

1 **Does plant species selection in functional active green walls influence VOC**
2 **phytoremediation efficiency?**

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4 **Peter J Irga^{ab, 1}, Thomas Pettit^b, Robert F Irga^{ab}, Naomi J Paull^b, Ashley NJ**
5 **Douglas^b, Fraser R Torpy^b**

6 ^a School of Civil and Environmental Engineering, Faculty of Engineering and
7 Information Technology, University of Technology Sydney

8

9 ^b Plants and Environmental Quality Research Group, School of Life Sciences, Faculty
10 of Science, University of Technology Sydney, P.O. Box 123, Broadway, Sydney, NSW
11 2007, Australia

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13 ¹ **Corresponding author:** Peter Irga

14

E-mail: Peter.Irga@uts.edu.au

15

Phone: +61 2 9514 9063

16

17 **ORCID Numbers:** Peter Irga – 0000-0001-5952-0658

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Fraser Torpy – 0000-0002-9137-6948

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20

21 **Abstract**

22 Volatile organic compounds (VOCs) are of public concern due to their adverse health
23 effects. Botanical air filtration is a promising technology for reducing indoor air
24 contaminants, but the underlying mechanisms are not fully understood. This study
25 assessed active botanical biofilters for their single-pass removal efficiency (SPRE) for
26 benzene, ethyl acetate and ambient total volatile organic compounds (TVOC)s, at
27 concentrations of *in situ* relevance. Biofilters containing four plant species
28 (*Chlorophytum orchidastrum*, *Nematanthus glabra*, *Nephrolepis cordifolia* ‘duffii’ and
29 *Schefflera arboricola*) were compared to discern whether plant selection influenced
30 VOC SPRE. Amongst all tested plant species, benzene SPREs were between 45.54–
31 59.50%, with *N. glabra* the most efficient. The botanical biofilters removed 32.36–
32 91.19% of ethyl acetate, with *C. orchidastrum* and *S. arboricola* recording significantly
33 higher ethyl acetate SPREs than *N. glabra* and *N. cordifolia*. These findings thus
34 indicate that plant type influences botanical biofilter VOC removal. It is proposed that
35 ethyl acetate SPREs were dependent on hydrophilic adsorbent sites, with increasing
36 root surface area, root diameter and root mass all associated with increasing ethyl
37 acetate SPRE. The high benzene SPRE of *N. glabra* is likely due to the high wax
38 content in its leaf cuticles. The SPREs for the relatively low levels of ambient TVOCs
39 were consistent amongst plant species, providing no evidence to suggest that *in situ*
40 TVOC removal is influenced by plant choice. Nonetheless, as inter-species differences
41 do exist for some VOCs, botanical biofilters using a mixture of plants is proposed.

42

43 **Keywords** Air pollution; botanical biofilters; phytoremediation; green walls; VOCs

44

45 **Introduction**

46 Air pollution in urban environments is a growing concern, with exposure directly linked
47 to seven million deaths globally in 2012 (WHO 2014). One major component of urban
48 air pollution is volatile organic compounds (VOCs), which include a range of organic
49 compounds that quickly vaporise at room temperature, such as ethyl acetate (EtOAc)
50 and benzene. VOC exposure can be damaging to human health, with complex and long-
51 term compounding effects that are difficult to treat (Deng et al. 2015, Deng et al. 2017).

52 Given increasing pollution concentrations and population densities in metropolitan
53 areas, there is a growing need to develop methods to maintain habitable urban living
54 environments (Irga et al. 2018). Urban developers are adopting sustainability
55 frameworks that require the employment of strategies to limit or mitigate pollution, and
56 demonstrate a positive impact on the environment (De Valck et al. 2019). As social and
57 technological changes are leading to increases in the proportion of time individuals
58 spend in indoor environments (Vardoulakis et al. 2015), the quality of indoor air is
59 becoming an increasingly important health factor. Indoor air quality is maintained
60 primarily by heating, ventilation and air conditioning (HVAC) systems, which have
61 variable control over the indoor atmospheric chemosphere (Irga and Torpy 2016).

62 The combined application of biotechnology, environmental engineering and
63 horticultural science has led to the development of biological air filters as a promising
64 avenue of research for the bioremediation of indoor air (Soreanu 2016). These systems
65 use natural bioagents (plants and/or microorganisms) to remove pollutants from the air
66 through an aerobic process, where the pollutants act as energy, carbon, and other
67 nutritional sources for the bioagents, or are otherwise absorbed into or adsorbed on to
68 the biological materials (Wei et al. 2017). Additionally, these systems can use
69 substrates containing a proportion of activated carbon and a range of other materials, to
70 assist with pollutant filtration or substrate microbial growth (Pettit et al. 2018, Torpy et
71 al. 2018). The efficiency with which biofilters can filter out and degrade VOCs from
72 indoor air indicates that they may be used to reduce inhabitant pollutant exposure
73 (Sriprapat and Thiravetyan 2013, Wolverton et al. 1984, Wood et al. 2006, Brillì et al
74 2018).

75 While static systems such as pot plants have been found to be ineffective for high
76 capacity contaminant removal (Llewellyn and Dixon 2011), research indicates that
77 'active green walls', which utilise mechanical assistance to funnel air into the biofilter
78 substrate, improves their bioremediation efficiency to the extent that functional air
79 remediation is probable (Torpy et al. 2015). These systems may also be practical for
80 large infrastructure use, given that they are accessible, robust, cost-effective and have
81 a low-energy footprint. Although the available types of botanical biofiltration systems
82 differ in design, they all use active airflow facilitated with devices such as impellers
83 that increase the airflow across or through the systems and therefore allow larger
84 volumes of air to be processed by the biofilter. Whilst there is a growing body of
85 literature that demonstrates the air pollutant remediation capabilities of this technology
86 (Pettit et al. 2019), to date, the potential for plant selection to enhance botanical
87 biofilters ability to filter some of the more dangerous air pollutants is required.

88 Plant selection is known to have an influence on VOC removal efficiency for static,
89 potted-plant systems (e.g. Kim et al. 2010). Whilst the nature of the plant characteristics
90 that determine these effects have yet to be resolved, there may be phylogenetic
91 associations where certain groups of plants are more effective for the removal of certain

92 forms of VOC (Kim et al. 2016). Whilst it has been shown that rhizospheric bacteria
93 are the major agents of removal for some VOCs (eg. Wood et al. 2002), there are clearly
94 plant-associated effects that may or may not (Irga et al. 2017) interact with the substrate
95 microbial community, or even subsume its activity for specific VOCs, as is the case for
96 CO₂ removal (Pettit et al. 2017). An alternate hypothesis is that different plants can
97 affect the abiotic chemical or physical properties of the substrate such that VOC
98 removal is altered (Deng and Deng 2018). Despite many years of research, these
99 patterns have yet to be resolved, and thus objective decisions on the most effective plant
100 species for VOC biofiltration cannot be made.

101 Previous work that has tested the removal of multiple VOCs has usually tested
102 pollutants with similar physio-chemical properties, for example numerous studies have
103 assessed VOCs focusing on benzene, toluene, ethyl benzene and xylene (BTEX).
104 Darlington et al. (2001) compared the biofilter removal rates of toluene, ethyl benzene
105 and *o*-xylene, finding that all compounds had similar removal rates, and suggested that
106 the limiting factor that affected VOC removal rates was transfer of gaseous pollutants
107 to the liquid phase, rather than microbial degradation. However, for higher
108 concentrations of more soluble compounds, which are easily absorbed into the liquid
109 phase, the rate of microbial degradation may be the primary limiting factor (Pettit et al
110 2018).

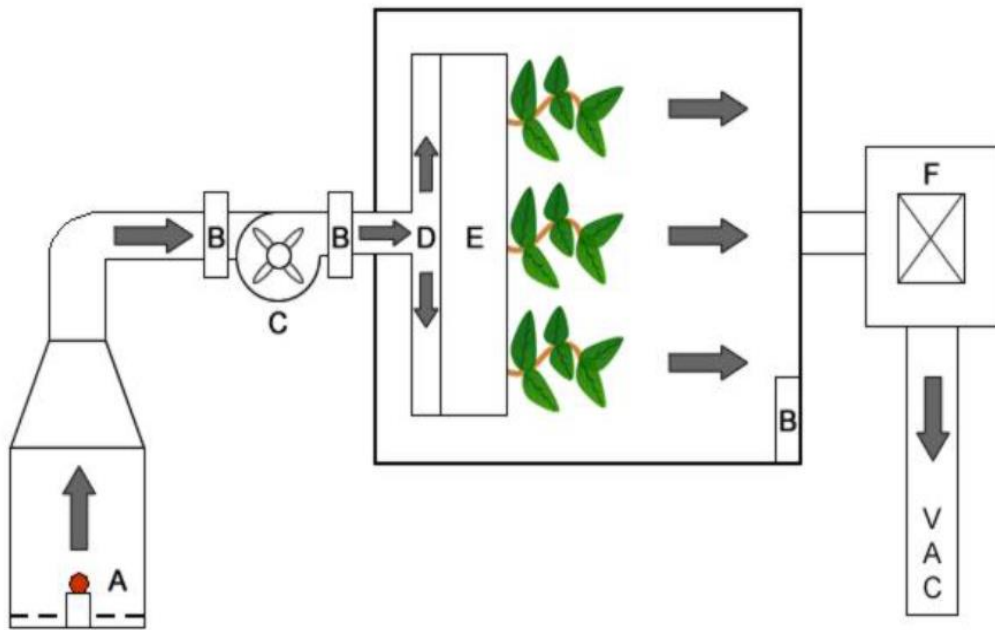
111 The current study investigates a range of common green wall plant species in an active
112 botanical biofilter to elucidate the influence of plant type on VOC removal efficiency.
113 This is the first study that compares the SPRE of VOCs with the explicit aim to identify
114 the most efficient plant wall species for active green wall VOC biofiltration. Further,
115 assessments were made to determine the correspondence between VOC filtration
116 efficiency and a range of plant and substrate characteristics so as to identify traits that
117 may be associated with increased VOC removal rates.

118 **Methods**

119 *Active living wall biofilter design*

120 The current study assessed an active green wall system previously described in Irga et
121 al. (2017; Figure 1). Briefly, the system utilizes assisted aeration by incorporating an
122 axial impeller to both increase gaseous pollutant exposure to the substrate and plant
123 rhizosphere, and to allow for particulate matter removal, which is filtered through the
124 substrate. The system is modular, allowing for flexibility in upscale design, with
125 module dimensions of 500 x 500 x 130 mm, with 16 circular compartments for plant
126 insertion. The module is constructed from polyethylene and contains a coconut coir
127 based substrate. When operational, air is drawn into the system, and flows through the
128 plant substrate matrix (25 L total substrate volume), contained within a tight weave
129 high density polyethylene (HDPE) bag – typically used as shade cloth, and returned to
130 the environment through the planted surface. Total airflow through the 0.25 m² front
131 surface area test system was 14.90 L/s (Abdo et al. 2016). This green wall system has
132 been previously demonstrated to be effective in the removal of VOCs, CO₂ and
133 particulate matter in laboratory trials (Pettit et al. 2017).

134



135

136 **Fig. 1. Schematic of the single pass removal efficiency test apparatus set up: A**
 137 **Combustion chamber; B Digital pressure differential sensor; C Axial impeller; D**
 138 **Plenum within system module; E Biofilter packing medium; F Photoionisation**
 139 **detector; VAC Exhaust vacuum pump.**

140

141 *Plant materials and measurement of plant morphological traits*

142 Four plant species were selected for this study (Figure 2), which are all commonly used
 143 vertical biowall plants, that encompass a range of phylogenetic, physiological and
 144 morphological variability. These plants were selected as they have previously been used
 145 in biowalls for phytoremediation assessments (Pettit et al. 2017). The four species
 146 tested were: (i) *Chlorophytum orchidastrum* Lindl. (Fire flash), a monocot; (ii)
 147 *Nematanthus glabra* Bailey. (Goldfish plant), a eudicot; (iii) *Nephrolepis cordifolia*
 148 (L.) C. Presl. var. 'duffii' (Lemon button fern), a monilophyte; and (iv) *Schefflera*
 149 *arboricola* Hayata. (Dwarf umbrella tree), also a eudicot. When not being tested, all
 150 plants within their green wall modules were maintained in a glasshouse lined with shade
 151 cloth, with an average temperature of $23.7 \pm 3.6^\circ\text{C}$, relative humidity of $68.1 \pm 16.0\%$,
 152 and a maximum mid-day light level of $90 \pm 10 \mu\text{mol.m}^{-2}.\text{s}^{-1}$ (4860 ± 54 lux). Plants
 153 were allowed to develop for 8 months under glasshouse conditions after planting and
 154 prior to testing. All modules were watered once weekly to saturation, as per industry
 155 standards.



156

157 **Fig. 2. Green wall plant species tested in the current experiment. From top left to**
158 **right; *Chlorophytum orchidastrum* (Fire flash), *Nematanthus glabra* (Goldfish**
159 **plant), *Nephrolepis cordifolia* (Lemon button fern), and *Schefflera arboricola***
160 **(Dwarf umbrella tree).**

161 At the conclusion of the VOC removal trials, plants were carefully removed from the
162 biofilter. The substrate was gently washed from the plants, and the plants were assessed
163 for several morphological characteristics that could have influenced VOC removal
164 efficiency, with four replicates per plant trait.

165 Average root diameter was recorded using callipers, by taking four composite
166 measurements from each plant, from four plant replicates per species. Root and leaf
167 mass fresh weights were recorded with scales. Root surface areas were determined by
168 creating plant pressings of the samples between two sheets of clear Perspex. Images of
169 each pressing were taken with a camera (Canon 1100D, 18 mm lens, Canon Australia
170 Pty Ltd, Macquarie Park, Australia) placed ~100 cm vertically above the Perspex sheets
171 containing the leaves and roots. Image analysis software (Fiji Image J 1.50g; National
172 Institutes of Health, Bethesda, Maryland, USA) was utilised to measure root surface
173 area by multiplying the two-dimensional root surface area by π . Leaf area was
174 determined using portable leaf area machine (Licor LI-3000-A, Lincoln, Nebraska,
175 USA).

176 Plant morphological data is shown in Table 1. For all variables, there were substantial
177 variations in morphology amongst the test species.

178 Table 1. Plant morphological data of each plant species. (Means \pm SEM, n =4).

Species	<i>Chlorophytum orchidastrum</i>	<i>Nematanthus glabra</i>	<i>Nephrolepis cordifolia</i>	<i>Schefflera arboricola</i>
Root diameter (mm)	3.55 \pm 0.32	0.52 \pm 0.06	0.68 \pm 0.07	3.95 \pm 0.79
Root mass fresh weight (g)	13.71 \pm 1.51	1.22 \pm 0.61	3.27 \pm 0.93	16.01 \pm 3.36
Leaf mass fresh weight (g)	26 \pm 3.34	30.22 \pm 16.64	214.42 \pm 46.2	245.6 \pm 53.81
Root surface area (cm ²)	150.8 \pm 20.01	63.71 \pm 28.45	6.92 \pm 0.35	33.6 \pm 4.31
Leaf surface area (cm ²)	731.17 \pm 229.5	255.6 \pm 121.9	15 \pm 0.65	255.63 \pm 1.75

179

180 *Experimental set up*

181

182 Experiments used a flow-through chamber system to assess the SPREs of benzene,
183 ethyl acetate and TVOCs. This set up used a sealed Perspex chamber (0.6 X 0.6 X 0.6
184 m; 216 L), of which one of the sides could be removed and resealed using adhesive
185 foam rubber and adjustable metal clamps, thus allowing green wall module placement
186 in the chamber. Ducting was fixed to one side of the chamber, which led to a second
187 chamber in which air pollutants were generated. The generated air pollutants flowed
188 through the fitted ducting with active airflow provided by a 16 W fan housed within the
189 ducting, before flowing through the biofilter. A second fan within the sealed Perspex
190 chamber encouraged mixing of the chamber atmosphere, creating a homogenous
191 concentration of pollutants within the chamber, before exhaust into another ducting
192 system on the opposite side of the chamber. This led to a third chamber that housed a
193 photo-ionisation detector (PID) that was used to monitor the concentration of air
194 pollutants. Air was exhausted to waste through a vacuum exhaust after sampling.

195 Single pass removal efficiency represents the percentage of a VOC removed from the
196 air stream as it passed through the biofilter, relative to the control treatment. In order to
197 determine the removal efficiency for each VOC, all trials were run independently, i.e.
198 with a single pollutant per run, with three replicates per treatment. Gaseous ethyl acetate
199 (EtOAc) and benzene were chosen to assess how the system comparatively treats
200 hydrophilic VOCs (EtOAc: solubility at 25°C = ~80.3 g/L) and hydrophobic VOCs
201 (benzene: solubility at 25°C = ~1.71 g/L). Each VOC was generated by placing 4.0 mL
202 of the liquid chemical into a 10 mL sealed glass vial and extracting 2.5 mL of the
203 vapour-saturated headspace with a gas chromatograph plunger-in-needle style syringe,
204 which was then injected into the pollutant generation chamber of the flow through
205 system. This process produced a pulse of VOC through the flow through duct. The
206 benzene treatment was thus a ~10 minute pulse, reaching a peak concentration of 4.170
207 \pm 0.144 ppm ~60 seconds after injection. The EtOAc treatment generated a ~8 minute
208 pulse, reaching a peak concentration mean of 3.997 \pm 0.074 SD ppm, 45 seconds after

209 injection. Additionally, the system was tested for the SPRE of ambient total VOCs
210 (TVOCs) using laboratory air supplied by the building's HVAC system, thus reflecting
211 the usual concentration of TVOCs in the room's normal operational state (~35 ppb).
212 Given that this experiment was performed in a general use research laboratory, the
213 TVOC concentration would be expected to be greater than that experienced in most
214 other building types. The concentration of the effluent gas was monitored with a PID
215 (ppbRAE 3000, RAE Systems, San Jose, CA, USA), with corrections applied as per
216 the manufacturer's instructions for the two VOCs.

217 Blank data (chamber with no green wall module present) for all VOC treatments was
218 also collected and used to calculate the background VOC removal efficiency of the flow
219 through system. The empty flow-through chamber was exposed to the stream of
220 gaseous VOC with identical concentration and flow conditions as the biofilter trials.
221 Calculations of specific VOC and TVOC removal efficiencies were thus based on
222 measurements at the same sampling point in the system with or without the biofiltration
223 system present.

224

225 *Data analysis*

226

227 After the data was checked for normality with Kolmogorov–Smirnov test and checked
228 for homogeneity of variance with Levene's test; one factor ANOVAs followed by
229 Tukey's *post hoc* tests (IBM SPSS Statistics Version 21, IBM Corp, Armonk, NY,
230 USA) were conducted to compare the SPREs of EtOAc, benzene and TVOCs amongst
231 the different plant species. The presence and strength of the relationship between
232 benzene removal and EtOAc removal across treatments was examined by computing
233 the Pearson correlation coefficient. Further, statistical associations between plant
234 morphological traits across plant species and pollutant removal were also tested with
235 Pearson correlations.

236

237 **Results**

238

239 EtOAc and benzene removal rates for each plant species are shown in Figs. 3 and 4.
240 Ambient indoor TVOC removal is shown in Fig. 5. VOC removal was achieved in all
241 treatments, across all plant species.

242 The system when tested for EtOAc removal recorded removal efficiencies in the range
243 of 32.36–91.19%. EtOAc removal efficiencies were $39.97 \pm 5.17\%$ for *N. glabra*, 80.69
244 $\pm 5.97\%$ for *S. arboricola*, $64.02 \pm 1.06\%$ for *N. cordifolia*, and $82.61 \pm 5.97\%$ for *C.*
245 *orchidastrum*. Significant differences in EtOAc removal were observed among plant
246 species tested (df 3,8 F=14.19, P=0.001), with *C. orchidastrum* and *S. arboricola*
247 recording significantly higher EtOAc SPREs than *N. glabra* and *N. cordifolia* (Figure
248 3, P<0.05 for all differences mentioned).

249 The system when tested for benzene removal recorded SPREs between 45.54–59.50%.
250 Benzene single pass removal efficiencies were $58.78 \pm 1.07\%$ for *N. glabra*, $51.01 \pm$
251 3.03% for *S. arboricola*, $48.00 \pm 2.19\%$ for *N. cordifolia*, and $47.65 \pm 1.46\%$ for *C.*
252 *orchidastrum*. Significant differences in benzene removal were observed among the
253 plant species tested (df 3,8 F=18.61, P=0.001), with *N. glabra* recording significantly
254 greater benzene SPRE than the other plant species (Figure 4, P<0.05 for all
255 comparisons).

256 When comparing ambient TVOC removal efficiencies, no significant differences
257 amongst plant species were observed (Figure 5, df 3,8 F=0.01, P=0.998).

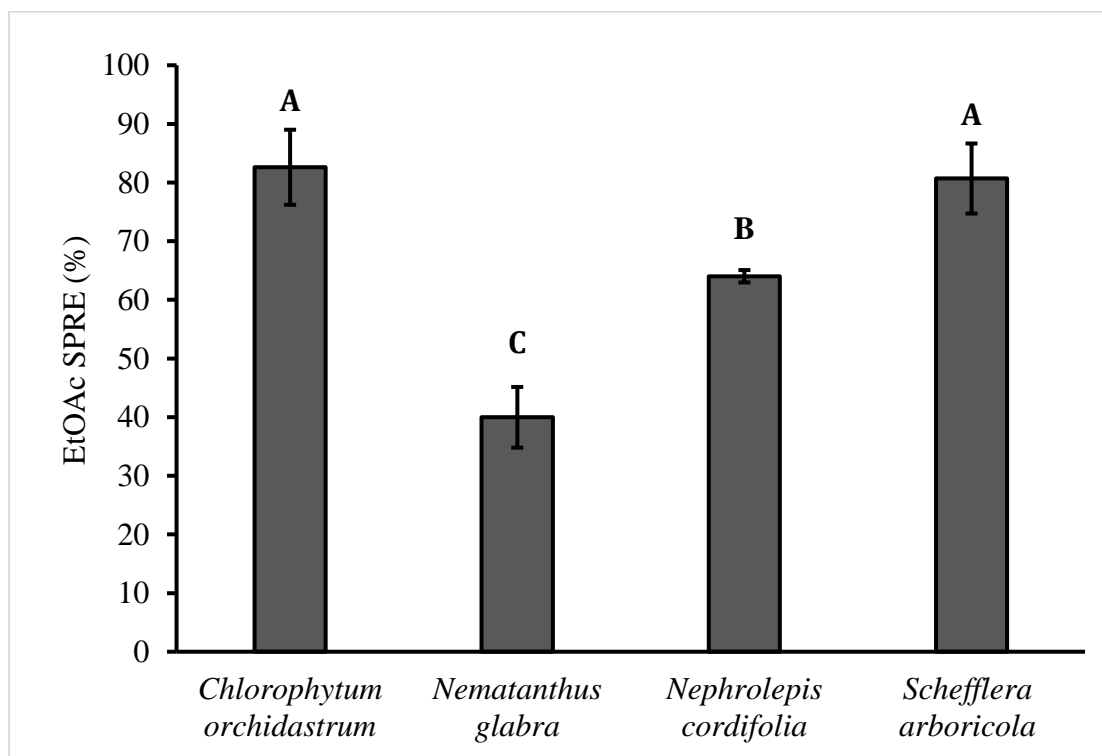
258 Plant species that were efficient for EtOAc removal demonstrated lower efficiency for
259 benzene removal, and *vice versa*. When removal efficiencies were combined across
260 species, benzene removal rates were significantly negatively correlated with EtOAc
261 removal ($r=-0.688$, $P=0.013$).

262 EtOAc SPRE was significantly positively correlated with root surface area ($r=0.694$,
263 $P=0.005$), root mass ($r=0.666$, $P=0.005$), root diameter ($r=0.479$, $P=0.05$), and leaf area
264 ($r=0.664$, $P=0.005$). No associations were found between EtOAc SPRE and leaf mass.

265 Benzene SPREs were not positively associated with any of the plant traits measured,
266 however they were significantly negatively correlated with root surface area ($r=-0.699$,
267 $P=0.003$), root mass ($r=-0.318$, $P=0.036$) and root diameter ($r=-0.479$, $P=0.05$). No
268 significant associations were found between benzene SPRE and any leaf traits.

269 It should be noted, whilst many correlations between VOC removal and plant traits
270 were statistically significant, the values of the correlation coefficients obtained were
271 moderately low, with none exceeding $r = 0.7$.

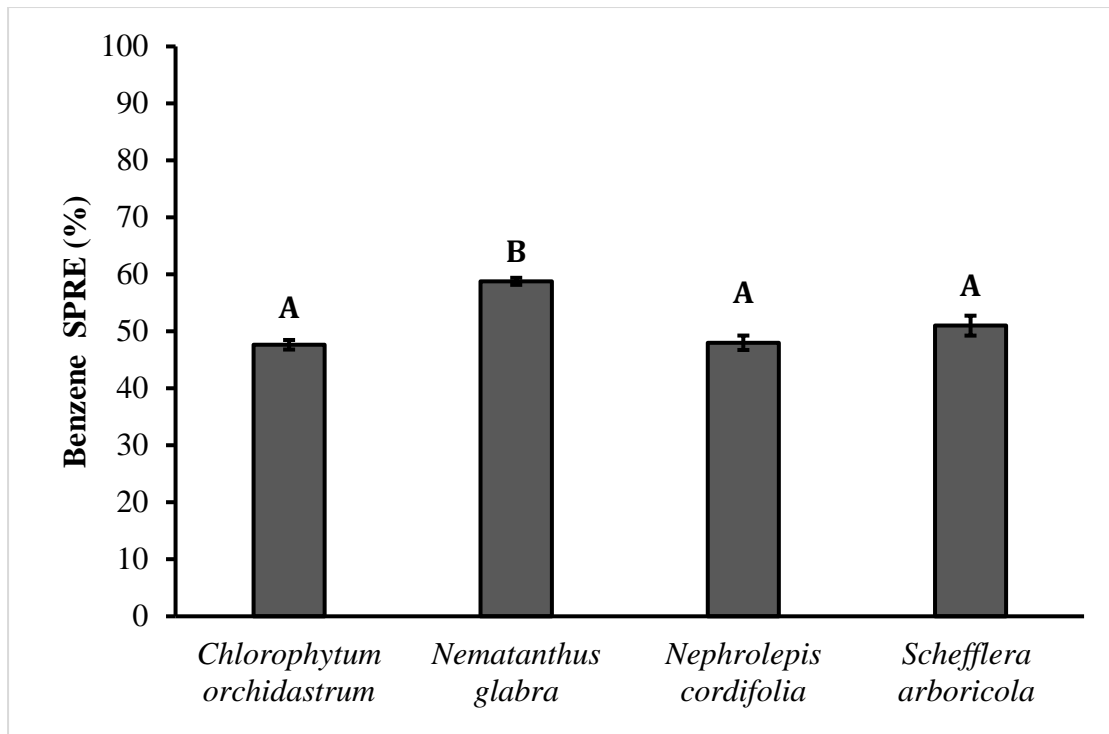
272 As no significant differences amongst plant species were observed in TVOC removal
273 efficiencies, no plant trait associations were tested with this treatment.



274

275 **Fig. 3.** Average levels of EtOAc removal across plant species (Means \pm SEM, $n=3$).
276 Species sharing the same letter are not significantly different using Tukey's post hoc
277 test, $P>0.05$.

278



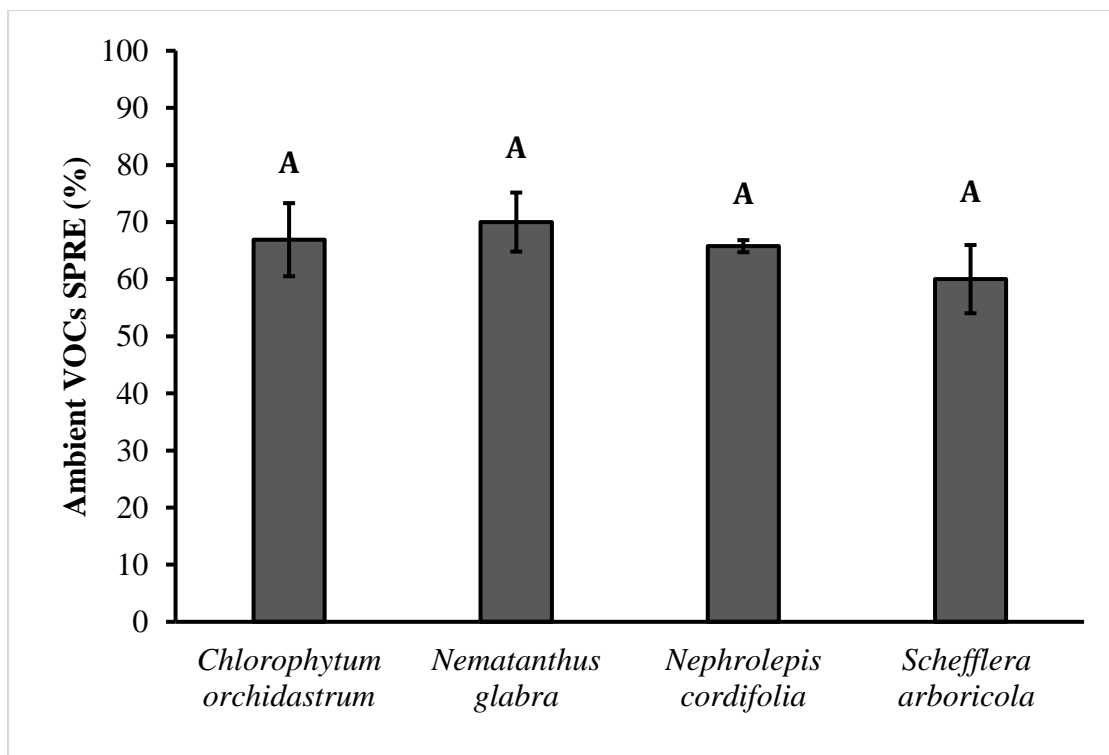
279

280 **Fig. 4.** Average levels of benzene removal across plant species (Means \pm SEM, n =3).

281 Species sharing the same letter are not significantly different using Tukey's post hoc

282 test, $P > 0.05$.

283



284

285 **Fig. 5.** Average levels of ambient indoor VOC removal across plant species (Means \pm

286 SEM, n =3). Species sharing the same letter are not significantly different using

287 Tukey's post hoc test, $P > 0.05$.

288 **Discussion**

289 If active green walls are to be used as functional air cleaning devices, the VOC removal
290 efficiency of these systems must be developed so as to provide maximized air cleaning
291 efficiency, whilst minimising additional energy use. To date, there is a scarcity of
292 literature on the relative contribution of the botanical component to the overall VOC
293 filtration ability of these systems. The primary observation of the current work is that
294 plant type does influence the system's capacity for VOC removal. It is well established
295 that potted plants can effectively improve indoor air quality by reducing hazardous
296 VOCs (Ugrekheldze et al. 1997, Kim and Jeon 2009, Sriprapat and Thiravetyan 2013,
297 Dela Cruz et al. 2014, Kim et al. 2016, Hörmann et al. 2018). However, it is known
298 that the efficiency of VOC removal varies substantially both among plant species (Kim
299 et al. 2018, Yoo et al. 2006), and with the molecular characteristics of individual
300 compounds. The current work extends this understanding to active botanical biofilters,
301 where less botanical influence might be expected due to the reduced VOC residence
302 time within the biological components of these systems.

303
304 Given that increasing root surface area, root diameter and root mass were all associated
305 with increasing EtOAc removal, it is proposed that the plant roots may provide
306 hydrophilic adsorbent sites for EtOAc, or facilitate high microbial activity in the
307 substrate stimulated by root exudates acting as nutrients for soil microorganisms (Kim
308 et al. 2018). Alternatively, it is possible that root exudates may alter the chemical
309 composition of the rhizosphere and thus influence the capacity of specific VOCs to
310 adsorb to the substrate (Pettit et al. 2018). Furthermore, with the substantial air flow
311 inherent in active biofiltration, certain root morphologies may enable increases in
312 exposure at the substrate/root/pollutant interface; potentially elevating the EtOAc
313 removal capacity. Additionally, it is plausible that the aerial plant parts could influence
314 EtOAc removal, with both the stomata and cuticle on the leaves creating pathways for
315 VOC removal, as has been proposed in previous work (Gkorezis et al. 2016, Jindachot
316 et al. 2018), and observed in the current study in the positive correlations between leaf
317 surface area and EtOAc SPRE. However, EtOAc is a relatively hydrophilic VOC, and
318 therefore will not diffuse readily through the cuticle due to its waxy nature, and thus
319 may largely be taken up through the stomata when they are open. This proposal is
320 supported by the current data, as *N. glabra* would not have been capable of stomatal
321 activity during the day as it is a CAM plant, and thus only opens its stomata in the dark,
322 a strategy that has evolved to limit moisture loss (Paull et al. 2018). In any case, the
323 correlations detected between EtOAc removal and plant characteristics in the current
324 study provide evidence that the belowground components of plants are the major
325 regulator of VOC removal in active air phytoremediation systems.

326
327 The benzene removal data was reasonably consistent across plant species, with less
328 than 15% variability between the most and least effective plants. In contrast to the
329 EtOAc removal trials, the most effective plant species for benzene removal was *N.*
330 *glabra*, likely due to the high wax content in its leaf cuticles, although stomatal benzene
331 uptake has also been proposed in previous work (Setsungnern et al. 2017).
332 Alternatively, *S. arboricola* has been shown to have high benzene removal efficiency,
333 which has been previously attributed to its relatively large leaf area (Parseh et al. 2018),
334 along with a significant waxy cuticle comprised of alpha-linoleic acid and dodecyl
335 cyclohexane (Treesubstorn and Thiravetyan 2012). Interestingly, in this study, no
336 significant associations were found between benzene SPRE and any leaf traits, and thus
337 we cannot determine the pathway for benzene removal observed for *N. glabra* in the

338 current work, nor can we eliminate effects that this species may have had on the
339 substrate as the means by which enhanced benzene removal was afforded. It is
340 recommended that in future work, quantitative assessments of leaf hydrophobic
341 compounds, such as waxes, be made to determine whether they have a major effect on
342 hydrophobic VOC removal. Work using substrates in which various plant species have
343 been grown, but subsequently removed, would also be of value to elucidate plant-
344 mediated substrate effects on VOC removal.

345 As this experiment assessed SPRE, the removal of each chemical was dependent upon
346 its residence time within the active green wall system (i.e. the time that the polluted air
347 stream was in contact with the growth substrate and plant foliage). Due to the limited
348 residence times in these single pass experiments, it is likely that the removal processes
349 in these trials were predominantly sorption process as opposed to microbial degradation
350 processes. Whilst several static chamber studies have found that the potted-plants'
351 microbial community plays a significant role in VOC removal (Aydogan and Montoya
352 2011, Orwell et al. 2006), the very short residence time of pollutants in the current trials
353 (<10 min in all cases) would probably limit the time available for microbial metabolism
354 to occur. Mikkonen et al. (2018) observed a decrease in a green wall system's microbial
355 diversity after it had been exposed to VOCs for 16 weeks, as heterotrophic bacterial
356 groups that could use the VOCs as a nutrient source had been favorably selected and
357 became numerically dominant in the community. Whilst it is thus possible that
358 prolonged exposure to VOCs would increase the bacterial community's VOC
359 degradation capacity, this effect may not affect an active green wall's *in situ* VOC air
360 cleaning efficiency to the same degree, due to the short pollutant residence time
361 (Weyens et al. 2015).

362

363 It is clear from the current findings that plant selection will effect VOC removal in
364 active botanical biofilter systems. This may be of value in functional biofilter design,
365 especially if hydrophilic VOCs are problematic in a specific application. The general
366 *in situ* importance of these effects may, however, not be of great magnitude, as all plants
367 tested has considerable VOC removal efficiencies; thus a biofilter of adequate size
368 relative to the concentration of VOCs encountered should lead to major VOC
369 reductions, irrespective of the performance of the individual species selected.

370

371 A further consideration in plant selection relates to long-term VOC effects on plant
372 health. Whilst several common green wall plants have been shown to have excellent
373 short term / high concentration pollutant tolerance (Paull et al. 2018), there have yet to
374 be long term trials pollutant exposure trials. The use of active airflow through the
375 plants' substrate may have unexpected effects on plant health, and these conditions
376 must be tested as a key contributor to the whole-of-life costs of botanical pollutant
377 removal systems. Similarly, substrate changes over long-term exposure remain
378 unknown, beyond those effects related to microbial community shift, previously
379 described.

380

381 A potential solution for the observed variability in VOC removal amongst plant species
382 is through green wall design, with targeted combinations of different species growing
383 together. Uniform plant types in large scale green walls are rarely encountered, thus the
384 combined removal efficiencies of biodiverse green walls could be used to account for
385 a diversity of VOCs. However, with the introduction of active air flow through green
386 wall systems, very little is known about the potential influence this will have on VOC
387 degrading bacteria in the substrate, or whether they play an important role in VOC

388 degradation, as has been shown to be the case in static systems (Wood et al. 2002). It
389 is thus proposed that experiments using radiolabelled VOCs will be required to test the
390 role of rhizospheric bacteria to utilize and degrade airborne VOCs in constant air flux
391 conditions.

392 **Summary and conclusion**

393 There is a growing body of evidence indicating that botanical biofiltration has major
394 potential for the low energy use removal of a broad range of air pollutants. However
395 the physical, chemical and biological functions of these systems remain poorly
396 described, and thus evidence supporting the design criteria for tailoring or maximising
397 pollutant filtration efficiency is still weak. The current work assessed the capacity of
398 several common green wall plants for removing two major classes of VOC providing a
399 baseline indication of the plant species' removal efficiencies for model hydrophobic
400 and hydrophilic VOCs. The findings suggest that target pollutant dependent botanical
401 biofilter plant selection are possible, as whilst all plant species were successful in
402 removing ambient TVOCs and benzene, there were substantial differences between
403 species in hydrophilic VOC removal. The authors propose that future work should
404 examine plant effects on biofilter substrates to determine the specific physical, chemical
405 or biological processes that are associated with VOC removal.

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