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| 1  | Transport of small and neutral solutes through reverse osmosis  |                             |
|----|---|-----------------------------|
| 2  | membranes: Role of skin layer conformation of   | Formatted: Font color: Auto |
| 3  | the polyamide film  |                             |
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# 21 Abstract

22 The polyamide skin layer of reverse osmosis (RO) membranes was characterised using 23 advanced and complementary analytical techniques to investigate the mechanisms underlying 24 the permeation of contaminants of emerging concern in potable water reuse - N-25 nitrosodimethylamine (NDMA) and N-nitrosomethylethylamine (NMEA). This study used 26 five RO membrane samples with similar membrane properties. The five RO membrane samples 27 spanned over a large range of water permeance (0.9-5.8 L/m<sup>2</sup>hbar) as well as permeation of 28 NDMA (9-66%) and NMEA (3-29%). Despite such distinct these differences among the five 29 RO membranes, characterisations of the skin layer using positron annihilation lifetime spectroscopy, atomic force microscopy and field emission scanning electron microscopy 30 31 revealed almost no variation difference-in their free-volume hole-radius (0.270-0.275 nm), 32 effective surface area (198-212%) and thickness (30-35 nm) of the skin layer. The results 33 suggest that there could be other RO skin layer properties, such as the interconnectivity of the 34 protuberances within the polyamide skin layer additional to the free-volume hole-size and 35 thickness of the skin layer, which can also govern water and solute permeation.

Keywords: free-volume hole; *N*-nitrosodimethylamine; positron annihilation lifetime
spectroscopy; potable reuse; reverse osmosis.

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## 39 1. Introduction

40 N-nitrosodimethylamine (NDMA;  $C_2H_6N_2O$ ) and N-nitrosomethylethylamine (NMEA; 41 C<sub>3</sub>H<sub>8</sub>N<sub>2</sub>O) are micropollutants of significant concern in potable water reuse since they are 42 probable carcinogenic chemicals [1]. With a molecular weight of 74 g/mol, NDMA is the 43 smallest in the N-nitrosamine group. NDMA and NMEA are neutral and hydrophilic compounds at environmental pH (i.e. pH 6-8). Although reverse osmosis (RO) membrane 44 45 separation can achieve excellent rejection of a range of impurities in reclaimed water including salts, macro-organics, and many micropollutants, the rejection of NDMA, NMEA and several 46 47 other N-nitrosamines is low and highly variable because of its small molecular size and lack of 48 charge [2-5]. Thus, they are often detectable in RO permeate at concentrations higher than guideline or target values set by water authorities around the world. For example, California 49 50 has established a notification level of 10 ng/L for NDMA and a public health goal of 3 ng/L 51 [6]. Similarly, in Australia, the guideline value of NDMA in water intended for potable reuse 52 has been also set at 10 ng/L [7]. The low and highly variable separation performance of RO 53 with respect to NDMA rejection necessitates post treatment by advanced oxidation (UV 54 irradiation and H<sub>2</sub>O<sub>2</sub>) [8]. Recent research [9] suggests that NDMA rejection by RO 55 membranes varies significantly amongst the many RO membranes available on the market. Thus, further insights which lead to better membrane selection and improvement of the 56 57 separation performance of RO for N-nitrosamine removal can directly contribute to the 58 economic viability and public safety of potable water reuse.

Given the significant importance of low molecular weight micropollutants in potable reuse, numerous previous studies have been conducted to reveal the permeation mechanisms of micropollutants through RO membranes [10-13]. The significance of steric (size) interaction between solutes and the free-volume holes within the RO membrane active skin layer has been Field Code Changed

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64 molecular size (e.g. minimum molecular width or projected area) of uncharged solutes and 65 their removals by RO has been established [14, 15]. Nevertheless, mechanisms underlying the 66 difference in NDMA permeation among RO membranes are still poorly understood from the 67 perspective of membrane properties. This is mainly due to analytical limitations in 68 characterising the membrane skin layer at sub-nanometre scale resolution.

clearly demonstrated from the viewpoint of solute properties. A strong correlation between

63

69 The free-volume holes – holes in the membrane skin layer in polymeric matrix – are thought 70 to play an important role in water and solute transport through the RO membrane. Nevertheless, 71 findings to date remain inconclusive. The free-volume hole size can be determined by positron 72 annihilation lifetime spectroscopy (PALS) [16]. Previous measurements of the free-volume 73 hole-radius of commercial RO membranes [16-18] varied considerably from 0.20 and 0.29 nm. 74 Several previous studies [18-20] have demonstrated a strong correlation between solute 75 permeation and free-volume hole-size. In contrast, no clear correlation between solute 76 permeation and free-volume hole-size was reported by several other studies [17, 21, 22]. To 77 date, there have been very few PALS studies on the characterisation of RO membranes due to 78 the limited availability of slow positron beam based instruments.

79 The thickness of the skin layer has also been considered as an important property of an RO 80 membrane governing water and solute transport. According to the solution-diffusion model 81 [23-25], permeation of solutes and solution through RO membranes occurs via their penetration 82 into the membrane material and diffusion through the RO membrane. The key role of the 83 polymeric membrane thickness on solute permeation can also be supported by the finding that 84 the thickness is inversely proportional to water permeance and there is a trade-off between 85 water permeance and solute selectivity [26, 27]. This mechanism is plausible, however, it is 86 difficult to validate for commercial RO membranes. Recent RO membranes are designed with

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87 a rougher membrane surface that holds a higher surface area and a higher permeance [28-32]. 88 As a result, they commonly have a so-called "ridge-and-valley" structure and hollow interior 89 of crumpled nodules throughout the surface of the skin layer [29]. The entire thickness of the 90 skin layer of commercial RO membranes is about 200-300 nm as reported by previous studies 91 using transmission electron microscopy (TEM) [33-36]. The crumpled film forming the 92 crumpled structure of the skin layer and the flat film comprising the interface between 93 polyamide and polysulfone layers both have a thickness as low as 20 nm [31, 37, 38]. 94 Characterisation using a field emission - scanning electron microscope (FE-SEM) [37] allows 95 for a quantification of each polyamide film. Nevertheless, due to the complex inner structure 96 and rough surface morphology of the RO membrane skin layer and its variation among RO 97 membranes, it is still difficult to identify the location of the most important polyamide films 98 and quantify their thickness for comparison among different RO membranes.

99 A systematic evaluation of several RO membranes with similar chemical ingredients can 100 potentially yield new insights into the role of the membrane skin layer on solute permeation. 101 This study aims to identify key structure parameters that govern NDMA transport in the RO 102 process by characterising the skin layer properties of five RO membranes that have similar 103 chemical composition and skin layer structure. State-of-the-art analytical techniques including 104 PALS, FE-SEM and atomic force microscopy (AFM) were used for the characterisation of the 105 skin layer including free-volume hole-size, thickness and surface area, respectively.

## 106 2. Materials and methods

#### 107 2.1. Chemicals

| 108 | Analytical grade NDMA and N-nitrosomethylethylamine (NMEA) (Table 1) were purchased          |
|-----|--|
| 109 | from Ultra Scientific (Kingstown, RI, USA). All stock solutions were prepared in methanol to |

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- 110 obtain 1 µg/mL of each chemical and were stored at 4 °C in the dark. Both chemicals can be
- 111 classified as hydrophilic and neutral (uncharged) at environmental pH (pH 6 to 8) [39].

112 **Table 1** – Structure and properties of the selected *N*-nitrosamines.

| Compound   | NDMA         | NMEA  |
|--|--------------|---|
| Structure  | н₃с−√сн₃     | № <sup>0</sup><br>H <sub>3</sub> с_NСН <sub>3</sub> |
| Molecular formula  | $C_2H_6N_2O$ | C <sub>3</sub> H <sub>8</sub> N <sub>2</sub> O      |
| Molecular weight [g/mol]                                   | 74.05        | 88.06   |
| Log  D at pH8 <sup>a</sup> [-]                             | 0.04         | 0.4   |
| pKa at pH8ª [-]  | 3.5          | 3.4   |
| Minimum projection area <sup>a, b</sup> [nm <sup>2</sup> ] | 0.20         | 0.22  |

<sup>a</sup>Chemicalize (http://www.chemicalize.org).

<sup>b</sup> Minimum projection area is the area of the compound projected with the minimum plane of

115 its circular disk, based on the van der Waals radius.

#### 116 2.2. Membranes and membrane treatment system

117 Two commercially available RO membranes – namely ESPA2 and ESPAB – and a prototype

118 RO membrane were obtained as flat sheet samples from Hydranautics/Nitto (Osaka, Japan).

119 The active skin layers of these membranes have similar chemical ingredients although the

120 detailed information is proprietary. The ESPA2 membrane has been employed in many potable

121 water reuse schemes [14], while the ESPAB membrane is designed for boron removal and has

122 been widely used in the second pass of RO seawater desalination plants. In addition, samples

123 of the ESPAB and Prototype membranes were also subjected to heat treatment to alter the

124 physical properties. These heat-treated samples are designated as heated ESPAB and heated

125 Prototype, respectively. Thus, in total, five different membrane samples were used in this

126 investigation.

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#### 127 2.3. Experimental protocols

# 128 2.3.1. Heat treatment

Heat treatment was conducted by heating the RO membrane coupons in 80 °C ultrapure water solution. The RO membrane coupons were first rinsed with ultrapure water (18.0 MΩcm). Thereafter, each coupon was stored in a 200 mL beaker filled with ultrapure water, and the beakers were placed in a temperature-controlled water bath (SWB-11A, AS ONE, Osaka, Japan) that maintained the water temperature at 80 °C. After 4 h of heat treatment, the membrane coupons were rinsed with ultrapure water and stored at 4 °C in the dark.

#### 135 2.3.2. RO filtration experiments

136 The separation of NDMA and NMEA by each RO membrane was evaluated in ultrapure water 137 using the bench-scale cross-flow RO system (Fig. S1). Filtration experiments were started with 138 permeance evaluation in which RO membrane filtration experiments were conducted with 139 ultrapure water at 2000 kPa to measure the pure water permeance. Thereafter, NDMA and 140 NMEA stock solution was added to obtain 200 ng/L of each compound in the feed solution. 141 The membrane system was operated at a 20 L/m<sup>2</sup>h permeate flux and 20 °C feed temperature. 142 Concentrations of NDMA and NMEA were determined by high-performance liquid 143 chromatography-photochemical reaction-chemiluminescence as described in our previous 144 studies [40, 41]. The passage is defined as  $R = 100 \times C_p/C_f$ , where  $C_p$  and  $C_f$  are solute 145 concentration in the permeate and feed, respectively.

# 146 2.4. Membrane characterisations

- 147 2.4.1. Surface chemistry
- 148 Major functional groups of RO membranes was analysed using Fourier transform infrared
- 149 spectroscopy (FTIR) spectrophotometer (Nicolet iS5, Thermo Fisher Scientific, Waltham, MA,
- 150 USA) in attenuated total reflection (ATR) method. The RO membrane samples were freeze-

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| 151 | dried for 24 hours using a freeze drier (FD-1000, Tokyo Rikakikai, Tokyo, Japan). The                                       |                    |    |
|-----|---|--------------------|----|
| 152 | spectrum was obtained in the range of 400-4000 cm <sup>-1</sup> at 1 cm <sup>-1</sup> resolution.                           |                    |    |
| 153 | 2.4.2. Positron annihilation lifetime spectroscopy (PALS)   |                    |    |
| 154 | The free-volume hole-radius of each RO membrane was measured using PALS with a slow   |                    |    |
| 155 | positron beam that is housed at the National Institute of Advanced Industrial Science and                                   |                    |    |
| 156 | Technology (AIST) in Tsukuba, Japan. Details of PALS for the analysis of RO membranes are                                   |                    |    |
| 157 | provided elsewhere [9]. The radius of free-volume hole $(r)$ of the skin layer in RO membranes                              | Field Code Changed | 1  |
| 158 | was determined from the pick-off annihilation lifetime of <i>ortho</i> -positronium ( $\tau_{o-Ps}$ ) using the             |                    |    |
| 159 | Tao-Eldrup model [42, 43]:  | Field Code Changed | i) |
| 160 | $\tau_{o-Ps} = 0.5 \left[ 1 - \frac{r}{r+0.166} + \frac{1}{2\pi} \sin\left(\frac{2\pi r}{r+0.166}\right) \right]^{-1} $ (1) | Field Code Changed | 1  |
| 161 | where $r (< 1 \text{nm})$ is approximated as a spherical shape. Positron irradiation was carried out                        |                    |    |
| 162 | under vacuum (~ $10^{-5}$ Pa) and about $2 \times 10^{6}$ positron annihilation events were collected for the               |                    |    |
| 163 | positron lifetime spectrum of each sample. Spectra were analysed using a non-linear least-                                  |                    |    |
| 164 | squares fitting program. Unless otherwise stated, the incident energy $(E_{in})$ was set at 1.0 keV,                        |                    |    |
| 165 | which corresponds to a mean implantation depth of 31 nm from the top (implantation depth                                    |                    |    |
| 166 | range = 0–90 nm) with a material density of 1.3 g/cm <sup>3</sup> (Fig. 1). This incident energy was                        |                    |    |
| 167 | selected according to previous studies [18, 19] that revealed the lowest free-volume hole-radius                            | Field Code Changed | 1  |
| 168 | of polyamide RO at 1.0 keV. The dry material density of RO membranes (1.3 g/cm <sup>3</sup> ) was                           |                    |    |
|     |   |                    |    |
| 169 | determined based on the data reported by Kolev and Freger [44].   | Field Code Changed | 1  |

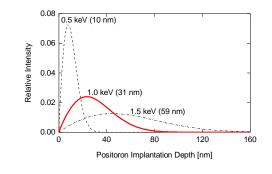


Fig. 1 – Distribution of positron implantation depth in a polyamide membrane sample with a
material density of 1.3 g/cm<sup>3</sup> at a positron incident energy of 0.5, 1.0 or 1.5 keV. The mean
implantation depth at each positron incident energy is shown in brackets.

174 2.4.3. Atomic force microscopic analysis

170

175 Membrane surface area was analysed using an atomic force microscope (AFM) (MFP-3D-SA, 176 Asylum Research - Oxford Instruments Company, CA, USA). Membranes underwent sample 177 pre-treatment steps involving the replacement of water in the membranes with tert-Butyl 178 alcohol followed by freeze drying. Images were obtained in air using tapping mode with a 179 silicate cantilever. The scanning area was 5  $\mu$ m  $\times$  5  $\mu$ m. The effective surface area of each membrane was calculated based on the data of three samples. Effective surface area here was 180 defined as a ratio between the actual (measured) area and the sample area as described in the 181 182 following formula:

183 Effecticve surface area [%] = 
$$\frac{Actual area [\mu m^2]}{sample area [\mu m^2]} \times 100$$
 (2)

### 184 2.4.4. Field emission scanning electron microscopic analysis

185 Cross-sectional images of the RO membrane skin layer were attained using a field emission -186 scanning electron microscope (FE-SEM) instrument (S-4800, Hitachi, Japan) at 3 kV 187 accelerating voltage. Cross-sections of each membrane sample were prepared by freezefracturing wet membranes in liquid nitrogen followed by air drying. Thereafter, the target cross-sectional area was coated with conductive material prior to the analysis. The thickness of the skin layer of each RO membrane was calculated by determining the average of 5 different locations.

# 192 **3. Results and discussion**

# 193 **3.1.** Solute permeation

194 The five RO membrane samples examined in this study spanned over a large range of water 195 permeance as well as solute passage with respect to both NDMA and NEMA (Fig. 2). Heat 196 treatment was effective to reduce solute passage and water permeance. After heat treatment, NDMA passage through the ESPAB and the Prototype membranes decreased from 56 to 37%197 198 and from 18 to 9%, respectively. The pure water permeance of these membrane also 199 proportionally decreased as can be seen from Fig. 2. A strong linear correlation between solute 200 passage with respect to both NDMA and NMEA and water permeance can be confirmed in Fig. 2. As noted in section 2.2, all five RO membrane samples were from the same manufacturer 201 202 with similar chemical ingredients of the active skin layer. Thus, data from Fig. 2 allow for a 203 systematic investigation of the role of the active skin layer in transport of small and neutral 204solutes as well as water across the membrane.

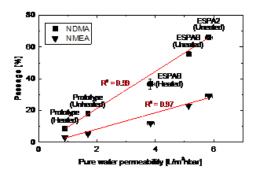




Fig. 2 – Correlation between the passage of NDMA and NMEA in pure water, and pure water permeance (feed temperature =  $20.0 \pm 0.1$  °C and permeate flux =  $20 \text{ L/m}^2\text{h}$ ). Error bars show the range of two replicate experiments.

# 209 3.2. Characterisations of the RO skin layer

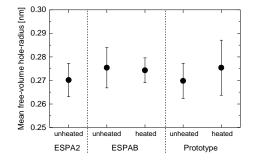
210 3.2.1. Membrane chemistry

211 Variation in the top layer chemistry of RO membranes can be qualitatively evaluated by 212 examining the FTIR spectra [45, 46]. Notable peaks for fully aromatic polyamide were 213 observed at 1668, 1608 and 1539 cm<sup>-1</sup> that correspond to C=O and C-N stretching and C-C-N 214 deformation vibration (amide I), N-H deformation vibration and C=C ring stretching vibration 215 of aromatic amide, and N-H in-place bending and N-C stretching vibration of a -CO-NH- group 216 (amide II), respectively [47] (Fig. S2). Peaks at 1586, 1505, 1488 and 1245 cm<sup>-1</sup> can be 217 assignable to polysulfone. The ratio in peak intensity between 1668 cm<sup>-1</sup> (C=O stretching of 218 the amide group formed by the reaction between diamine and acid chloride) and 1245 cm<sup>-1</sup> (C-219 O stretching of the polysulfone support) was 0.21, 0.21 and 0.17 for ESPA2, ESPAB and 220 Prototype, respectively. This indicates that these RO membranes have a-similar chemical 221 propertycomposition. In contrast, It is noted that heat treatment increased the peak intensity 222 ratio from 0.21 to 0.30 and from 0.17 to 0.18 for ESPAB and Prototype membrane, respectively. 223 The cause of the changes in the peak intensity for ESPAB after heat treatment remains unclear, 224 but it will be in the scope of our future study.

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### 225 3.2.2. Free-volume hole-radius

226 The mean free-volume hole-radius of the selected RO membranes was determined at a mean 227 implantation depth of 31 nm using  $\tau_{o-Ps}$  values (pick-off annihilation lifetime of o-Ps) from 228 PALS analysis (Table S3). The free-volume hole-radius of the three unheated RO membranes 229 (i.e. ESPA2, ESPAB and Prototype) was almost identical, ranging from 0.270 to 0.275 nm (Fig. 230 3). Heat treatment did not show any discernible impact on the free-volume hole-radius. It is 231 noteworthy that PALS analysis at other implantation depths (i.e. 10 and 59 nm) of the ESPAB 232 membrane did not show any significant variation in the free-volume hole-radius due to heat 233 treatment (Fig. S4). It is noted that current PALS technique cannot confirm a small difference 234 in free-volume hole-radius of RO membranes less than 0.01 nm due to the inherent errors in 235 PALS and the inhomogeneity of the membrane samples. Thus, the free-volume hole-radius of 236 all five membrane samples in Fig. 3 are considered to be similar.



237

Fig. 3 – Free-volume hole-radius of the five RO membranes. The data here is the average and
range of two replicates.

The cross-sectional areas of spherical free-volume holes with radii of 0.270 and 0.275 nm are 0.23 and 0.24 nm<sup>2</sup>, respectively. These values are comparable to the minimum projection area of NDMA (0.20 nm<sup>2</sup>) and NMEA (0.22 nm<sup>2</sup>) (**Table 1**). Because the passage of NDMA and NMEA varied with a difference of only 0.02 nm<sup>2</sup> in the minimum projection area of the two molecules, a variation of 0.01 nm<sup>2</sup> in free-volume hole-area among the five RO membranes may still be an important factor. However, there was no observable correlation between the measured free-volume hole-radius and the passage of NDMA and NMEA (**Fig. S5**). Given the similar free-volume hole-size of the five membrane samples, these results suggest that a factor other than the free-volume hole-size can also govern the permeation of NDMA and NMEA by these RO membranes.

250 3.2.3. Effective surface area

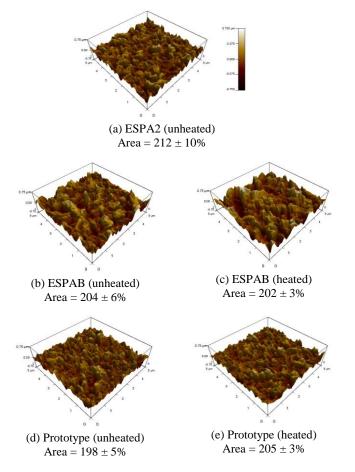
251 The effective membrane surface area was determined by taking into account the topography of 252 the RO skin layer at the microscopic level (i.e. surface roughness) using AFM. Indeed, at the 253 microscopic level, the effective membrane surface area can differ considerably from the surface 254 area normally used to calculate the permeate flux [9]. It is noted that permeate flux 255 considerably influences NDMA permeation [13]. Since the skin layer can play an important 256 role in solute permeation through the RO membrane as proposed in literature [30, 38], it is 257 important to take into account the effective membrane surface area when assessing solute 258 permeation.

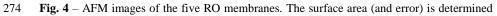
259 Despite the large variation in the visualized "ridge-and-valley" structure among the three 260 different types of RO membranes (i.e. ESPA2, ESPAB and Prototype), their effective surface 261 area was almost identical, ranging from 198 and 212% (Fig. 4). In other words, the effective 262 membrane surface area at the microscopic level is approximately two times the plain area. 263 Likewise, heat treatment did not cause any discernible changes in the effective surface area. 264 Results from Fig. 4 confirm that separation experiments in this study were also at the same permeate flux for a systematic comparison of solute permeation among all selected RO 265 membranes. More importantly, the observation of the large variation in permeance (Fig. 2) and 266 267 almost identical effective surface area (Fig. 4) among the RO membranes suggests that in this Field Code Changed
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- study, water permeance is not influenced by the effective surface area of the RO skin layer. It
- 269 is noteworthy that the surface area determined by AFM does not necessarily represent the entire
- 270 surface area. Surface areas through which feed solution can penetrate include the backside of
- 271 bent protuberance and confined rumpled films with packed protuberances that cannot be
- 272 measured by AFM [32]. Therefore, actual surface area taking account of all morphology is

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273 necessary to conclusively determine the role of surface area for water permeance.





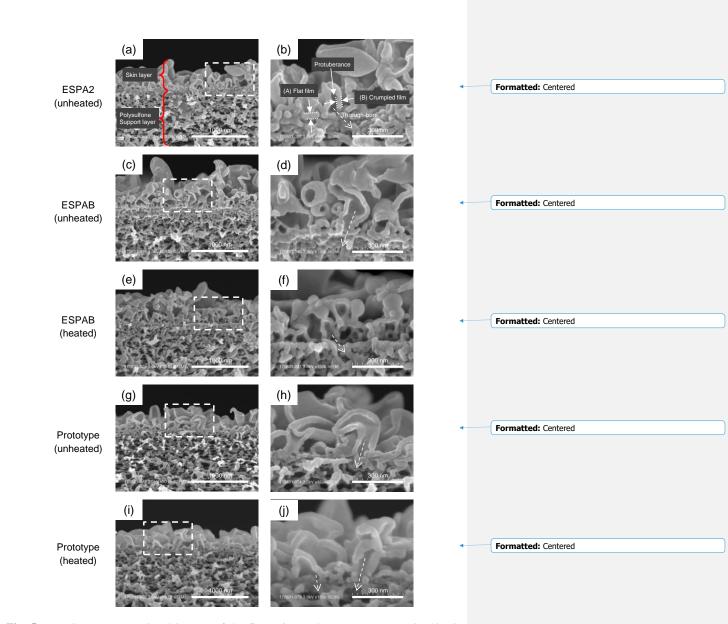
275 from the average (and measurement variation) of three membrane coupons.

276 3.2.4. Thickness

277 The thickness of the skin layer of the selected RO membranes was evaluated using a cross-278 sectional view obtained by FE-SEM. The FE-SEM analysis (Fig. 5) revealed a distinctive 279 "ridge-and-valley" structure and a hollow interior of crumpled nodules throughout the skin 280 layer of all RO membranes investigated in this study. These complex structures are similar to 281 other commercial RO membranes recently reported in literature [36-38]. The estimated 282 thickness of the skin layer was 300-400 nm for ESPA2 and ESPAB RO membranes and 200-283 300 nm for Prototype RO membranes. There was no apparent change in the skin layer thickness 284 after heat treatment. The skin layer of these membranes comprised of two major polyamide 285 films (A) a flat polyamide film that forms a film base at the interface with the polysulfone 286 support layer and (B) a crumpled polyamide film that forms crumpled nodules with internal hollow structure (also called as protuberance) (Fig. 5). Ultrahigh resolution FE-SEM images 287 288 in Fig. 5 show a similar thickness between the flat polyamide film and crumpled polyamide 289 film. In addition, it appears that the crumpled polyamide film is almost always on top of the 290 flat polyamide film. Using SEM, Yan et al. [37] also reported that the ESPA2 RO membrane 291 has a through-bore that interconnects the cavity of the protuberance and open structure of the 292 polysulfone support layer. The interconnections can also be found at the other four RO 293 membranes used in this study. The majority of the protuberances did not appear to have 294 interconnections. Although high water permeance of RO membranes (e.g. ESPA2) can be 295 associated with the number of the interconnections, it was not possible to quantify the 296 connectivity through the FE-SEM cross-sectional images.

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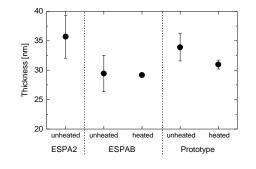


298 Fig. 5 – FE-SEM cross-sectional images of the five RO membranes: (a, c, e, g, i) skin &

299 polysulfone layers (scale = 1000 nm) and (b, d, f, h, j) areas enclosed with dash lines in the

300 skin & polysulfone layers (scale = 300 nm).

301 Given the importance of the crumpled polyamide films on water and solute permeation [30, 302 37], the crumpled film thickness was measured for all the RO membranes. It is noted that the 303 values were attained through two representative FE-SEM images; thus, there could be a 304 variation in thickness throughout the RO membranes. The results revealed that unheated 305 ESPA2, ESPAB and Prototype RO membranes had similar crumpled film thickness of 36, 29, 306 and 34 nm, respectively (Fig. 6). Results in Fig. 6 indicate that there was no discernible 307 variation in thickness among the three RO membranes in this study. In addition, heat treatment 308 did not appear to alter the crumpled film thickness. Indeed, differences in the crumpled film 309 thickness between heated and unheated samples were within the measurement error margin (i.e. 310 standard deviation of two samples of the same membrane). As a result, in this study, variation 311 in water flux and the passage of NDMA and NMEA cannot be attributed to the measured 312 crumpled film thickness (Fig. S7), suggesting that other skin layer properties such as 313 conformation of the crumpled polyamide films could also govern their permeation.



314

Fig. 6 – Thickness of the crumpled film of the five RO membranes. The thickness was
determined based on two RO membrane coupons, each of which was measured at 5 locations
(Table S6).

### 318 3.3. Discussions

Comprehensive analysis of the skin layer of five RO membrane samples using PALS, FE-SEM and AFM revealed that there could be other RO skin layer properties besides the free-volume hole-radius and thickness of the crumpled film that can govern water and solute permeation. This is a significant finding in membrane transport, because the free-volume hole-radius and thickness of the crumpled film have often been considered the only membrane properties governing the membrane transport.

325 FE-SEM images obtained here identified that the free-volume hole-radius analysed by PALS was likely to result from the crumpled polyamide film. According to the distribution of positron 326 327 implantation depth (Fig. 1), most positrons of PALS at a positron incident energy of 1 keV 328 were expected to have annihilated within the crumpled polyamide films that have the thickness 329 of about 30 nm. The flat polyamide film is mostly covered by several layers of the crumpled 330 film and it is 300 to 400 nm away from the top ridge (Fig. 5). Thus, the flat polyamide film 331 located at the interface with the polysulfone supporting layer is not accessible by positrons with 332 1 keV. Although the flat polyamide film beneath the crumpled films can be reached by 333 increasing the positron incident energy (Fig. 1), increasing the incident energy also broadens 334 out the positron implantation depth distribution, meaning that signals can be obtained from 335 both the flat and crumpled films. As a result, thickness of the flat film that has far less surface 336 area than the crumpled film cannot be accurately determined. If the flat polyamide film plays 337 an important role in solute permeation, a sample preparation method that enables us to 338 preferentially analyse the flat polyamide films should be developed.

In regard to the flat polyamide film, perhaps water permeance and separation performance of
the selected RO membranes is related to the degree of the interconnections between the cavity
of the protuberance and the open structure of the polysulfone support layer. In other words, RO

342 membranes with less interconnections have less water permeance but high separation capability. 343 The importance of hollow protuberance on the water permeance of RO membranes has been 344 confirmed by Pacheco et al. [48] who evaluated 3D images of two commercial RO membranes 345 using TEM tomography technique. The FE-SEM cross-sectional images in Fig. 5 revealed that 346 not all of the protuberances have interconnections toward the polysulfone support layer. 347 Protuberances without interconnections allow the solute and solvent (water) to permeate 348 through two barriers - crumpled film and flat film, which could reduce water permeance but 349 improve the separation performance. The projected area TEM technique previously reported 350 by Pacheco et al. [36] and Yan et al. [37] has the potential to visualize the structure inside the 351 ridges including the interconnections from its top view, but the correlation between the 352 interconnections and separation performance has not yet been quantified. 353 There are some other limitations and challenges for the characterization of the RO skin layer 354 with respect to PALS analysis. The analysis here was conducted under dry conditions, while 355 wet RO membranes may have swelling effects [49], which can expand the polymer network

and alter the water permeance and selectivity [50, 51]. In addition, the size distribution of freevolume holes may be more important than mean free-volume hole-radius, because the narrower passages connecting the major free-volume holes could actually determine solute permeations as suggested by Dražević et al. [52]. More accurate analysis with wet membrane samples and the determination of size distribution of free-volume holes requires a significant improvement in PALS method, thus, it is a scope of our future study.

In addition to the swelling effects, chemical properties of the internal skin layer could be a major contributor to a variation in diffusion coefficient and sorption coefficient of water and solutes, which ultimately leads to a variation in their water permeance and separation performance. Typically, increases in the degree of polyamide cross-linking can cause less water Formatted: Font color: Auto

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and solutes to sorb onto the polymer due to restriction in swelling effects [53]. This could cause a decrease in both effective water and salt diffusion coefficients [54], leading to a decrease in water permeance but an increase in selectivity  $(D_w/D_s)$  (trade-off theory) [26]. If that is the case, chemical properties of the skin layer such as cross-linking levels should be more important than the physical properties analysed in this study (i.e. free-volume hole-radius and thickness) to determine the major skin layer properties. 

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### 372 4. Conclusions

373 This study shows that RO membranes with distinct separation performance and water 374 permeance can have similar crumpled film properties including free-volume hole-size and 375 thickness. PALS, AFM and FE-SEM revealed that major crumpled polyamide film properties 376 such as free-volume hole-size, effective surface area and thickness are almost identical among 377 three different types of unheated RO (ESPA2, ESPAB and Prototype) and two heated RO 378 (ESPAB and Prototype). The results suggest that there exist at least another RO skin layer 379 property other than the free-volume hole-size and thickness that can also govern the transport 380 of water and small and neutral solutes such as NDMA and NMEA that are of signifincant 381 concern in potable water reuse. Such property is likely to be the protuberance conformation or 382 interconnectivity of the protuberance within the membrane polyamide skin layer. In addition, 383 FE-SEM data also reveal that current PALS technique may not be suitable for determining free-volume hole-radius of the flat polyamide film located at the interface between the 384 385 polyamide skin and the polysulfone supporting layer beneath the crumpled polyamide films. 386 Further advances in analytical technique that allows for the quantification of interconnections 387 between the protuberances and the polysulfone support layer, the comprehensive 388 characterisation of RO membranes by PALS (size distribution and wet conditions), and the

389 evaluation of chemical properties of the polyamide films is necessary to fully decode the

390 permeation mechanism of NDMA.

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### 397 6. References

- US Environmental Protection Agency, Integrated Risk Information System (IRIS), N nitrosodimethylamine. Office of Research and Development (ORD), National Center
   for Environmental Assessment (1987) www.epa.gov/iris/subst/0045.htm, in, 1987.
- 401 [2] M.J. Farré, K. Döderer, L. Hearn, Y. Poussade, J. Keller, W. Gernjak, Understanding
  402 the operational parameters affecting NDMA formation at Advanced Water Treatment
  403 Plants, J. Hazard. Mater., 185 (2011) 1575-1581.
- 404 [3] C. Bellona, J.E. Drewes, G. Oelker, J. Luna, G. Filteau, G. Amy, Comparing nanofiltration and reverse osmosis for drinking water augmentation, Journal AWWA, 406 100 (2008) 102-116.
- 407 [4] T. Fujioka, S.J. Khan, J.A. McDonald, A. Roux, Y. Poussade, J.E. Drewes, L.D.
  408 Nghiem, *N*-nitrosamine rejection by reverse osmosis membranes: A full-scale study,
  409 Water Res., 47 (2013) 6141-6148.
- 410 [5] M.H. Plumlee, M. López-Mesas, A. Heidlberger, K.P. Ishida, M. Reinhard, N411 nitrosodimethylamine (NDMA) removal by reverse osmosis and UV treatment and
  412 analysis via LC-MS/MS, Water Res., 42 (2008) 347-355.
- 413 [6] CDPH, NDMA and other Nitrosamines Drinking water issues, California Department
   414 of Public Health 2011.
- 415 [7] NRMMC, EPHC, AHMC, Australian guidelines for water recycling: Managing health
  416 and environmental risks (Phase 2): Augmentation of drinking water supplies,
  417 Environment Protection and Heritage Council, National Health and Medical Research
  418 Council, Natural Resource Management Ministerial Council, Canberra, 2008.
- 419 [8] H.L. Leverenz, G. Tchobanoglous, T. Asano, Direct potable reuse: A future imperative,
  420 Journal of Water Reuse and Desalination, 1 (2011) 2-10.

- 421 [9] T. Fujioka, N. Oshima, R. Suzuki, W.E. Price, L.D. Nghiem, Probing the internal 422 structure of reverse osmosis membranes by positron annihilation lifetime spectroscopy: 423 Gaining more insight into the transport of water and small solutes, J. Membr. Sci., 486 424 (2015) 106-118.
- A.R.D. Verliefde, S.G.J. Heijman, E.R. Cornelissen, G.L. Amy, B. Van der Bruggen,
  J.C. van Dijk, Rejection of trace organic pollutants with high pressure membranes
  (NF/RO), Environ. Prog., 27 (2008) 180-188.
- 428 [11] C. Bellona, J.E. Drewes, P. Xu, G. Amy, Factors affecting the rejection of organic solutes during NF/RO treatment - A literature review, Water Res., 38 (2004) 2795-2809.
- L.D. Nghiem, P.J. Coleman, NF/RO filtration of the hydrophobic ionogenic compound
  triclosan: Transport mechanisms and the influence of membrane fouling, Sep. Purif.
  Technol., 62 (2008) 709-716.
- T. Fujioka, L.D. Nghiem, S.J. Khan, J.A. McDonald, Y. Poussade, J.E. Drewes, Effects
  of feed solution characteristics on the rejection of *N*-nitrosamines by reverse osmosis
  membranes, J. Membr. Sci., 409–410 (2012) 66-74.
- T. Fujioka, S.J. Khan, Y. Poussade, J.E. Drewes, L.D. Nghiem, *N*-nitrosamine removal
  by reverse osmosis for indirect potable water reuse A critical review based on
  observations from laboratory-, pilot- and full-scale studies, Sep. Purif. Technol., 98
  (2012) 503-515.
- Y. Kiso, K. Muroshige, T. Oguchi, M. Hirose, T. Ohara, T. Shintani, Pore radius estimation based on organic solute molecular shape and effects of pressure on pore radius for a reverse osmosis membrane, J. Membr. Sci., 369 (2011) 290-298.
- 443 [16] S.H. Kim, S.-Y. Kwak, T. Suzuki, Positron annihilation spectroscopic evidence to 444 demonstrate the flux-enhancement mechanism in morphology-controlled thin-film-445 composite (TFC) membrane, Environ. Sci. Technol., 39 (2005) 1764-1770.
- T. Fujioka, N. Oshima, R. Suzuki, S.J. Khan, A. Roux, Y. Poussade, J.E. Drewes, L.D.
  Nghiem, Rejection of small and uncharged chemicals of emerging concern by reverse
  osmosis membranes: The role of free volume space within the active skin layer, Sep.
  Purif. Technol., 116 (2013) 426-432.
- Z. Chen, K. Ito, H. Yanagishita, N. Oshima, R. Suzuki, Y. Kobayashi, Correlation study
  between free-volume holes and molecular separations of composite membranes for
  reverse osmosis processes by means of variable-energy positron annihilation
  techniques, J. Phys. Chem. C, 115 (2011) 18055-18060.
- K. Ito, Z. Chen, W. Zhou, N. Oshima, H. Yanagishita, R. Suzuki, Y. Kobayashi,
  Subnanoscopic holes in composite membranes for desalination elucidated by energytunable positron annihilation, Jpn. J. Poly. Sci. Technol., 69 (2012) 443-447.
- M. Henmi, Y. Fusaoka, H. Tomioka, M. Kurihara, High performance RO membranes
  for desalination and wastewater reclamation and their operation results, Water Sci.
  Technol., 62 (2010) 2134-2140.

- T. Sasaki, H. Tomioka, K. Nakatsuji, Composite semipermeable membrane, production
  process thereof, and element, fluid separation equipment and treatment method for
  boron-containing water using the same, in: U.S. Patent (Ed.) United States Patent,
  Toray Industries, Inc., USA, 2010.
- T. Fujioka, N. Oshima, R. Suzuki, M. Higgins, W.E. Price, R.K. Henderson, L.D.
  Nghiem, Effect of heat treatment on fouling resistance and the rejection of small and neutral solutes by reverse osmosis membranes, Water Sci. Technol. Water Supply, 15 (2015) 510-516.
- 468 [23] J.G. Wijmans, R.W. Baker, The solution-diffusion model: a review, J. Membr. Sci.,
  469 107 (1995) 1-21.
- 470 [24] S. Bandini, L. Bruni, 2.04 Transport Phenomena in Nanofiltration Membranes, in: E.
  471 Drioli, L. Giorno (Eds.) Comprehensive Membrane Science and Engineering, Elsevier,
  472 Oxford, 2010, pp. 67-89.
- J. Wang, D.S. Dlamini, A.K. Mishra, M.T.M. Pendergast, M.C.Y. Wong, B.B. Mamba,
  V. Freger, A.R.D. Verliefde, E.M.V. Hoek, A critical review of transport through
  osmotic membranes, J. Membr. Sci., 454 (2014) 516-537.
- 476 [26] G.M. Geise, H.B. Park, A.C. Sagle, B.D. Freeman, J.E. McGrath, Water permeability
  477 and water/salt selectivity tradeoff in polymers for desalination, J. Membr. Sci., 369
  478 (2011) 130-138.
- 479 [27] H. Ju, A.C. Sagle, B.D. Freeman, J.I. Mardel, A.J. Hill, Characterization of sodium chloride and water transport in crosslinked poly(ethylene oxide) hydrogels, J. Membr.
  481 Sci., 358 (2010) 131-141.
- 482 [28] M. Hirose, H. Ito, Y. Kamiyama, Effect of skin layer surface structures on the flux
  483 behaviour of RO membranes, J. Membr. Sci., 121 (1996) 209-215.
- 484 [29] J.R. Werber, C.O. Osuji, M. Elimelech, Materials for next-generation desalination and
   485 water purification membranes, Nature Reviews: Materials, 1 (2016) 16018.
- 486 [30] S. Karan, Z. Jiang, A.G. Livingston, Sub–10 nm polyamide nanofilms with ultrafast
   487 solvent transport for molecular separation, Science, 348 (2015) 1347-1351.
- L. Lin, R. Lopez, G.Z. Ramon, O. Coronell, Investigating the void structure of the polyamide active layers of thin-film composite membranes, J. Membr. Sci., 497 (2016) 365-376.
- 491 [32] M.M. Kłosowski, C.M. McGilvery, Y. Li, P. Abellan, Q. Ramasse, J.T. Cabral, A.G.
  492 Livingston, A.E. Porter, Micro-to nano-scale characterisation of polyamide structures
  493 of the SW30HR RO membrane using advanced electron microscopy and stain tracers,
  494 J. Membr. Sci., 520 (2016) 465-476.
- 495 [33] A.K. Ghosh, B.-H. Jeong, X. Huang, E.M.V. Hoek, Impacts of reaction and curing 496 conditions on polyamide composite reverse osmosis membrane properties, J. Membr. 497 Sci., 311 (2008) 34-45.

- 498 [34] C.Y. Tang, Y.-N. Kwon, J.O. Leckie, Characterization of humic acid fouled reverse
   499 osmosis and nanofiltration membranes by transmission electron microscopy and
   500 streaming potential measurements, Environ. Sci. Technol., 41 (2006) 942-949.
- [35] V. Freger, Swelling and morphology of the skin layer of polyamide composite
   membranes: an atomic force microscopy study, Environ. Sci. Technol., 38 (2004) 3168 3175.
- [36] F.A. Pacheco, I. Pinnau, M. Reinhard, J.O. Leckie, Characterization of isolated
   polyamide thin films of RO and NF membranes using novel TEM techniques, J. Membr.
   Sci., 358 (2010) 51-59.
- 507 [37] H. Yan, X. Miao, J. Xu, G. Pan, Y. Zhang, Y. Shi, M. Guo, Y. Liu, The porous structure
  508 of the fully-aromatic polyamide film in reverse osmosis membranes, J. Membr. Sci.,
  509 475 (2015) 504-510.
- [38] Y. Li, M.M. Kłosowski, C.M. McGilvery, A.E. Porter, A.G. Livingston, J.T. Cabral,
   Probing flow activity in polyamide layer of reverse osmosis membrane with
   nanoparticle tracers, J. Membr. Sci., 534 (2017) 9-17.
- [39] B. Van der Bruggen, A. Verliefde, L. Braeken, E.R. Cornelissen, K. Moons, J.Q.J.C.
  Verberk, H.J.C. van Dijk, G. Amy, Assessment of a semi-quantitative method for
  estimation of the rejection of organic compounds in aqueous solution in nanofiltration,
  J. Chem. Technol. Biotechnol., 81 (2006) 1166-1176.
- 517 [40] T. Fujioka, H. Takeuchi, H. Tanaka, L.D. Nghiem, K.P. Ishida, H. Kodamatani, A rapid
   518 and reliable technique for *N*-nitrosodimethylamine analysis in reclaimed water by
   519 HPLC-photochemical reaction-chemiluminescence, Chemosphere, 161 (2016) 104 520 111.
- 521 [41] H. Kodamatani, H. Yamasaki, T. Sakaguchi, S. Itoh, Y. Iwaya, M. Saga, K. Saito, R.
  522 Kanzaki, T. Tomiyasu, Rapid method for monitoring N-nitrosodimethylamine in
  523 drinking water at the ng/L level without pre-concentration using high-performance
  524 liquid chromatography-chemiluminescence detection, J. Chromatogr. A, 1460 (2016)
  525 202-206.
- M. Eldrup, D. Lightbody, J.N. Sherwood, The temperature dependence of positron lifetimes in solid pivalic acid, Chem. Phys., 63 (1981) 51-58.
- 528 [43] S.J. Tao, Positronium annihilation in molecular substances, J. Chem. Phys., 56 (1972)
   5499-5510.
- 530 [44] V. Kolev, V. Freger, Hydration, porosity and water dynamics in the polyamide layer
   531 of reverse osmosis membranes: A molecular dynamics study, Polymer, 55 (2014) 1420 532 1426.
- M. Bass, V. Freger, Facile evaluation of coating thickness on membranes using ATR FTIR, J. Membr. Sci., 492 (2015) 348-354.
- E. Dražević, K. Košutić, V. Freger, Permeability and selectivity of reverse osmosis
   membranes: Correlation to swelling revisited, Water Res., 49 (2014) 444-452.

- 537 [47] C.Y. Tang, Y.-N. Kwon, J.O. Leckie, Effect of membrane chemistry and coating layer
   538 on physiochemical properties of thin film composite polyamide RO and NF
   539 membranes: I. FTIR and XPS characterization of polyamide and coating layer
   540 chemistry, Desalination, 242 (2009) 149-167.
- [48] F. Pacheco, R. Sougrat, M. Reinhard, J.O. Leckie, I. Pinnau, 3D visualization of the internal nanostructure of polyamide thin films in RO membranes, J. Membr. Sci., 501 (2016) 33-44.
- 544 [49] J. Lee, C.M. Doherty, A.J. Hill, S.E. Kentish, Water vapor sorption and free volume in
  545 the aromatic polyamide layer of reverse osmosis membranes, J. Membr. Sci., 425–426
  546 (2013) 217-226.
- 547 [50] E.P. Chan, A.P. Young, J.-H. Lee, J.Y. Chung, C.M. Stafford, Swelling of ultrathin
  548 crosslinked polyamide water desalination membranes, J. Polym. Sci., Part B: Polym.
  549 Phys., 51 (2013) 385-391.
- Y.-H. Huang, W.-C. Chao, W.-S. Hung, Q.-F. An, K.-S. Chang, S.-H. Huang, K.-L.
  Tung, K.-R. Lee, J.-Y. Lai, Investigation of fine-structure of polyamide thin-film composite membrane under swelling effect by positron annihilation lifetime spectroscopy and molecular dynamics simulation, J. Membr. Sci., 417–418 (2012) 201-209.
- E. Dražević, K. Košutić, V. Kolev, V. Freger, Does Hindered Transport Theory Apply
   to Desalination Membranes?, Environ. Sci. Technol., 48 (2014) 11471-11478.
- [53] G.M. Geise, D.R. Paul, B.D. Freeman, Fundamental water and salt transport properties
   of polymeric materials, Prog. Polym. Sci., 39 (2014) 1-42.
- T.A. Jadwin, A.S. Hoffman, W.R. Vieth, Crosslinked poly(hydroxyethyl methacrylate)
  membranes for desalination by reverse osmosis, J. Appl. Polym. Sci., 14 (1970) 13391359.
- 562