





Genome Sequences of Seven Foot-and-Mouth Disease Virus Isolates Collected from Serial Samples from One Persistently Infected Carrier Cow in Vietnam

Steven J. Pauszek, a Miranda R. Bertram, a,b Le T. Vu,c Ethan J. Hartwig, a George R. Smoliga, a Barbara Brito, a, b Carolina Stenfeldt, a, c Kimberley VanderWaal, c Ian H. Fish, a,b Vo V. Hung, d Nguyen T. Phuong, d Bui H. Hoang, d Luis L. Rodriguez, a Do H. Dung, e Donathan Arzta

Foreign Animal Disease Research Unit, Plum Island Animal Disease Center, ARS, USDA, Orient Point, New York, USAa; Oak Ridge Institute for Science and Education, PIADC Research Participation Program, Oak Ridge, Tennessee, USAb; STEMMA Laboratory, Veterinary Population Medicine, University of Minnesota, St. Paul, Minnesota, USAc; Regional Animal Health Office No. 6, Department of Animal Health, Ministry of Agriculture and Rural Development, Ho Chi Minh City, Vietnama; Department of Animal Health, Ministry of Agriculture and Rural Development, Hanoi, Vietname

ABSTRACT Several foot-and-mouth disease virus (FMDV) carrier cattle were identified in Vietnam by the recovery of infectious virus from oropharyngeal fluid. This report contains the first near-complete genome sequences of seven viruses from sequential samples from one carrier animal collected over the course of 1 year. The characterization of within-host viral evolution has implications for FMDV control strategies.

oot-and-mouth disease virus (FMDV; Aphthovirus, Picornaviridae) causes one of the most important transboundary livestock diseases. Acute FMD is characterized by the formation of characteristic vesicles on the feet and in the oral cavity (1-3). Following acute infection, a large proportion of FMDV-infected ruminants become persistently infected carriers, defined by the detection of FMDV in oropharyngeal fluid (OPF) beyond 28 days postinfection (4, 5). Transmission from carrier cattle to naive cattle has not been convincingly demonstrated (6-9). However, depopulation, quarantine, and trade restrictions are imposed subsequent to FMD outbreaks, due largely to the perceived risk of transmission from carrier animals (10–12).

In a recent study in Vietnam, FMDV O/ME-SA/PanAsia was the lineage most frequently recovered from persistently infected animals, and it was the most common lineage circulating in the region during the study (13, 14). Analysis of the VP1 sequences of serial isolates from persistently infected animals demonstrated genetic divergence between viruses isolated from individual animals over the course of 1 year (13, 14). However, the VP1 region covers only about 7.5% of the FMDV genome (15), and analysis of the whole genome is required for a more detailed elucidation of evolutionary changes that occur during persistent infection.

The viruses described herein, O/VIT/366/2012_pro, O/VIT/383/2012_pro, O/VIT/407/ 2012_pro, O/VIT/414/2012_pro, O/VIT/428/2013_pro, O/VIT/431/2013_pro, and O/VIT/ 433/2013_pro, were isolated from OPF samples collected from one FMDV carrier cow (Bos indicus) in Long An Province, Vietnam, between 7 June 2012 and 17 June 2013. All isolates belong to the O/ME-SA/PanAsia lineage. There was no clinical or molecular evidence of incursion of novel FMDV in the herd during the study period, and the most recent outbreak in the herd was reported to have occurred in 2010. Virus isolation was

Received 10 July 2017 Accepted 12 July 2017 Published 24 August 2017

Citation Pauszek SJ, Bertram MR, Vu LT, Hartwig EJ, Smoliga GR, Brito B, Stenfeldt C, VanderWaal K, Fish IH, Hung VV, Phuong NT, Hoang BH, Rodriguez LL, Dung DH, Arzt J. 2017. Genome sequences of seven foot-andmouth disease virus isolates collected from serial samples from one persistently infected carrier cow in Vietnam. Genome Announc 5:e00849-17. https://doi.org/10.1128/ genomeA.00849-17.

This is a work of the U.S. Government and is not subject to copyright protection in the United States, Foreign copyrights may apply. Address correspondence to Jonathan Arzt, jonathan.arzt@ars.usda.gov.

enameAnnouncements^T Pauszek et al.

achieved from all samples by a single passage in LFBK- $\alpha_V \beta_6$ cell culture (16), as previously described (17). Viral RNA was extracted, and the partial 5' untranslated region (UTR), complete open reading frame (ORF), and partial 3' UTR were covered with three overlapping reverse transcription-PCR (RT-PCR) amplicons and were Sanger sequenced, as previously described (18). Chromatogram analysis and consensus sequence identification were performed with Sequencher version 5.4.6, as previously described (19). These sequences code for the complete 6,999-nucleotide (nt) ORF, 115 to 116 nt in the 5' UTR, and 27 to 45 nt in the 3' UTR. Among the seven sequences, the pairwise nucleotide and amino acid differences within the ORF region ranged from 26 to 147 nt, and 2 to 19 amino acids (aa), respectively. There was one insertion in the 5' UTR of the isolate O/VIT/428/2013_pro which was not observed in temporally subsequent isolates. Across these samples, 25 sites exhibit nonsynonymous substitutions. These observations suggest a dynamically evolving viral population in carrier animals.

To our knowledge, this is the first report of near-complete sequences of FMDVs isolated from sequential samples obtained from one persistently infected individual under natural conditions. The characterization of these viruses provides insights into within-host evolution of FMDV during persistent infection and has implications for FMD control in areas that are endemic for the disease. Further characterization of FMDV evolution within carriers will help clarify the role of persistently infected animals in FMD epidemiology and this potential source of outbreaks and emergence of novel viral strains.

Accession number(s). The genome nucleotide sequences of O/VIT/366/2012_pro, O/VIT/383/2012_pro, O/VIT/407/2012_pro, O/VIT/414/2012_pro, O/VIT/428/2013_pro, O/VIT/431/2013_pro, and O/VIT/433/2013_pro described herein have been deposited in GenBank under the accession numbers MF143572 to MF143578, respectively. The versions described in this paper are the first versions, MF143572.1 to MF143578.1.

ACKNOWLEDGMENTS

This research was funded in part by ARS-CRIS project 1940-32000-061-00D. Additional funding was provided by the Cooperative Biological Engagement Program of the U.S. Department of Defense, Defense Threat Reduction Agency.

REFERENCES

- 1. Arzt J, Juleff N, Zhang Z, Rodriguez LL. 2011. The pathogenesis of foot-and-mouth disease I: viral pathways in cattle. Transbound Emerg Dis 58:291-304. https://doi.org/10.1111/j.1865-1682.2011.01204.x.
- 2. Arzt J, Baxt B, Grubman MJ, Jackson T, Juleff N, Rhyan J, Rieder E, Waters R, Rodriguez LL. 2011. The pathogenesis of foot-and-mouth disease II: viral pathways in swine, small ruminants, and wildlife; myotropism, chronic syndromes, and molecular virus-host interactions. Transbound Emerg Dis 58:305-326. https://doi.org/10.1111/j .1865-1682.2011.01236.x.
- 3. Grubman MJ, Baxt B. 2004. Foot-and-mouth disease. Clin Microbiol Rev 17:465-493. https://doi.org/10.1128/CMR.17.2.465-493.2004.
- 4. Van Bekkum JG, Frenkel HS, Frederiks HHJ, Frenkel S. 1959. Observations on the carrier state of cattle exposed to foot-and-mouth disease virus. Bull Off Int Epizoot 51:917-922.
- 5. Sutmoller P, Gaggero A. 1965. Foot-and-mouth disease carriers. Vet Rec 77:968-969. https://doi.org/10.1136/vr.77.33.968.
- 6. Moonen P, Jacobs L, Crienen A, Dekker A. 2004. Detection of carriers of foot-and-mouth disease virus among vaccinated cattle. Vet Microbiol 103:151-160. https://doi.org/10.1016/j.vetmic.2004.07.005.
- 7. Parthiban AB, Mahapatra M, Gubbins S, Parida S. 2015. Virus excretion from foot-and-mouth disease virus carrier cattle and their potential role in causing new outbreaks. PLoS One 10:e0128815. https://doi.org/10 .1371/journal.pone.0128815.
- 8. Bengis RG, Thomson GR, Hedger RS, De Vos V, Pini A. 1986. Foot-andmouth disease and the African buffalo (Syncerus caffer). 1. Carriers as a source of infection for cattle. Onderstepoort J Vet Res 53:69-73.
- 9. Ilott MC, Salt JS, Gaskell RM, Kitching RP. 1997. Dexamethasone inhibits virus production and the secretory IgA response in oesophagealpharyngeal fluid in cattle persistently infected with foot-and-mouth

- disease virus. Epidemiol Infect 118:181–187. https://doi.org/10.1017/ S0950268896007376.
- 10. Bouma A, Elbers ARW, Dekker A, de Koeijer A, Bartels C, Vellema P, van der Wal P, van Rooij EMA, Pluimers FH, de Jong MCM. 2003. The foot-and-mouth disease epidemic in the Netherlands in 2001. Prev Vet Med 57:155-166. https://doi.org/10.1016/S0167-5877(02)00217-9.
- 11. Mansley LM, Donaldson Al, Thrusfield MV, Honhold N. 2011. Destructive tension: mathematics versus experience—the progress and control of the 2001 foot and mouth disease epidemic in Great Britain. Rev Sci Tech 30:483-498. https://doi.org/10.20506/rst.30.2.2054.
- 12. Junker F, Ilicic-Komorowska J, van Tongeren F. 2009. Impact of animal disease outbreaks and alternative control practices on agricultural markets and trade: the case of FMD. OECD food, agriculture and fisheries papers, no 19. OECD Publishing, Paris, France.
- 13. de Carvalho Ferreira HC, Pauszek SJ, Ludi A, Huston CL, Pacheco JM, Le VT, Nguyen PT, Bui HH, Nguyen TD, Nguyen T, Nguyen TT, Ngo LT, Do DH, Rodriguez L, Arzt J. 2017. An integrative analysis of foot-and-mouth disease virus carriers in Vietnam achieved through targeted surveillance and molecular epidemiology. Transbound Emerg Dis 64:547-563. https://doi.org/10.1111/tbed.12403.
- 14. Brito B, Pauszek SJ, Eschbaumer M, Stenfeldt C, de Carvalho Ferreira HC, Vu LT, Phuong NT, Hoang BH, Tho ND, Dong PV, Minh PQ, Long NT, King DP, Knowles NJ, Dung DH, Rodriguez LL, Arzt J. 2017. Phylodynamics of foot-and-mouth disease virus O/PanAsia in Vietnam 2010-2014. Vet Res 48:24. https://doi.org/10.1186/s13567-017-0424-7.
- 15. Carrillo C, Tulman ER, Delhon G, Lu Z, Carreno A, Vagnozzi A, Kutish GF, Rock DL. 2005. Comparative genomics of foot-and-mouth disease virus. J Virol 79:6487-6504. https://doi.org/10.1128/JVI.79.10.6487-6504.2005.
- 16. LaRocco M, Krug PW, Kramer E, Ahmed Z, Pacheco JM, Duque H, Baxt B,

- Rodriguez LL. 2013. A continuous bovine kidney cell line constitutively expressing bovine $\alpha_V \beta_6$ integrin has increased susceptibility to foot-and-mouth disease virus. J Clin Microbiol 51:1714–1720. https://doi.org/10.1128/JCM.03370-12.
- 17. Pacheco JM, Arzt J, Rodriguez LL. 2010. Early events in the pathogenesis of foot-and-mouth disease in cattle after controlled aerosol exposure. Vet J 183:46–53. https://doi.org/10.1016/j.tvjl.2008.08.023.
- 18. Arzt J, Brito BP, Pauszek SJ, Hartwig EJ, Smoliga GR, Vu LT, Vu PP,
- Stenfeldt C, Rodriguez LL, Long NT, Dung DH. 2017. Genome sequence of foot-and-mouth disease virus serotype O lineage Ind-2001d collected in Vietnam in 2015. Genome Announc 5(18):e00223-17. https://doi.org/10.1128/genomeA.00223-17.
- Pacheco JM, Piccone ME, Rieder E, Pauszek SJ, Borca MV, Rodriguez LL. 2010. Domain disruptions of individual 3B proteins of foot-and-mouth disease virus do not alter growth in cell culture or virulence in cattle. Virology 405:149–156. https://doi.org/10.1016/j.virol.2010.05.036.