



**Genomic Characterisation of Extra-intestinal  
Pathogenic *Escherichia coli* from Human  
Infection**

A thesis submitted for the degree of

Doctor of Philosophy

by

Jessica McKinnon

Bachelor of Biotechnology (Honours)

## **Certificate of Authorship**

I declare that this thesis by compilation is submitted in fulfilment of the requirements for the award of Doctor of Philosophy, in the School of Life Sciences at the University of Technology Sydney.

This thesis is wholly my own work unless otherwise reference or acknowledged. In addition, I certify that all information sources and literature used are indicated in the thesis. This document has not been submitted for qualifications at any other academic institution.

This research is supported by an Australian Government Research Training Program.

Production Note:

Signature removed prior to publication.

Jessica McKinnon

May 2020

## Peer Reviewed Publications

### **Published with First Authorship (\* authors contributed equally)**

Darling AE\*, **McKinnon J\***, Worden P, Santos J, Charles IG, Roy Chowdhury P and Djordjevic SP. (2014). A draft genome of *Escherichia coli* sequence type 127 strain 2009-46. *Gut Pathogens* 6(32).

**McKinnon J\***, Roy Chowdhury P\* and Djordjevic SP. (2018). Genomic analysis of multidrug-resistant *Escherichia coli* ST58 causing urosepsis. *International Journal of Antimicrobial Agents* 52(3):430-435.

Roy Chowdhury P\*, **McKinnon J\***, Liu M and Djordjevic SP. (2018). Multidrug resistant uropathogenic *Escherichia coli* ST405 with a novel, composite IS26 transposon in a unique chromosomal location. *Frontiers* 8(9):3212.

**McKinnon J**, Roy Chowdhury P and Djordjevic SP. (2020) Molecular analysis of an IncF ColV-like plasmid lineage that carries a complex resistance locus with a trackable genetic signature. *Microb Drug Resist.*

Bogema DR\*, **McKinnon J\***, Liu M, Hitchick N, Miller N, Venturini C, Iredell J, Darling AE, Roy Chowdhury P, Djordjevic SP. (2020). Whole-genome analysis of extraintestinal *Escherichia coli* sequence type 73 from a single hospital over a 2 year period identified different circulating clonal groups. *Microb Genom* 6(1).

### **In Preparation**

**McKinnon J**, Roy Chowdhury P and Djordjevic SP. (2019). Whole genome sequencing of 83 class 1 integron positive clinical isolates from urinary tract infection.

### **Co-Authorships**

Melvold JA, Wyrsh ER, **McKinnon J**, Roy Chowdhury P, Charles IG and Djordjevic, SP. (2017). Identification of a novel qnrA allele, qnrA8, in environmental *Shewanella* algae. *Journal of Antimicrobial Chemotherapy* 72, 2949-2952.

Roy Chowdhury P, **McKinnon J**, Wyrsh E, Hammond JM, Charles IG and Djordjevic SP. (2014). Genomic interplay in bacterial communities: implications for growth promoting practices in animal husbandry. *Frontiers in Microbiology* 5.

## **Conference Presentations**

- Molecular Dissection of Drug Resistance Regions in Clinical *Escherichia coli* Isolates from a Sydney Hospital

**New Horizons: Research and Education for Optimal Health**

**Wednesday 20<sup>th</sup> November, 2013, Poster 3**

- Comparative Genomics of Sequential *Escherichia coli* Strains 2009-49 and 2009-52: A Micro-Evolutionary Event in Real-Time?

**BacPath 13: Molecular Analysis of Bacterial Pathogens**

**Tuesday 29<sup>th</sup> September, 2015, Poster Session 3**

- Identification of a Unique Antibiotic Resistance and Virulence Locus from a Human Blood-Stream Infection

**International Conference on Plasmid Biology**

**Poster presentation, 2016**

- A uropathogenic *Escherichia coli* ST405 with a unique complex resistance region

**Mobile Genetic Element: Transposable Elements, Integrative and Plasmids**

**Oral presentation number 8, 2017**

## **Abstract**

Tracking resistance genes based on specific structural features of class 1 integrons is an integral part of clinical epidemiology. A class 1 integron is a gene capture and expression unit, most frequently found in Gram-negative bacteria. They are known to be one of the greatest contributors to the spread of multi-drug resistance genes within clinical isolates.

Recent studies indicate that certain insertion elements target specific regions of class 1 integrons creating unique structures. Consequently, the resistance pool in such integrons goes unnoticed in standard molecular screening methodologies, although they are equally efficient in expressing and disseminating resistance genes. Examples of such class 1 integrons with atypical structures, now realised to have widely disseminated within human and animal *E. coli* populations worldwide, were generated by the insertion of a genetic element known as IS26. This project aimed to determine the presence of such structures, as well as characterise the diverse complex resistance loci (CRL) and virulence cargo of extraintestinal pathogenic *E. coli* samples in a specific Sydney hospital. This was achieved by identifying the virulence-associated and antimicrobial resistance gene pool harboured by these strains with a particular focus on insertion or deleterious molecular events, discerning the locations of CRL within genome or plasmid where possible and observing trends within clonal groups.

Targeted PCR, Sanger sequencing and next generation short and long read genome sequencing techniques in conjunction with bioinformatic analyses were used to characterise clinical *E. coli* samples from Sydney Adventist Hospital isolated between 2009 and 2011.

Our data suggests that antibiotic resistance is readily transferring between host populations via lateral gene transfer of mobile elements in Sydney through the observation of unique molecular signatures. Clonal groups were identified in the cohort which share virulence and antimicrobial resistance traits, in some cases at seemingly differing stages of evolution. Large scale studies such as this provide insight to the mechanisms and forces driving the dissemination of antimicrobial resistance.

## **Contents**

Certificate of Authorship .....	2
Peer Reviewed Publications.....	3
Conference Presentations.....	4
Abstract.....	5
Abbreviations.....	12
Chapter 1: Background .....	14
<b>1.1 Antibiotics and the Evolution of Resistance</b> .....	15
<b>1.2 Lateral Gene Transfer</b> .....	18
<b>1.2.1 The Concept</b> .....	18
<b>1.2.2 Homologous Recombination</b> .....	20
<b>1.2.3 Site-specific Recombination</b> .....	21
<b>1.3 Established <i>Escherichia coli</i> Lineages</b> .....	21
<b>1.3.1 Lateral Gene Transfer and Drug Resistance in <i>E. coli</i></b> .....	24
<b>1.4 Mobile Genetic Elements</b> .....	25
<b>1.4.1 Plasmids</b> .....	26
<b>1.4.2 Transposons and Insertion Elements</b> .....	29
<b>1.4.3 Integrons</b> .....	30
<b>1.4.4 Genomic Islands</b> .....	33
<b>1.4.5 Complex Resistance Loci</b> .....	33
<b>1.5 Class 1 Integrons: Evolution and Zoonosis</b> .....	37
<b>1.6 Experimental Rationale and Approach</b> .....	40
<b>1.6.1 List of Methodologies and Software</b> .....	41
Chapter 2: A draft genome of <i>Escherichia coli</i> sequence type 127 strain 2009-46 .....	44
<b>Abstract</b> .....	46

<b>Background</b> .....	46
<b>Results</b> .....	46
<b>Conclusion</b> .....	46
<b>Background</b> .....	46
<b>Methods</b> .....	47
<b>Genome sequencing</b> .....	47
<b>Assembly and annotation</b> .....	48
<b>Quality assurance</b> .....	48
<b>Initial findings</b> .....	48
<b>Antibiotic resistance profile</b> .....	50
<b>Future directions</b> .....	51
<b>Availability of supporting data</b> .....	51
<b>Competing interests</b> .....	51
<b>Authors' contributions</b> .....	51
<b>Acknowledgements</b> .....	51
<b>References</b> .....	55
Chapter 3: Multidrug resistant uropathogenic <i>Escherichia coli</i> ST405 with a novel, composite IS26 transposon in a unique chromosomal location .....	
	57
<b>Abstract</b> .....	59
<b>1. Introduction</b> .....	60
<b>2. Materials and Methods</b> .....	61
<b>2.1 Strains, isolation and culture conditions</b> .....	61
<b>2.2 DNA purification</b> .....	61
<b>2.3 Fosmid library construction, screening and PCR conditions</b> .....	62
<b>2.4 Amplicon and whole genomes sequencing</b> .....	62

<b>2.5 Bioinformatics</b> .....	63
<b>3. Results</b> .....	63
<b>3.1 Genomic analysis of strains 2009-27 and 2009-30</b> .....	63
<b>3.2 Plasmid analysis</b> .....	64
<b>3.3 Characterisation of the CRL in strain 2009-27</b> .....	65
<b>3.4 Prevalence of Tn6242</b> .....	66
<b>3.5 Comparative phylogenomics of ST405 strains</b> .....	66
<b>3.6 Evolution of Tn6242</b> .....	67
<b>3.7 Generation of laterally mobile translocatable units from Tn6242</b> .....	68
<b>4. Discussion</b> .....	68
<b>Acknowledgements</b> .....	70
<b>Author contributions</b> .....	70
<b>Conflict of Interest</b> .....	70
<b>Ethics statement</b> .....	70
<b>References</b> .....	75
Chapter 4: Whole genome analysis of ExPEC ST73 from a single hospital over a 2-year period	
identified different circulating clonal groups.....	80
<b>Abstract</b> .....	82
<b>Data Summary</b> .....	82
<b>Impact Statement</b> .....	83
<b>Introduction</b> .....	84
<b>Methods</b> .....	85
<b>Isolate source and culture conditions</b> .....	85
<b>Nucleic acid purification and whole genome sequencing</b> .....	86
<b>Genome assembly and gene presence</b> .....	86



<b>Archived sequence read selection</b> .....	86
<b>S1-PFGE analysis</b> .....	87
<b>SNP based phylogenetic analyses</b> .....	87
<b>Results</b> .....	88
<b>Assembly information and statistics</b> .....	88
<b>Public read high-throughput sequencing analysis</b> .....	88
<b>Virulence profiles of Sydney strains</b> .....	89
<b>Antibiotic resistance</b> .....	90
<b>Structure of class 1 integrons in ST73 strains from Sydney</b> .....	91
<b>Discussion</b> .....	93
<b>Author statements</b> .....	95
<b>Funding information</b> .....	95
<b>Acknowledgements</b> .....	96
<b>Ethical statement</b> .....	96
<b>Conflicts of interest</b> .....	96
<b>Abbreviations</b> .....	96
<b>References</b> .....	103
<b>Chapter 5: Genomic analysis of multidrug-resistant <i>Escherichia coli</i> ST58 causing urosepsis.</b>	108
<b>Abstract</b> .....	110
<b>1. Introduction</b> .....	111
<b>Materials and Methods</b> .....	112
<b>2.1. Bacterial strains</b> .....	112
<b>2.2. Calibrated dichotomous sensitivity (CDS) testing</b> .....	112
<b>2.3. Whole-genome sequencing and analyses</b> .....	113
<b>2.4. Single nucleotide polymorphisms (SNPs)</b> .....	113

<b>2.5. Nucleotide sequence accession nos.</b> .....	114
<b>3. Results and Discussion</b> .....	114
<b>3.1. Whole-genome sequencing and comparative genomic analyses</b> .....	114
<b>3.2. Virulence-associated genes (VAGs) in ST58 strains</b> .....	115
<b>3.3. Characterisation of the complex resistance locus in the Sydney ST58 strains</b> .....	115
<b>3.4. pSDJ2009-52F carries the Tn1721/Tn21 hybrid transposon</b> .....	116
<b>4. Conclusion</b> .....	117
<b>Acknowledgments</b> .....	118
<b>Funding</b> .....	118
<b>Competing interests</b> .....	118
<b>Ethical approval</b> .....	118
<b>References</b> .....	123
 Chapter 6: Molecular analysis of an IncF ColV-like plasmid lineage that carries a complex resistance locus with a trackable genetic signature.....	 125
<b>Abstract</b> .....	127
<b>Introduction:</b> .....	127
<b>Methods:</b> .....	129
<b>Sequencing of pSDJ2009-52F</b> .....	129
<b>Single Nucleotide Polymorphism (SNP) Analysis</b> .....	129
<b>BEAST2 Analysis</b> .....	129
<b>Results:</b> .....	130
<b>Plasmid Information</b> .....	130
<b>Virulence and Antibiotic Resistance Gene Carriage</b> .....	130
<b>Comparative SNP Analyses</b> .....	131
<b>BEAST2 Analysis</b> .....	132

<b>Molecular Signatures</b> .....	132
<b>Discussion</b> .....	133
<b>CRedit Statement</b> .....	134
<b>Funding Statement</b> .....	135
<b>References</b> .....	141
Chapter 7: General Discussion.....	144
<b>7.1 Dominant Lineages in Clinical ExPEC and Trends in Antibiotic Resistance</b> .....	145
<b>7.2 IS26-mediated Insertion Signatures and Zoonosis</b> .....	150
<b>7.3.1 Limitations</b> .....	156
References.....	158

## **Abbreviations**

AIEC	adherent-invasive <i>E. coli</i>
APEC	avian pathogenic <i>E. coli</i>
bp	base pair
CDS	calibrated dichotomous sensitivity
CRL	complex resistance loci/locus
CS	conserved segment
DAEC	diffusely adhering <i>E. coli</i>
EAEC	enteroaggregative <i>E. coli</i>
EHEC	enterohemorrhagic <i>E. coli</i>
EIEC	enteroinvasive <i>E. coli</i>
EPEC	enteropathogenic <i>E. coli</i>
ESBL	extended spectrum $\beta$ -lactamase
ETEC	enterotoxigenic <i>E. coli</i>
ExPEC	extra-intestinal pathogenic <i>E. coli</i>
GI	genomic island
GIT	gastro-intestinal tract
HC	hemorrhagic colitis
HUS	haemolytic uraemic syndrome
Inc	incompatibility
IPEC	intestinal pathogenic <i>E. coli</i>
IS	insertion sequence
kb	kilobase
LB	Luria-Bertani/lysogeny broth
LGT	lateral gene transfer
MDR	multi-drug resistant
MGE	mobile genetic element
MLST	multi-locus sequence typing

NMEC	neonatal meningitis <i>E. coli</i>
nt	nucleotide
PCR	polymerase chain reaction
PFGE	pulsed field gel electrophoresis
SMAC	sorbitol-MacConkey agar
SMRT	single molecule real time
SNP	single nucleotide polymorphism
SRA	sequence read archive
ST	sequence type
TU	translocatable unit
UPEC	uropathogenic <i>E. coli</i>
UTI	urinary tract infection
VAG	virulence-associated gene