

Novel Fingermark Detection Methods Using Biomolecular Recognition

by

Rolanda Lam

A thesis submitted for the

Degree of Doctor of Philosophy (Science)

University of Technology Sydney

February 2018

Certificate of Authorship and Originality

I certify that the work in this thesis has not previously been submitted for a degree nor has it been submitted as part of the requirements for a degree except as fully acknowledged within the text.

I also certify that the thesis has been written by me. Any help that I have received in my research work and the preparation of the thesis itself has been acknowledged. In addition, I certify that all the information sources and literature used are indicated in the thesis.

Production Note: Signature removed prior to publication.

Rolanda Lam February 17, 2018 Dedicated to Somebody I Loved Dearly – You will forever be in my thoughts.

Acknowledgements

This research was supported by the Australian Research Council (ARC) Linkage Project LP130101019. Therefore, I'd like to thank Claude for giving me plenty of opportunities to develop both professionally and academically during my time in Australia, as well as Xanthe, Chris, and the rest of the ARC linkage project committee (academic and industry partners) for choosing me to join their team.

I met a lot of people during my three years at the University of Technology Sydney (UTS), most within the UTS:Centre for Forensic Science, but some in other research areas too. I wish all the Honours and Higher Degree Research students (former and current) all the best. Within the UTS Fingerprint research group, I'd like to share my gratitude for two unsung heroes. Seb, your evaluation program saved me countless hours of image processing and data analysis…I don't even know how to thank you, but thank you! Scott, you may be younger than me, but you are wise…thanks for all the advice and chats!

I'd like to thank everybody in the ever-growing (or is it now ever-diminishing?) o-o family. Without you guys, I don't know how I would have gotten through these past three years. I'm going to miss our 12:00 lunches and you helping me prove to my family and friends back home that I had the illusion of a social life. Good luck organising things without me! NoNo, there's still so much left on our Aussie bucket list…#FunwithRo will just have to continue elsewhere in the years to come!

Back on Canadian soil, I'd like to thank Della for bringing this opportunity to my attention…and constantly reminding me to come back. You may have had ulterior motives, but they were spoiled by funny man Brian. Brian, the Royal Canadian Mounted Police (RCMP) isn't going to be the same without you. While I didn't get to see everybody during my quick visits, I know that I had the support of many RCMP colleagues, especially Maryse who was holding down the National Division Ident fort in my absence.

A huge thank you goes out to Maria for taking a chance on me when I first asked for a tour of her Carleton University lab and then agreeing to have me there for four months. I know you'll never lead me astray and am looking forward to the possibilities of conducting more research with you. Annamaria, without you, I wouldn't have been able to make as much progress as quickly as I did with the aptamer development, so thank you, thank you, thank you! You're a great teacher and it was nice to know that, even though we barely knew each other, we were there for one another to get through whatever life threw our way. I'd like to thank Karl, Carlos, and Alex who assisted me with my GC-MS and sequencing samples. I'd also like to thank the rest of the DeRosa lab group for welcoming the student from Australia who sounded just like you!

I know I was horrible at keeping in touch with everybody back home, but I'll be sure to catch up with all my friends in the coming months. A special shout out goes to Katreena, who frequently told me to quit just so she wouldn't miss me anymore. You have one funny way of showing you care!

Last, but not least, I'd like to thank my family. In particular, Mom, Dad – Thanks for your support and encouraging me to take the initial leap to move to the other side of the world. Beck – Thanks for checking up on me frequently. Bert, Rich – Thanks for "visiting" me.

For those who know me, I like to plan…and let's just say things did not go as planned after I accepted the offer to do a PhD at UTS. But hey, when life gives you lemons, make lemonade, right? (Bet you were all thinking I'd end with "eh", eh?)

Table of Contents

List of Figures

Figure 2-8 Schematic diagrams depicting manner in which quartered fingermarks were processed to compare the eight different time intervals.. 69

Figure 2-9 Schematic diagrams depicting the different processing configurations for: (a) latent fingermarks; (b) blood-contaminated fingermarks; and (c) saliva- and semencontaminated fingermarks. (aq = aqueous working solution; $BSA = a$ queous working solution containing bovine serum albumin; $EG =$ glycolic working solution; $M# =$ multiplex solution containing $\#$ antibodies/aptamers; MeOH = ice-cold methanol fixing solution used; 5-SSA = 5-sulfosalicylic acid fixing solution used) 72

Figure 2-10 Schematic diagram showing how a split fingermark was processed. Used multiplex solution was created by exposing it to additional fingermarks pre-treated with the various routine technique sequences. For blood-contaminated fingermarks, 5- SSA fixing solution was used prior to the multiplex solution application. ($aq = aqueous$) working solution; $M#$ = multiplex solution containing # antibodies/aptamers) 76

Figure 2-11 Natural fingermark deposited on PVDF and processed for 15 minutes (top right), 30 minutes (bottom right), 45 minutes (top left), and one hour (bottom left), visualised at 590 nm with a 650 nm bandpass filter. .. 79

Figure 2-12 Natural fingermark deposited on ziplock bag and processed for 1.25 hours (top left), 1.5 hours (bottom left), 1.75 hours (top right), and two hours (bottom right), visualised at 590 nm with a 650 nm bandpass filter. .. 80

Figure 2-13 Average CAST scores calculated for latent fingermarks developed on PVDF and ziplock bag to compare eight different processing times. 81

Figure 2-14 Frequency of CAST scores for latent fingermarks developed on PVDF and ziplock bag to assess four shorter processing times. ... 81

Figure 2-15 Results for latent fingermarks on non-porous substrates (a-c) and semiporous substrates (d) organised by average CAST scores and working solutions. In Figure 2-15(d), glossy magazine was not processed with M4(BSA), while glossy cardboard was not processed with either M6 working solution. (aq = aqueous working solution; $BSA = aqueous working solution containing bovine serum albumin; $EG = a$$

glycolic working solution; $M# =$ multiplex solution containing $#$ antibodies/aptamers) ... 83

Figure 2-16 Blood-contaminated fingermarks on (a) garbage bag and (b) beverage can, both fixed with 5-SSA solution and processed with glycolic working solution with four components (top left), glycolic working solution with eight components (bottom left), aqueous working solution with four components (top right), and aqueous working solution with eight components (bottom right). Both developed fingermarks were visualised at 590 nm with a 650 nm bandpass filter. .. 85

Figure 2-17 Quartered blood-contaminated fingermark developed with a multiplex solution of six components on a light grey shopping bag with: (a) MeOH fixing/aqueous working solutions; (b) MeOH fixing/glycolic working solutions; (c) 5- SSA fixing/aqueous working solutions; and (d) 5-SSA fixing/glycolic working solutions. All quarter marks were visualised under an excitation of 590 nm with a 650 nm bandpass filter. .. 86

Figure 2-18 Average CAST score results comparing blood-contaminated fingermarks fixed with two different fixing solutions, but all processed with a multiplex solution of six components. (aq = aqueous working solution; $EG =$ glycolic working solution; $M#$ $=$ multiplex solution containing $#$ antibodies/aptamers; MeOH $=$ ice-cold methanol fixing solution used; 5-SSA = 5-sulfosalicylic acid fixing solution used) 87

Figure 2-19 Average CAST score results comparing blood-contaminated fingermarks fixed with two different fixing solutions and multiplex working solutions containing different number of components. (aq = aqueous working solution; $EG =$ glycolic working solution; $M#$ = multiplex solution containing # antibodies/aptamers; MeOH = ice-cold methanol fixing solution used; 5-SSA = 5-sulfosalicylic acid fixing solution used) .. 87

Figure 2-20 Comparison of aqueous (left halves) and glycolic (right halves) working solutions for fingermarks contaminated with (a) saliva on garbage bag; (b) semen on beverage can; and (c) saliva on glossy magazine. Saliva-contaminated fingermarks were visualised at 590 nm with a 650 nm bandpass filter, while the semencontaminated fingermark was visualised at 530 nm with a 610 nm bandpass filter. 89 Figure 2-21 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the comparison between aqueous and glycolic working solutions for saliva- and semen-contaminated fingermarks (blue and red columns, respectively). Cling film was not used for saliva-contaminated fingermarks. All fingermarks with no development were removed from this analysis, resulting in no usable data for salivacontaminated fingermarks on glossy magazine. A positive value favours the glycolic working solution. .. 90

Figure 2-22 Comparison of average CAST scores for corresponding latent fingermark halves. Non-porous substrates were only processed with the multiplex, CA, and CA \rightarrow R6G, while semi-porous substrates were processed with the multiplex, CA, CA \rightarrow IND-Zn, and CA → IND-Zn → PD.. 91

Figure 2-23 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the comparison between the multiplex solution with routine sequences for latent fingermarks. Assessments were performed under each technique's optimal viewing conditions (Table 2-2). Negative values correspond to a decrease in enhancement of the multiplex solution when compared to the routine technique sequence. The multiplex was compared to CA and $CA \rightarrow R6G$ on non-porous substrates, and CA, CA \rightarrow IND-Zn, and CA \rightarrow IND-Zn \rightarrow PD on semi-porous substrates. .. 93

Figure 2-24 Comparison of average CAST scores for blood-contaminated fingermark halves processed with the multiplex solution and routine technique sequences. 94

Figure 2-25 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the comparison between the multiplex solution with blood reagent sequences. Assessments were performed under each technique's optimal viewing conditions (Table 2-2). Negative values correspond to a decrease in enhancement of the multiplex solution when compared to the routine technique sequence. 95

Figure 2-26 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the comparison of $CA \rightarrow$ dyestain \rightarrow multiplex with $CA \rightarrow$ multiplex (blue) and $CA \rightarrow$ dyestain (red). Negative values favour $CA \rightarrow$ dyestain \rightarrow multiplex. ... 97 Figure 2-27 Average enhancement scores (for a comparative scale between -2 and +2) resulting from comparisons of routine (a) latent and (b) blood reagent sequences with and without the multiplex solution as a pre-treatment. Assessments were performed under each technique's optimal viewing conditions (Table 2-2). Negative values correspond to a decrease in enhancement of the sequences with the multiplex solutions when compared to the routine technique sequences alone. ... 99

Figure 2-28 Representative images of split fingermarks visualised under their respective viewing conditions depicting results for the multiplex solution as a pretreatment: (a) latent fingermark on a ziplock bag treated with multiplex \rightarrow CA (left) and CA only (right); and (b) blood-contaminated fingermark on a beverage can treated with CA \rightarrow multiplex \rightarrow AY7 (left) and CA \rightarrow AY7 (right). Each half illustrated was visualised and recorded under optimal viewing conditions for the last enhancement technique implemented (Table 2-2). ... 100

Figure 2-29 Average enhancement scores (for a comparative scale between -2 and +2) resulting from comparisons of routine (a) latent and (b) blood reagent sequences with and without the multiplex solution as a post-treatment. Assessments were performed under each technique's optimal viewing conditions (Table 2-2). Negative values correspond to a decrease in enhancement of the sequences with the multiplex solutions when compared to the routine technique sequences alone. ... 101

Figure 2-30 Representative images of fingermark halves visualised under their respective viewing conditions depicting results for the multiplex solution as a posttreatment: (a) latent fingermark half on a shopping bag treated with CA only (left) and then $CA \rightarrow$ multiplex (right); and (b) blood-contaminated fingermark half on a plastic bottle treated with CA \rightarrow AY7 (left) and then CA \rightarrow AY7 \rightarrow multiplex (right). Each half illustrated was visualised and recorded under optimal viewing conditions for the last enhancement technique implemented (Table 2-2).. 102

Figure 3-1 Schematic diagram of the SELEX process (Reproduced from Stoltenburg et al. [129]). ... 109

Figure 3-3 Polymerase chain reaction temperature program used. 126

Figure 3-4 Example PAGE gels from Round 11 visualised under (a) UV and (b) fluorescence. The bottom band outlined in red was cut out for subsequent desalting and DNA quantification. ... 127

Figure 3-5 Example UV-vis absorbance spectrum from Round 9 with a 50x dilution of DNA sample. ... 128

Figure 3-6 Graph showing percentage of DNA bound in each negative and positive selection round of fingermark-SELEX. Selection was monitored by the fluorophore attached to the DNA in all rounds except Round 1. The concentration of DNA was lowered in Rounds 2 and 3 to increase stringency. ... 134

Figure 3-7 Sequencing data distribution for select SELEX rounds. $#$ = selection round; N = negative selection round; P = positive selection round) 138

Figure 3-8 Squalene peak (highlighted in red box) was detected in Round 6 (top), but not in subsequent Rounds 7 (middle) and 8 (bottom). .. 140

Figure 3-9 Round 9 samples (fingermarks on garbage bag plastic) using DCM on nonpolar column (top), ECF on non-polar column (middle), and ECF on polar column (bottom). .. 141

Figure 3-10 Round 10 samples (fingermarks on copy paper) using ECF and run on a polar column (top) and non-polar column (bottom). .. 142

Figure 4-1 Charged fingermarks deposited on PVDF, aged for 24 hours, and then developed with aptamer-based reagent, each with a different lysozyme aptamer sequence (Reproduced from Wood et al. [195]). .. 148

Figure 4-2 Principle of nanoplasmonic imaging of latent fingermarks with and without cocaine present (Reproduced from Li et al.[196]). With respects to detection, a more acceptable term for "addicts" would be "users". .. 149

Figure 4-3 (a) Latent fingermarks deposited on marble and visualised with the aid of (b) FAM-labelled, (c) quantum dot-functionalised, and (d) UCNP-functionalised lysozyme-binding aptamer (Reproduced from Wang et al. [197]). 150

Figure 4-4 Overview of pilot study involving "fingermark" aptamer candidates from fingermark-SELEX process. ... 153

Figure 4-5 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the comparison between working solutions containing the corresponding short (i.e., 1, 2, 3, 4) and long (i.e., 5, 6, 7, 8, respectively) aptamer candidate sequences. Positive values correspond to an increase in enhancement with the working solution containing the longer aptamer candidate sequence when compared to that with its corresponding shorter aptamer candidate sequence. 168

Figure 4-6 Development obtained on PVDF with working solutions containing corresponding short (left) and long (right) aptamer candidate sequences under: (a) white light and (b) luminescence conditions (530 nm with 590 nm bandpass filter). 169

Figure 4-7 Three-day-old (top row) and 12-day-old (bottom row) fingermarks developed and observed under luminescence conditions (530 nm with 590 nm bandpass filter) on (from left to right) Australian garbage bag, Canadian garbage bag, Australian copy paper, and Canadian copy paper. .. 170

Figure 4-8 Average enhancement scores (for a comparative scale between -2 and $+2$) resulting from the direct comparison of pairs of longer aptamer candidate sequences. Positive values correspond to an increase in enhancement with the aptamer candidate sequence listed second in its pairing when compared to the aptamer candidate sequence listed first. .. 171

Figure 4-9 Examples of reverse development on Australian garbage bag (left) and ziplock bag (right) observed under luminescence conditions (530 nm with 590 nm bandpass filter). ... 172

Figure 4-10 Chemical spot test results for fresh working solution (Sequence 5) by Method 1 (top to bottom): 0.001 M, 0.01 M, and 0.1 M for (left to right): (a) L-serine, glycine, L-ornithine, L-alanine, L-threonine, L-histidine, L-valine, L-leucine, L- isoleucine, L-lysine, and L-phenylalanine with water control; and (b) stearic acid, oleic acid, squalene, palmitic acid*, and cholesterol* with dichloromethane and blank substrate controls. *Denotes chemical solutions prepared to 0.05 M instead of 0.1 M.

... 174

Figure 4-11 Distribution of CAST scores for larger donor population organised by aptamer candidate sequences per substrate. .. 176

Figure 4-12 Natural fingermarks from the larger donor population study illustrating various types of development: (a) 2-week-old fingermark aluminium foil with continuous ridge detail; (b) 3-day-old fingermark on plastic grocery bag with reverse development; and (c) 2-week-old fingermark on aluminium foil with spotty development indicating pore locations. All fingermarks were visualised at 530 nm with 590 nm bandpass filter. ... 176

Figure 4-13 Three-day-old natural fingermarks developed with Method 1 (left halves) and: (a) Method 2 on Australian garbage bag; (b) Method 3 on Australian garbage bag; and (c) Method 4 on aluminium foil. All fingermarks were visualised at 530 nm with 590 nm bandpass filter. ... 178

Figure 4-14 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the direct comparison of Method 1 to three other working-rinse solution combinations. Positive values correspond to an increase in enhancement with Method 1 when compared to the others.. 178

Figure 4-15 Comparison of CAST score frequency distribution between $CA \rightarrow R6G$ and working solutions containing "fingermark" aptamer candidates ("FM") per substrate... 180

Figure 4-16 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the direct comparison of working solutions containing "fingermark" aptamer candidates to $CA \rightarrow R6G$. Negative values correspond to a decrease in enhancement with the working solutions when compared to the routine sequence. .. 181

Figure 4-17 Distribution of CAST score frequency as percentages for each working solution per substrate, due to additional samples processed with "fingermark" aptamer candidate-containing reagent ("FM") and lysozyme aptamer-based reagent. 182

Figure 4-18 One-week-old natural fingermark half deposited by a male donor on aluminium foil, developed with the working solution containing Sequence #5, and visualised at 530 nm with a 590 nm bandpass filter. .. 183

Figure 4-19 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the direct comparison of working solutions containing "fingermark" aptamer candidates to those containing aptamers against cathepsin, IgE, or lysozyme. Positive values correspond to an increase in enhancement of the novel reagent when compared to the single-target aptamers... 184

Figure 4-20 One-week-old fingermark on aluminium foil with both halves imaged: (a) under optimal viewing conditions (e.g., exposure time adjusted) for both halves; and (b) at the same exposure time. (left half = working solution with "fingermark" aptamer candidate; right half = working solution with single-target aptamer) 185

Figure 4-21 Distribution of CAST score frequency as percentages for each working solution per substrate since "fingermark" aptamer candidate-containing reagent ("FM") processed all corresponding halves to the three single-target antibodies. 187

Figure 4-22 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the direct comparison of working solutions containing "fingermark" aptamer candidates to those containing anti- cathepsin D, anti-L-amino acid, or antiserotonin. Positive values correspond to an increase in enhancement of the novel reagent when compared to the single-target antibodies. ... 189

Figure 4-23 Normal development with working solution containing tris buffer (left) compared to reverse development with working solution containing PBS with non-fat milk (right). Fingermark visualised at 530 nm with 590 nm bandpass filter. 189

Figure 4-24 Distribution of CAST score frequency as percentages for each working solution per substrate since "fingermark" aptamer candidate-containing reagent("FM") processed all corresponding halves to the two multiplex solutions. (M4 and "M8" = multiplex solutions of four and seven antibodies, respectively) 190

Figure 4-25 Average enhancement scores (for a comparative scale between -2 and +2) resulting from the direct comparison of working solutions containing "fingermark" aptamer candidates to a multiplex solution of four (blue columns) and of seven (red columns) antibodies. Positive values correspond to an increase in enhancement of the novel reagent when compared to the multiplex solutions. .. 192

Figure 4-26 One-week-old fingermarks developed on: (a) Australian garbage bag with working solution containing "fingermark" aptamer candidate (left half) and multiplex of four antibodies (right half); and (b) grocery bag with working solution containing "fingermark" aptamer candidate (left half) and multiplex of seven antibodies (right half). Fingermarks visualised at 530 nm with 590 nm bandpass filter. 192

Figure X-1 Two-day-old sebaceous fingermark on PVDF developed with working solution containing Aptamer 1 (left) and Aptamer 2 (right) viewed under (a) white light and (b) 505 nm with 555 nm bandpass filter. \ldots \ldots

Figure XI-1 Three-day-old sebaceous fingermarks deposited on (a) Australian and (b) Canadian copy paper and subjected to the working solution for 15 (left halves) and 20 (right halves) seconds. Fingermarks were visualised at 530 nm with a 590 nm bandpass filter. 219

List of Tables

List of Abbreviations

Abstract

Over the past decade, there has been a resurgence of interest to design fingermark enhancement reagents capable of biomolecular recognition; such reagents would offer high selectivity and sensitivity, two areas where some believe improvement is desired with current fingermark detection methods. In addition to these, a high degree of adaptability for visualisation can be achieved with biomolecular recognition probes, such as antibodies and aptamers, allowing for the selection of the most appropriate visualisation wavelength for a particular luminescent probe or substrate without the need for sophisticated instrumentation or imaging systems. However, the major hurdle to overcome is the balance between sensitivity and selectivity. Single-target biomolecular recognition may be highly selective, purported to have better detection limits than chemical reactions or stains, and can provide information about identity and/or activity, but often results in incomplete ridge pattern development because only a fraction of the fingermark residue is being specifically targeted.

Consequently, the development and evaluation of multi-target biomolecular reagents for fingermark enhancement was investigated, with the focus on endogenous eccrine secretions. A variety of parameters (i.e., processing time, fixing and working solution conditions) were optimised on a wide range of non-porous and semi-porous substrates representative of casework materials to assess the suitability of the biomolecular reagents for potential operational use. The relative performance of biomolecular reagents was compared to that of routine methods applied to latent and body fluidcontaminated fingermarks. The incorporation of these novel reagents into routine technique sequences was also investigated. The experimental results indicated that the multi-target biomolecular reagents were not a suitable alternative to routine detection methods, did not provide any significant enhancement when included in routine sequences; however, they may still have potential for a niche application yet to be identified.

While a larger fraction of the fingermark was being targeted by multi-target reagents, the resulting development seemed to be influenced by inter-donor variability; it was unknown which combination of biomolecular recognition probes would be the most "universal". The focus of this research shifted to aptamers due to their many advantageous features over antibodies, one being their versatile *in vitro* selection process called Systematic Evolution of Ligands by EXponential enrichment or SELEX. Up to sixteen fingermark donors deposited variously aged natural fingermarks onto two realistic substrates (i.e., pooled target approach), which were then subjected to a novel SELEX variation termed fingermark-SELEX. Select DNA aptamer candidates, developed specifically against genuine fingermark residues, were subsequently incorporated into a fingermark enhancement reagent. The proof-ofconcept work demonstrated this novel reagent's ability to successfully develop friction ridge detail on non-porous substrates. Its relative performance was superior to that of single-target and multi-target biomolecular reagents previously designed within the same research group. This study has further opened up the possibilities of incorporating biomolecular recognition into fingermark detection methods by recognising and tapping into the potential of SELEX and resulting aptamer candidates in this forensic discipline.

Publications and Presentations

PEER-REVIEWED PUBLICATIONS

1. **Lam, R.**; Hofstetter, O.; Lennard, C.; Roux, C.; Spindler, X. (2016) Evaluation of Multi-Target Immunogenic Reagents for the Detection of Latent and Body Fluid-Contaminated Fingermarks. *For. Sci. Int.* 264, 168-175.

ORAL PRESENTATIONS (Presenter = Underlined)

- 1. **Lam, R.** Novel Fingermark Detection Methods Using Biomolecular Recognition. University of Technology Sydney Stage 3 Seminar, July 21, 2017, Broadway, NSW, Australia.
- 2. Spindler, X.; **Lam, R.**; Sullivan-Davenport, K.; Dilag, J.; Hofstetter, O.; Lennard, C.; Roux, C. Possibilities and Challenges in Using Biomolecular Recognition for Latent Fingermark Detection. Australian and New Zealand Forensic Science Society 23rd International Symposium on the Forensic Sciences, September 22, 2016, Auckland, New Zealand.
- 3. **Lam, R.** Universal Immunogenic Reagents for the Detection of Latent Fingermarks. University of Technology Sydney Stage 2 Seminar, July 8, 2016, Broadway, NSW, Australia.
- 4. **Lam, R.;** Ruscito, A.; DeRosa, M.C.; Spindler, X.; Lennard, C.; Roux, C. Fingermark-SELEX: A Novel Approach to Develop DNA Aptamers for Fingermark Detection. 60th Annual Conference of the Canadian Society of Forensic Sciences, May 19, 2016, Montreal, QC, Canada.
- 5. Spindler, X.; **Lam, R.**; Dilag, J.; Sullivan, K.; Hofstetter, O.; Lennard, C.; Roux, C. Optimisation and Evaluation of Multi-Target Biomolecular Reagents for Latent and Bloody Fingermark Detection: Latest Developments. International Fingerprint Research Group Meeting, October 23, 2015, Patiala, India.
- 6. Spindler, X.; **Lam, R.**; Dilag, J.; Lennard, C.; Roux, C. Next-Generation Fingermark Reagents: Molecular Recognition, Multispectral Imaging and Mapping. 7th European Academy of Forensic Science Conference, September 8, 2015, Prague, Czech Republic.
- 7. **Lam, R.**; Spindler, X.; Lennard, C.; Roux, C. Optimisation of Multi-Target Immunogenic Reagents and Comparison to Routine Detection Methods for Latent and Body Fluid-Contaminated Fingermarks. $7th$ European Academy of Forensic Science Conference, September 8, 2015, Prague, Czech Republic.
- 8. **Lam, R.**; Spindler, X.; Lennard, C.; Roux, C. Optimisation of Multi-Target Immunogenic Reagents and Comparison to Routine Detection Methods for Latent and Body Fluid-Contaminated Fingermarks. 4th Doctoral School of the École des Sciences Criminelles, August 26, 2015, Les Diablerets, Switzerland.
- 9. **Lam, R.** Optimisation of Multi-Target Immunogenic Reagents and Comparison to Routine Detection Methods for Latent and Body Fluid-Contaminated Fingermarks. UTS-UWS Forensic Science Research Student Forum, June 30, 2015, Penrith, NSW, Australia.

POSTER PRESENTATIONS (Presenter = Underlined)

1. **Lam, R.;** Ruscito, A.; DeRosa, M.C.; Spindler, X.; Lennard, C.; Roux, C. Fingermark-SELEX: A Novel Approach to Develop DNA Aptamers for Fingermark Detection. 21st Triennial Meeting of the International Association of Forensic Sciences 2017, August 24, 2017, Toronto, ON, Canada.

- 2. **Lam, R.;** Ruscito, A.; DeRosa, M.C.; Roux, C. "Fingermark" Aptamers: From Random Oligonucleotide Library to Fingermark Detection Reagent. International Association for Identification's 102nd International Forensic Educational Conference, August 8, 2017, Atlanta, GA, USA.
- 3. **Lam, R.**; Ruscito, A.; DeRosa, M.C.; Spindler, X.; Lennard, C.; Roux, C. Fingermark-SELEX: A Novel Approach to Developing Immunogenic Reagents for Fingermark Detection. Australian and New Zealand Forensic Science Society 23rd International Symposium on the Forensic Sciences, September 22, 2016, Auckland, New Zealand.