

Complete Genome Sequence of *Stenotrophomonas maltophilia* Strain CF13, Recovered from Sputum from an Australian Cystic Fibrosis Patient

Mohammad Hamidian,^a James Lazenby,^a Joyce To,^a Rebecca Hartstein,^a Jana Soares,^a Samantha McNamara,^a Cynthia B. Whitchurch^a

Microbiology[®]

Resource Announcements

^aThe ithree institute, University of Technology Sydney, Ultimo, NSW, Australia

AMERICAN SOCIETY FOR

MICROBIOLOGY

ABSTRACT Stenotrophomonas maltophilia isolate CF13 is a multidrug-resistant isolate that was recovered in Sydney, Australia, in 2011, from a sputum sample from an individual with cystic fibrosis. The genome sequence of CF13 was completed using long- and short-read technologies.

Stenotrophomonas maltophilia is as an important opportunistic pathogen associated with antibiotic-resistant respiratory infections globally (1–3). A collection of putative *Pseudomonas aeruginosa* isolates were isolated from sputum samples from individuals with cystic fibrosis at NSW Health Pathology (Prince of Wales Hospital, Randwick, NSW, Australia) in 2011 as described previously (4). One multidrug-resistant isolate (CF13) was selected for sequence analysis and was found in this study to be *S. maltophilia*. Antibiotic sensitivity testing was performed by standard broth microdilution (5, 6). The MICs for CF13 were as follows: 4 mg/liter (gentamicin), 32 mg/liter (amikacin), 128 mg/liter (meropenem), 2 mg/liter (polymyxin B), 2 mg/liter (colistin), >128 mg/liter (aztreonam), 256 mg/liter (ceftazidime), 64 mg/liter (ticarcillin-clavulanic acid), and 128 mg/liter (fosfomycin).

Whole-cell genomic DNA of CF13 was isolated using the DNeasy microbial kit (Qiagen), from cells grown overnight at 37°C in LB inoculated from a frozen stock derived from a single colony. Illumina HiSeg and MinION sequencing was performed at the ithree Core Sequencing Facility, University of Technology Sydney (Ultimo, Australia). Library construction for Illumina MiSeq sequencing was performed following the adapted Nextera Flex library preparation kit process Hackflex, as described previously (7). The library for MinION sequencing was prepared using the rapid barcoding seguencing kit (SQK-RBK004) and was sequenced using a FLO-MIN106D flow cell (identification number FAL30280) according to the manufacturer's instructions (Oxford Nanopore Technologies, Inc., Oxford, UK). Illumina HiSeq sequencing generated 1,013,693 paired-end short reads with 50-fold coverage and an average length of 250 bp, and MinION sequencing produced a total of 23,907 reads with an N_{50} value of 18.2 kbp and 30-fold coverage. FastQC (v.0.11.9) (https://www.bioinformatics.babraham .ac.uk/projects/fastqc) and Filtlong (v.0.2.0) (https://github.com/rrwick/Filtlong) were used to check the quality of Illumina and MinION reads, respectively. Filtlong filtered long reads by quality, length, and comparison to a reference genome (K279a, GenBank accession number AM743169), excluding reads that did not belong to S. maltophilia. Illumina and MinION reads were trimmed using the Trimmomatic (v.0.39) (8) and Porechop (v.0.2.3) (9) programs, respectively, using default parameters. The highquality Illumina and MinION reads were assembled de novo using a hybrid assembly approach with the Unicycler program (v.0.4.7) (10), using default parameters, which resulted in a single chromosomal contig. Unicycler was used to rotate and open the chromosome at *dnaA* to determine that the genome was circular and complete, a

Citation Hamidian M, Lazenby J, To J, Hartstein R, Soares J, McNamara S, Whitchurch CB. 2020. Complete genome sequence of *Stenotrophomonas maltophilia* strain CF13, recovered from sputum from an Australian cystic fibrosis patient. Microbiol Resour Announc 9:e00628-20. https://doi.org/10.1128/ MRA.00628-20.

Editor Catherine Putonti, Loyola University Chicago

Copyright © 2020 Hamidian et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Mohammad Hamidian, mohammad.hamidian@uts.edu.au.

Received 29 May 2020 Accepted 11 July 2020 Published 6 August 2020

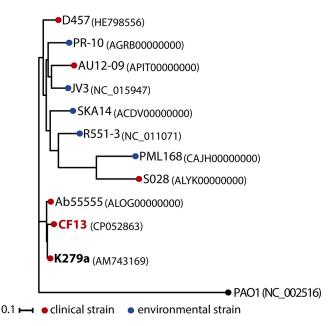


FIG 1 Core genome phylogenetic tree of *S. maltophilia* strains. Red and blue nodes indicate isolates recovered from clinical and environmental sources, respectively. The black node shows *Pseudomonas aeruginosa* PAO1, which was used as an outgroup. Numbers next to the strain names indicate GenBank accession numbers. Sequence reads were mapped to the reference strain K279a (17) using Snippy (v.4.6.0) (https://github.com/tseemann/snippy) to generate a whole-genome alignment. Variant sites were called using SAMtools (v.1.3.1.24) (9), and a maximum likelihood tree was inferred using RAxML (v.8) (21) with the generalized time-reversible gamma model of nucleotide substitution. Ten independent runs of RAxML with 1,000 bootstraps each gave nearly identical results; therefore, the first replicate was taken as a single representative result, with support values calculated from 1,000 bootstraps.

method and program that were validated previously (10). The chromosome, consisting of 4,591,696 bases with a GC content of 66.46%, was annotated using the NCBI Prokaryotic Genome Annotation Pipeline (v.4.11) (11). The chromosome of CF13 encodes a total of 4,086 putative proteins, with 69 tRNAs and 6 rRNA regions. CF13 carries no plasmid.

The complete genome sequence shows that CF13 belongs to a novel sequence type (ST) of *S. maltophilia*, namely, ST371 (with *atpD-5*, *gapA-62*, *guaA-289*, *mutM-158*, *nuoD-25*, *ppsA-116*, and *recA-103*). To determine the relationship of CF13 to other known strains (12–17), a phylogenetic tree was constructed. CF13 was found to be clustered with the reference strain K279a (Fig. 1). The population structure of *S. maltophilia* consists of 23 monophyletic lineages, with the largest lineage, Sm6, known as *S. maltophilia sensu stricto*, containing most strains characterized to date, including K279a (18). Hence, CF13 also belongs to Sm6.

CF13 contains the aminoglycoside resistance genes aph(3')-II and aac(6')-I and the bla_{L1} resistance gene, which encodes a broad-spectrum Ambler class B metalloenzyme. CF13 also carries the oqxB gene, which, in conjunction with oqxA, encodes a resistance-nodulation-division (RND)-type efflux pump that is involved in resistance to multiple antibiotics (19, 20). However, only a partial segment of oqxA was found in CF13.

Data availability. The complete genome sequence has been deposited in DDBJ/ ENA/GenBank under the accession number CP052863. Illumina and MinION sequence reads have been deposited in the Sequence Read Archive (SRA) database under the accession numbers SRR9903515 and SRR11687566, respectively.

ACKNOWLEDGMENTS

M.H. is supported by a UTS Chancellor's Research Fellowship (grant CPDRF PRO17-4005).

Kerry Varettas from NSW Health Pathology (Prince Of Wales Hospital, Randwick, NSW, Australia) provided the strains used in this study.

REFERENCES

- 1. Brooke JS. 2012. *Stenotrophomonas maltophilia*: an emerging global opportunistic pathogen. Clin Microbiol Rev 25:2–41. https://doi.org/10 .1128/CMR.00019-11.
- Chang YT, Lin CY, Chen YH, Hsueh PR. 2015. Update on infections caused by *Stenotrophomonas maltophilia* with particular attention to resistance mechanisms and therapeutic options. Front Microbiol 6:893. https://doi .org/10.3389/fmicb.2015.00893.
- Sanchez MB. 2015. Antibiotic resistance in the opportunistic pathogen Stenotrophomonas maltophilia. Front Microbiol 6:658. https://doi.org/10 .3389/fmicb.2015.00658.
- O'Carroll MR, Syrmis MW, Wainwright CE, Greer RM, Mitchell P, Coulter C, Sloots TP, Nissen MD, Bell SC. 2004. Clonal strains of *Pseudomonas aeruginosa* in paediatric and adult cystic fibrosis units. Eur Respir J 24:101–106. https://doi.org/10.1183/09031936.04.00122903.
- Andrews JM. 2001. Determination of minimum inhibitory concentrations. J Antimicrob Chemother 48(Suppl 1):5–16. https://doi.org/10 .1093/jac/48.suppl_1.5.
- Wiegand I, Hilpert K, Hancock RE. 2008. Agar and broth dilution methods to determine the minimal inhibitory concentration (MIC) of antimicrobial substances. Nat Protoc 3:163–175. https://doi.org/10.1038/nprot.2007 .521.
- Gaio D, To J, Liu M, Monahan L, Anantanawat K, Darling AE. 2019. Hackflex: low cost Illumina sequencing library construction for high sample counts. bioRxiv https://doi.org/10.1101/779215.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. Microb Gen 3:e000132. https://doi.org/10.1099/mgen.0.000132.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- Zhang L, Morrison M, O Cuiv P, Evans P, Rickard CM. 2013. Genome sequence of *Stenotrophomonas maltophilia* strain AU12-09, isolated from an intravascular catheter. Genome Announc 1:e00195-13. https:// doi.org/10.1128/genomeA.00195-13.
- 13. Zhu B, Liu H, Tian W-X, Fan X-Y, Li B, Zhou X-P, Jin G-L, Xie G-L. 2012. Genome sequence of *Stenotrophomonas maltophilia* RR-10, isolated as

A Microbiology

an endophyte from rice root. J Bacteriol 194:1280–1281. https://doi.org/ 10.1128/JB.06702-11.

- Allen MJ, Tait K, Mühling M, Weynberg K, Bradley C, Trivedi U, Gharbi K, Nissimov J, Mavromatis K, Jensen CN, Grogan G, Ali ST. 2012. Genome sequence of *Stenotrophomonas maltophilia* PML168, which displays Baeyer-Villiger monooxygenase activity. J Bacteriol 194:4753–4754. https://doi.org/10.1128/JB.00949-12.
- Song S, Yuan X, Liu S, Zhang N, Wang Y, Ke Y, Xu J, Huang L, Chen Z, Li Y. 2012. Genome sequence of *Stenotrophomonas maltophilia* S028, an isolate harboring the AmpR-L2 resistance module. J Bacteriol 194:6696. https://doi.org/10.1128/JB.01809-12.
- Lira F, Hernández A, Belda E, Sánchez MB, Moya A, Silva FJ, Martínez JL. 2012. Whole-genome sequence of *Stenotrophomonas maltophilia* D457, a clinical isolate and a model strain. J Bacteriol 194:3563–3564. https:// doi.org/10.1128/JB.00602-12.
- Crossman LC, Gould VC, Dow JM, Vernikos GS, Okazaki A, Sebaihia M, Saunders D, Arrowsmith C, Carver T, Peters N, Adlem E, Kerhornou A, Lord A, Murphy L, Seeger K, Squares R, Rutter S, Quail MA, Rajandream MA, Harris D, Churcher C, Bentley SD, Parkhill J, Thomson NR, Avison MB. 2008. The complete genome, comparative and functional analysis of *Stenotrophomonas maltophilia* reveals an organism heavily shielded by drug resistance determinants. Genome Biol 9:R74. https://doi.org/10 .1186/gb-2008-9-4-r74.
- Gröschel MI, Meehan CJ, Barilar I, Diricks M, Gonzaga A, Steglich M, Conchillo-Solé O, Scherer I-C, Mamat U, Luz CF, De Bruyne K, Utpatel C, Yero D, Gibert I, Daura X, Kampmeier S, Rahman NA, Kresken M, van der Werf TS, Alio I, Streit WR, Zhou K, Schwartz T, Rossen JWA, Farhat MR, Schaible UE, Nübel U, Rupp J, Steinmann J, Niemann S, Kohl TA. 2020. The phylogenetic landscape and nosocomial spread of the multidrugresistant opportunist *Stenotrophomonas maltophilia*. Nat Commun 11: 2044. https://doi.org/10.1038/s41467-020-15123-0.
- Rodríguez-Martínez JM, Díaz de Alba P, Briales A, Machuca J, Lossa M, Fernández-Cuenca F, Rodríguez Baño J, Martínez-Martínez L, Pascual Á. 2013. Contribution of OqxAB efflux pumps to quinolone resistance in extended-spectrum-β-lactamase-producing *Klebsiella pneumoniae*. J Antimicrob Chemother 68:68–73. https://doi.org/10.1093/jac/dks377.
- Hansen LH, Johannesen E, Burmølle M, Sørensen AH, Sørensen SJ. 2004. Plasmid-encoded multidrug efflux pump conferring resistance to olaquindox in *Escherichia coli*. Antimicrob Agents Chemother 48:3332–3337. https://doi.org/10.1128/AAC.48.9.3332-3337.2004.
- Kozlov AM, Darriba D, Flouri T, Morel B, Stamatakis A. 2019. RAxML-NG: a fast, scalable and user-friendly tool for maximum likelihood phylogenetic inference. Bioinformatics 35:4453–4455. https://doi.org/10.1093/ bioinformatics/btz305.