Retracing the evolutionary history of the

Trypanosomatidae: the use of kinetoplast DNA in molecular systematics, species identification and diagnostics

By

Alexa Kaufer

2020

Certificate of Original Authorship

I, **Alexa Kaufer** declare that this thesis, is submitted in fulfilment of the requirements for the award of Doctor of Philosophy in the School of Life Sciences/Science at the University of Technology Sydney.

This thesis is wholly my own work unless otherwise reference or acknowledged. In addition, I certify that all information sources and literature used are indicated in the thesis.

This document has not been submitted for qualifications at any other academic institution.

This research is supported by the Australian Government Research Training Program.

I hereby certify that the above statements are true and correct:

Production Note:

Signature removed prior to publication.

Alexa Kaufer (PhD Candidate)

Date: January 2020

Acknowledgements

Firstly, I would like to acknowledge that this project would not have been possible without the incredible dedication and help of my supervisors Prof. John Ellis, Dr. Damien Stark and Dr. Joel Barratt. I would like to thank them for their amazing guidance, knowledge and support throughout these past years. Their mentoring has been an invaluable gift, for which I am extremely grateful.

I would also like to thank everyone else who has been involved throughout my candidature, particularly Larissa Calarco and Rory Gough. Their continual help and guidance were an immeasurable help, offering fellow moral support throughout one the hardest challenges I have ever undertaken.

For everyone I've thanked, my words of appreciation could never suffice; I could not have had a greater and more fulfilling candidature without each and every one of you.

Format of Thesis

I, Alexa Kaufer declare the format of this manuscript is 'Thesis by compilation'.

Publications Arising from Thesis

KAUFER, A., STARK, D. & ELLIS, J. 2019. Evolutionary Insight into the Trypanosomatidae Using Alignment-Free Phylogenomics of the Kinetoplast. *Pathogens*, 8, 157.

KAUFER, A., ELLIS, J. & STARK, D. 2019. Identification of Clinical Infections of *Leishmania* imported into Australia: Revising Speciation with Polymerase Chain Reaction-RFLP of the Kinetoplast Maxicircle. *The American Journal of Tropical Medicine and Hygiene*, 101, 599.

KAUFER, A., BARRATT, J., STARK, D. & ELLIS, J. 2019. The complete coding region of the maxicircle as a superior phylogenetic marker for exploring evolutionary relationships between members of the Leishmaniinae. *Infection, Genetics and Evolution*, 70, 90-100.

KAUFER, A., ELLIS, J., STARK, D. & BARRATT, J. 2017. The evolution of trypanosomatid taxonomy. *Parasites & Vectors*, 10, 287.

Table of Contents

Certificate of Original Authorship	i
Acknowledgements	ii
Format of Thesis	iii
Publications Arising from Thesis	iv
Table of Contents	v
List of Figures	vi
List of Tables	vii
Supplementary Information	ix
Thesis Abstract	x
Exegesis	xii
Chapter 1	1
Chapter 2	19
Chapter 3	31
Chapter 4	49
Chanton 5	62

List of Figures

Chapter 1
Figure 1. Vectors and invertebrate hosts of some trypanosomatids
Figure 2. The six major morphotype classes of trypanosomatids
Figure 3. Some clinical manifestations of leishmaniasis
Figure 4. Photomicrographs of stained smears showing Leishmania infections
Figure 5. The growth cycle of trypanosomatids within invertebrates
Figure 6. Diagrammatic representation of the three <i>Leishmania</i> sections proposed by Lainson & Shaw (1979)
Figure 7. Light and electron micrographs of Zelonia australiensis
Chapter 2
kinetoplast2
Figure 2. Graphical map of <i>Z. australiensis</i> maxicircle genome assembled from Illumina sequencing of total DNA with the regions targeted by long-range PCR highlighted
Figure 3. DNA electrophoresis of PCR products generated through optimised LR-PCR assays
Figure 4. Schematic diagram of the maxicircle genome sequence of various trypanosomatid spp. generated in this study
Figure 5. Inferred evolutionary relationship showing genetic distance between <i>Z. australiensis</i> and other trypanosomatids using the maxicircle coding region
Figure 6. Inferred evolutionary relationship between <i>Z. australiensis</i> and other trypanosomatids using the maxicircle coding region
Figure 7. Phylogenetic time tree inferring the evolutionary relationships between the Leishmaniinae and other trypanosomatids using the maxicircle coding region 27
Chapter 3
Figure 1. Graphical map of <i>Leishmania guyanensis</i> and <i>Phytomonas francai</i> maxicircle genomes assembled from Illumina and PacBio sequencing
Figure 2. Circular confirmation of GC plot and GC skew of the maxicircle kinetoplast for <i>Leishmania braziliensis, Trypanosoma brucei rhodesiense and Trypanosoma cruz</i>
Figure 3. Self dottup plot comparative analysis of the entire maxicircle genome (right panel) and divergent region (left panel) of <i>Leishmania braziliensis</i> (A), <i>Trypanosoma brucei rhodesiense</i> (B) and <i>Trypanosoma cruzi</i> (C) against their own sequences 33

Figure 4. Analysis of total repeated sequences in the maxicircle divergent region of various trypanosomatid species
Figure 5. Inferred evolutionary relationships between species of the trypanosomatid family using aligned and alignment-free analysis of the maxicircle coding region 40
Figure 6. Phylogenetic time tree demonstrating the complex evolutionary relationships of the trypanosomatid family using the maxicircle kDNA41
Chapter 4
Figure 1. World map of geographical distribution of clinically important, human-infecting <i>Leishmania</i> species
Figure 2. DNA electrophoresis of sensitivity test of ND7-PCR of L. (L.) major 56
Figure 3. PCR-RFLP analysis of <i>Leishmania</i> species with the restriction enzyme <i>Nla</i> III
Figure 4. PCR-RFLP analysis of <i>Leishmania</i> species with the restriction enzyme <i>Hpy</i> CH4IV
Figure 5. Inferred evolutionary relationships showing genetic distance between clinical isolate of imported cases of leishmaniasis
Figure 6. Levels of intra- and inter-species mean similarities of <i>Leishmania</i> spp 58
Chapter 5
Figure 1. The tree of life depicting the relationships of the eukaryotes
Figure 2. Classification of the trypanosomatid family, subfamily, genus, subgenus and species based on Espinosa et al. 2016 and Kaufer et al. 201974
Figure 3. The evolution of phylogenetic analyses inferred from various molecular targets over the years
Figure 4. Diagram of gene synteny of trypanosomatid maxicircle genomes based on Kaufer et al. 2019
Figure 5. Alignment-free phylogenetic workflow using the frequency feature profile (FFP) analysis

List of Tables

Chapter 1	1
Table 1. Currently recognised genera of the family Trypanosomatidae	3
Table 2. Historical overview of the studies describing unusual infections caused b monoxenous trypanosomatids	•
Chapter 2	19
Table 1. Main genetic information contained in the maxicircle	22
Table 2. Data generated from LR-PCR following QC grooming in this study	25
Chapter 3	31
Table 1. Comparison of the short-read Illumina and hybrid Illumina/PacBio assert of various trypanosomatid species showing output length and sequence identity (%)	-
Chapter 4	49
Table 1. List of bacterial and fungal species tested as a negative control group	53
Table 2. Reaction conditions for each restriction enzyme used for the ND7 PCR-RFLP	
Table 3. Number of SNPs (above diagonal) and indels (below diagonal) identified between the various <i>Leishmania</i> species	
Table 4. Patient details, associated risk factor, clinical presentation and species identified	55
Chapter 5	62
Table 1. Important diseases caused by trypanosomatids	70
Table 2. Genomic comparison of various trypanosomatid species	83

Supplementary Information

The thesis chapters (2-4) make reference to supplementary, supporting or additional information. This information is contained in files that can be found at the URL specified in the chapter and on the USB provided.

Thesis Abstract

The Trypanosomatidae (class Kinetoplastida) are a diverse and widespread group of protists characterised by their possession of a unique and highly specialised mitochondrial homologue known as the kinetoplast. All trypanosomatid parasites are exclusively parasitic, with the majority of genera being restricted to a single invertebrate host (i.e. monoxenous lifecycle). However, they are primarily known for their pathogenic dixenous members (i.e. having a two-host life cycle), that serve as the aetiologic agents of several important neglected tropical diseases (NTDs) including leishmaniasis, Chagas disease and human African Trypanosomiasis. Recent advancements in molecular biology have improved our knowledge of evolutionary relationships between trypanosomatid species by revolutionising the genetic approach to trypanosomatid systematics. The kinetoplast DNA, and more specifically, the maxicircle genome represents a valuable taxonomic marker given its unique presence across all Kinetoplastids. The research described in this thesis was performed to explore the suitability of the kinetoplast DNA as a much-needed standardised framework for the taxonomic classification and species identification of protozoans falling within the Trypanosomatidae family.

The main research outcome provides the most in-depth analysis of the trypanosomatid family to date, demonstrating extensive evidence for the superiority of the maxicircle for the taxonomic resolution of the Trypanosomatidae. Chapter 1 presents a comprehensive review outlining the important developments that have been made in the field of trypanosomatid taxonomy, advancing our current knowledge over the relationships between members of the trypanosomatid family. Chapter 2 demonstrates the superiority of the maxicircle genome for the phylogenetic inference of

the Leishmaniinae. Phylogenetic analyses provided support imperative towards the Supercontinents hypothesis of dixenous parasitism within the Trypanosomatidae. The comprehensive analysis of the entire family of trypanosomatid parasites in Chapter 3 revealed strong support for the multiple origin and independent evolution of dixenous parasitism within the trypanosomatid family. Kinetoplast DNA is an organelle exclusive to the Kinetoplastids, unique in its structure, function and mode of replication and thus represents a unique diagnostic target, with the potential to differentiate different species and strains of *Leishmania*. The implementation of novel diagnostic procedures for leishmaniasis such as the one reported in Chapter 4 is intended to establish a gold standard practice for the diagnosis and treatment of leishmaniasis. Ultimately, Chapter 5 reviews the completion of this thesis, demonstrating that use of the maxicircle genomes provide an excellent benchmark for future studies involving the phylogenetic analyses, taxonomic classification and species identification of the Trypanosomatidae.