting bacterial growth. 100 μl of bacterial suspension was added in a 96-well microtiter plate with equal
087 volume of antimicrobial compounds. Dimethyl sulfoxide (DMSO) was used as a positive control to achieve maximum
088 fluorescence while bacterial suspension containing only dye and HEPES buffer set as blank to subtract the background.
089 Fluorescence was measured with a luminescence spectrophotometer at 3 min intervals at an excitation wavelength of 622 nm and
090 an emission wavelength of 670 nm. The experiment was performed in triplicate.

091 4.8 Biofilm inhibition assay

092
093 Bacterial cultures (*S. aureus* and *E. coli*) were grown in LB₁₀ media overnight at 37 °C with shaking at 150 rpm. Cultures were
094 diluted (1:20) in LB medium and 200 μl aliquots were dispensed to flat bottom 96-well plate wells (Sarstedt Australia). Cultures
095 were supplemented with varying concentrations of synthetic compounds dissolved in DMSO. The cultures were grown in 96-well
096 plate wells overnight along with synthetic compounds. Control cultures were supplemented with an equal amount of DMSO.
097 Plates were sealed with self-adhesive microplate sealers (TopSeal-A, PerkinElmer) to allow air diffusion and to prevent
098 condensation. Cultures were incubated overnight at 37 °C with shaking at 150 rpm. Biofilms adhered on polystyrene substratum
099 were quantified by crystal violet staining as described previously[67]. All cultures were prepared in triplicate.

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112 References

- 113
- 114 [1] W.H. Organization, WHO Antimicrobial Resistance: Global Report on Surveillance, April 2014, in.
- 115 [2] C.K. Naber, Staphylococcus aureus bacteremia: epidemiology, pathophysiology, and management strategies,
116 Clinical infectious diseases, 48 (2009) S231-S237.
- 117 [3] K.L. Brown, R.E. Hancock, Cationic host defense (antimicrobial) peptides, Current opinion in immunology, 18 (2006)
118 24-30.
- 119 [4] R.E. Hancock, H.-G. Sahl, Antimicrobial and host-defense peptides as new anti-infective therapeutic strategies,
120 Nature biotechnology, 24 (2006) 1551-1557.
- 121 [5] S. Wagner, R. Sommer, S. Hinsberger, C. Lu, R.W. Hartmann, M. Empting, A. Titz, Novel strategies for the treatment
122 of Pseudomonas aeruginosa infections, Journal of medicinal chemistry, 59 (2016) 5929-5969.
- 123 [6] M.S. Butler, M.A. Blaskovich, M.A. Cooper, Antibiotics in the clinical pipeline at the end of 2015, The Journal of
124 Antibiotics, (2016).
- 125 [7] S. Choi, A. Isaacs, D. Clements, D. Liu, H. Kim, R.W. Scott, J.D. Winkler, W.F. DeGrado, De novo design and in vivo
126 activity of conformationally restrained antimicrobial arylamide foldamers, Proceedings of the National Academy of
127 Sciences, 106 (2009) 6968-6973.
- 128 [8] A.C. Nilsson, H. Janson, H. Wold, A. Fugelli, K. Andersson, C. Håkangård, P. Olsson, W.M. Olsen, LTX-109 is a novel
129 agent for nasal decolonization of methicillin-resistant and-sensitive Staphylococcus aureus, Antimicrobial agents and
130 chemotherapy, 59 (2015) 145-151.
- 131 [9] F. García-Olmedo, A. Molina, J.M. Alamillo, P. Rodríguez-Palenzuela, Plant defense peptides, Peptide Science, 47
132 (1998) 479-491.
- 133 [10] S. Kurata, Recognition and elimination of diversified pathogens in insect defense systems, Molecular diversity, 10
134 (2006) 599-605.
- 135 [11] A.C. Rinaldi, Antimicrobial peptides from amphibian skin: an expanding scenario: Commentary, Current opinion
136 in chemical biology, 6 (2002) 799-804.
- 137 [12] H. Sugiarto, P.-L. Yu, Avian antimicrobial peptides: the defense role of β -defensins, Biochemical and biophysical
138 research communications, 323 (2004) 721-727.
- 139 [13] K. Kuroda, G.A. Caputo, Antimicrobial polymers as synthetic mimics of host-defense peptides, Wiley
140 Interdisciplinary Reviews: Nanomedicine and Nanobiotechnology, 5 (2013) 49-66.
- 141 [14] B. Mensa, G.L. Howell, R. Scott, W.F. DeGrado, Comparative mechanistic studies of brilacidin, daptomycin, and
142 the antimicrobial peptide LL16, Antimicrobial agents and chemotherapy, 58 (2014) 5136-5145.
- 143 [15] Y. Ge, D.L. MacDonald, K.J. Holroyd, C. Thornsberry, H. Wexler, M. Zasloff, In vitro antibacterial properties of
144 pexiganan, an analog of magainin, Antimicrobial agents and chemotherapy, 43 (1999) 782-788.
- 145 [16] B.M. Peters, M.E. Shirtliff, M.A. Jabra-Rizk, Antimicrobial peptides: primeval molecules or future drugs?, PLoS
146 Pathog, 6 (2010) e1001067.
- 147 [17] A.A. Bahar, D. Ren, Antimicrobial peptides, Pharmaceuticals, 6 (2013) 1543-1575.
- 148 [18] P. Méndez-Samperio, Peptidomimetics as a new generation of antimicrobial agents: current progress, Infection
149 and drug resistance, 7 (2014) 229.
- 150 [19] Y. Chen, C.T. Mant, S.W. Farmer, R.E. Hancock, M.L. Vasil, R.S. Hodges, Rational design of α -helical antimicrobial
151 peptides with enhanced activities and specificity/therapeutic index, Journal of Biological Chemistry, 280 (2005) 12316-
152 12329.
- 153 [20] A.J. Karlsson, R.M. Flessner, S.H. Gellman, D.M. Lynn, S.P. Palecek, Polyelectrolyte multilayers fabricated from
154 antifungal β -peptides: design of surfaces that exhibit antifungal activity against Candida albicans, Biomacromolecules,
155 11 (2010) 2321-2328.
- 156 [21] N.P. Chongsiriwatana, J.A. Patch, A.M. Czyzewski, M.T. Dohm, A. Ivankin, D. Gidalevitz, R.N. Zuckermann, A.E.
157 Barron, Peptoids that mimic the structure, function, and mechanism of helical antimicrobial peptides, Proceedings of
158 the National Academy of Sciences, 105 (2008) 2794-2799.
- 159 [22] R. Kapoor, M.W. Wadman, M.T. Dohm, A.M. Czyzewski, A.M. Spormann, A.E. Barron, Antimicrobial peptoids are
160 effective against Pseudomonas aeruginosa biofilms, Antimicrobial agents and chemotherapy, 55 (2011) 3054-3057.
- 161 [23] M.L. Huang, S.B.Y. Shin, M.A. Benson, V.J. Torres, K. Kirshenbaum, A comparison of linear and cyclic peptoid
162 oligomers as potent antimicrobial agents, ChemMedChem, 7 (2012) 114-122.
- 163 [24] N. Srinivas, P. Jetter, B.J. Ueberbacher, M. Werneburg, K. Zerbe, J. Steinmann, B. Van der Meijden, F. Bernardini,
164 A. Lederer, R.L. Dias, Peptidomimetic antibiotics target outer-membrane biogenesis in Pseudomonas aeruginosa,
165 Science, 327 (2010) 1010-1013.

166 [25] S. Mosca, J. Keller, N. Azzouz, S. Wagner, A. Titz, P.H. Seeberger, G. Brezesinski, L. Hartmann, Amphiphilic Cationic
167 β 3R3-Peptides: Membrane Active Peptidomimetics and Their Potential as Antimicrobial Agents, *Biomacromolecules*,
168 15 (2014) 1687-1695.

169 [26] A. Makovitzki, D. Avrahami, Y. Shai, Ultrashort antibacterial and antifungal lipopeptides, *Proceedings of the*
170 *National Academy of Sciences*, 103 (2006) 15997-16002.

171 [27] C. Ghosh, J. Haldar, Membrane-Active Small Molecules: Designs Inspired by Antimicrobial Peptides,
172 *ChemMedChem*, 10 (2015) 1606-1624.

173 [28] R. Domalaon, G. G Zhanel, F. Schweizer, Short antimicrobial peptides and peptide scaffolds as promising
174 antibacterial agents, *Current topics in medicinal chemistry*, 16 (2016) 1217-1230.

175 [29] M.B. Strøm, Ø. Rekdal, J.S. Svendsen, Antimicrobial activity of short arginine-and tryptophan-rich peptides, *Journal*
176 *of Peptide Science*, 8 (2002) 431-437.

177 [30] M. Ahn, P. Gunasekaran, G. Rajasekaran, E.Y. Kim, S.-J. Lee, G. Bang, K. Cho, J.-K. Hyun, H.-J. Lee, Y.H. Jeon, Pyrazole
178 derived ultra-short antimicrobial peptidomimetics with potent anti-biofilm activity, *European Journal of Medicinal*
179 *Chemistry*, 125 (2017) 551-564.

180 [31] S.M. Wales, K.A. Hammer, A.M. King, A.J. Tague, D. Lyras, T.V. Riley, P.A. Keller, S.G. Pyne, Binaphthyl-1, 2, 3-
181 triazole peptidomimetics with activity against *Clostridium difficile* and other pathogenic bacteria, *Organic &*
182 *biomolecular chemistry*, 13 (2015) 5743-5756.

183 [32] C. Ghosh, G.B. Manjunath, M.M. Konai, D.S. Uppu, K. Paramanandham, B.R. Shome, R. Ravikumar, J. Haldar, Aryl-
184 alkyl-lysines: Membrane-active small molecules active against murine model of burn infection, *ACS Infectious*
185 *Diseases*, 2 (2015) 111-122.

186 [33] J. Hoque, M.M. Konai, S.S. Sequeira, S. Samaddar, J. Haldar, Antibacterial and Antibiofilm Activity of Cationic Small
187 Molecules with Spatial Positioning of Hydrophobicity: An in Vitro and in Vivo Evaluation, *Journal of Medicinal*
188 *Chemistry*, 59 (2016) 10750-10762.

189 [34] S. Nizalapur, K.K. Ho, Ö. Kimyon, E. Yee, T. Berry, M. Manefield, C.G. Cranfield, M. Willcox, D.S. Black, N. Kumar,
190 Synthesis and biological evaluation of N-naphthoyl-phenylglyoxamide-based small molecular antimicrobial peptide
191 mimics as novel antimicrobial agents and biofilm inhibitors, *Organic & biomolecular chemistry*, 14 (2016) 3623-3637.

192 [35] S. Nizalapur, O. Kimyon, E. Yee, K. Ho, T. Berry, M. Manefield, C.G. Cranfield, M. Willcox, D.S. Black, N. Kumar,
193 Amphipathic guanidine-embedded glyoxamide-based peptidomimetics as novel antibacterial agents and biofilm
194 disruptors, *Organic & Biomolecular Chemistry*, (2017).

195 [36] D.A. Horton, G.T. Bourne, M.L. Smythe, The combinatorial synthesis of bicyclic privileged structures or privileged
196 substructures, *Chemical Reviews*, 103 (2003) 893-930.

197 [37] M.E. Welsch, S.A. Snyder, B.R. Stockwell, Privileged scaffolds for library design and drug discovery, *Current opinion*
198 *in chemical biology*, 14 (2010) 347-361.

199 [38] H. Aldemir, R. Richarz, T.A. Gulder, The biocatalytic repertoire of natural biaryl formation, *Angewandte Chemie*
200 *International Edition*, 53 (2014) 8286-8293.

201 [39] G. Bringmann, T. Gulder, T.A. Gulder, M. Breuning, Atroposelective total synthesis of axially chiral biaryl natural
202 products, *Chemical reviews*, 111 (2010) 563-639.

203 [40] M.C. Kozłowski, B.J. Morgan, E.C. Linton, Total synthesis of chiral biaryl natural products by asymmetric biaryl
204 coupling, *Chemical Society reviews*, 38 (2009) 3193-3207.

205 [41] A. Isnansetyo, Y. Kamei, Anti-methicillin-resistant *Staphylococcus aureus* (MRSA) activity of MC21-B, an
206 antibacterial compound produced by the marine bacterium *Pseudoalteromonas phenolica* O-BC30 T, *International*
207 *journal of antimicrobial agents*, 34 (2009) 131-135.

208 [42] H. Rahman, B. Austin, W.J. Mitchell, P.C. Morris, D.J. Jamieson, D.R. Adams, A.M. Spragg, M. Schweizer, Novel
209 anti-infective compounds from marine bacteria, *Marine drugs*, 8 (2010) 498-518.

210 [43] C.C. CHANG, G.O. MORTON, J.C. JAMES, M.M. SIEGEL, N.A. KUCK, R.T. TESTA, D.B. BORDERS, LL-AF283 antibiotics,
211 cyclic biphenyl peptides, *The Journal of antibiotics*, 44 (1991) 674-677.

212 [44] I. UCHIDA, N. SHIGEMATSU, M. EZAKI, M. HASHIMOTO, H. AOKI, H. IMANAKA, Biphenomycins A and B, novel
213 peptide antibiotics. II. Structural elucidation of biphenomycins A and B, *The Journal of antibiotics*, 38 (1985) 1462-
214 1468.

215 [45] K. Pérez-Labrada, M.A. Cruz-Mendoza, A. Chávez-Riveros, E. Hernández-Vázquez, T. Torroba, L.D. Miranda,
216 Diversity-oriented synthesis and cytotoxic activity evaluation of biaryl-containing macrocycles, *Organic & Biomolecular*
217 *Chemistry*, (2017).

218 [46] D.I. Chan, E.J. Prenner, H.J. Vogel, Tryptophan-and arginine-rich antimicrobial peptides: structures and
219 mechanisms of action, *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1758 (2006) 1184-1202.

220 [47] L. Li, I. Vorobyov, T.W. Allen, The different interactions of lysine and arginine side chains with lipid membranes,
221 *The Journal of Physical Chemistry B*, 117 (2013) 11906-11920.

- 222 [48] T. Wieprecht, M. Dathe, M. Beyermann, E. Krause, W.L. Maloy, D.L. MacDonald, M. Bienert, Peptide
223 hydrophobicity controls the activity and selectivity of magainin 2 amide in interaction with membranes, *Biochemistry*,
224 36 (1997) 6124-6132.
- 225 [49] D.-K. Lee, A. Bhunia, S.A. Kotler, A. Ramamoorthy, Detergent-type membrane fragmentation by MSI-78, MSI-367,
226 MSI-594, and MSI-843 antimicrobial peptides and inhibition by cholesterol: a solid-state nuclear magnetic resonance
227 study, *Biochemistry*, 54 (2015) 1897-1907.
- 228 [50] A. Ramamoorthy, S. Thennarasu, D.-K. Lee, A. Tan, L. Maloy, Solid-state NMR investigation of the membrane-
229 disrupting mechanism of antimicrobial peptides MSI-78 and MSI-594 derived from magainin 2 and melittin, *Biophysical*
230 *journal*, 91 (2006) 206-216.
- 231 [51] L.M. Gottler, A. Ramamoorthy, Structure, membrane orientation, mechanism, and function of pexiganan—a
232 highly potent antimicrobial peptide designed from magainin, *Biochimica et Biophysica Acta (BBA)-Biomembranes*,
233 1788 (2009) 1680-1686.
- 234 [52] A.K. Jepson, J. Schwarz-Linek, L. Ryan, M.G. Ryadnov, W.C. Poon, What Is the 'Minimum Inhibitory
235 Concentration'(MIC) of Pexiganan Acting on *Escherichia coli*?—A Cautionary Case Study, in: *Biophysics of Infection*,
236 Springer, 2016, pp. 33-48.
- 237 [53] Q.Y. Lau, F.M. Ng, J.W.D. Cheong, Y.Y.A. Yap, Y.Y.F. Tan, R. Jureen, J. Hill, C.S.B. Chia, Discovery of an ultra-short
238 linear antibacterial tetrapeptide with anti-MRSA activity from a structure–activity relationship study, *European journal*
239 *of medicinal chemistry*, 105 (2015) 138-144.
- 240 [54] J. O'brien, I. Wilson, T. Orton, F. Pognan, Investigation of the Alamar Blue (resazurin) fluorescent dye for the
241 assessment of mammalian cell cytotoxicity, *European Journal of Biochemistry*, 267 (2000) 5421-5426.
- 242 [55] C.G. Cranfield, B.a. Cornell, S.L. Grage, P. Duckworth, S. Carne, A.S. Ulrich, B. Martinac, Transient potential
243 gradients and impedance measures of tethered bilayer lipid membranes: Pore-forming peptide insertion and the effect
244 of electroporation, *Biophysical Journal*, 106 (2014) 182-189.
- 245 [56] C.G. Cranfield, T. Bettler, B. Cornell, Nanoscale ion sequestration to determine the polarity selectivity of ion
246 conductance in carriers and channels, *Langmuir*, 31 (2015) 292-298.
- 247 [57] C. Cranfield, S. Carne, B. Martinac, B. Cornell, The Assembly and Use of Tethered Bilayer Lipid Membranes (tBLMs),
248 in: *Methods in Membrane Lipids*, Springer New York, 2015, pp. 45-53.
- 249 [58] C.G. Cranfield, T. Berry, S.A. Holt, K.R. Hossain, A.P. Le Brun, S. Carne, H. Al Khamici, H. Coster, S.M. Valenzuela, B.
250 Cornell, Evidence of the key role of H₃O⁺ in phospholipid membrane morphology, *Langmuir*, 32 (2016) 10725-10734.
- 251 [59] F.-Y. Chen, M.-T. Lee, H.W. Huang, Evidence for membrane thinning effect as the mechanism for peptide-induced
252 pore formation, *Biophysical journal*, 84 (2003) 3751-3758.
- 253 [60] P. Jorge, D. Grzywacz, W. Kamysz, A. Lourenço, M.O. Pereira, Searching for new strategies against biofilm
254 infections: Colistin-AMP combinations against *Pseudomonas aeruginosa* and *Staphylococcus aureus* single-and
255 double-species biofilms, *PloS one*, 12 (2017) e0174654.
- 256 [61] O. Cirioni, C. Silvestri, R. Ghiselli, W. Kamysz, D. Minardi, P. Castelli, F. Orlando, E. Kamysz, M. Provinciali, G.
257 Muzzonigro, In vitro and in vivo effects of sub-MICs of pexiganan and imipenem on *Pseudomonas aeruginosa* adhesion
258 and biofilm development, *Le infezioni in medicina: rivista periodica di eziologia, epidemiologia, diagnostica, clinica e*
259 *terapia delle patologie infettive*, 21 (2013) 287-295.
- 260 [62] G. Wang, B. Mishra, R.F. Epan, R.M. Epan, High-quality 3D structures shine light on antibacterial, anti-biofilm
261 and antiviral activities of human cathelicidin LL-37 and its fragments, *Biochimica et Biophysica Acta (BBA)-*
262 *Biomembranes*, 1838 (2014) 2160-2172.
- 263 [63] M.F. Mohamed, A. Abdelkhalek, M.N. Seleem, Evaluation of short synthetic antimicrobial peptides for treatment
264 of drug-resistant and intracellular *Staphylococcus aureus*, *Scientific reports*, 6 (2016) 29707.
- 265 [64] M.A. Wikler, Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically: approved
266 standard, *Clinical and Laboratory Standards Institute*, 2006.
- 267 [65] E. Pasquier, J. Ciccolini, M. Carre, S. Giacometti, R. Fanciullino, C. Pouchy, M.-P. Montero, C. Serdjebi, M. Kavallaris,
268 N. André, Propranolol potentiates the anti-angiogenic effects and anti-tumor efficacy of chemotherapy agents:
269 implication in breast cancer treatment, *Oncotarget*, 2 (2011) 797.
- 270 [66] M. Wu, E. Maier, R. Benz, R.E. Hancock, Mechanism of interaction of different classes of cationic antimicrobial
271 peptides with planar bilayers and with the cytoplasmic membrane of *Escherichia coli*, *Biochemistry*, 38 (1999) 7235-
272 7242.
- 273 [67] G.A. O'Toole, Microtiter dish biofilm formation assay, *Journal of visualized experiments: JoVE*, (2011).