1	Promising yet variable performance of	f cross-taxon biodiversity	surrogates: a test in two marine
---	---------------------------------------	----------------------------	----------------------------------

- 2 habitats at multiple times
- 3
- 4 William Gladstone^{1*} (https://orcid.org/0000-0003-4517-4605), Brad Murray¹
- 5 (https://orcid.org/0000-0002-4734-5976) and Pat Hutchings^{3,4} (https://orcid.org/0000-0001-7521-
- 6 3930)
- 7 1. Faculty of Science, University of Technology Sydney, PO Box 123, Broadway NSW 2007, Australia
- 8 3. Australian Museum Research Institute, Australian Museum, 1 William Street, Sydney NSW 2010,
- 9 Australia
- 10 4. Department of Biological Sciences, Macquarie University, North Ryde, NSW 2109, Australia
- 11 * corresponding author: William.Gladstone@uts.edu.au
- 12

13	Acknowledgements We thank the Department of Defence for funding this project through CSIRO
14	Division of Fisheries; Angela Meza, Fiona Mackillop and Rob Patterson for assistance in the collection
15	and sorting of samples; Ian Loch and Winston Ponder for assistance in identification of molluscs;
16	Pauline McWilliams for assistance in identification of crustaceans and Roger Springthorpe for
17	assistance with the decapods; Winston Ponder for assistance with the mollusc literature; and Daniel
18	Krix for producing the map of study sites and graphs of the surrogates' biodiversity variables. A

19 representative collection of the fauna is deposited in the Australian Museum.

20 Abstract

21 Surrogates are a potential solution to the often-cited problem of there being insufficient information 22 for biodiversity assessments or conservation planning. Cross-taxon surrogacy is the ability of a group 23 of well-known taxa to represent variation in other poorly known taxa. To date, tests of the 24 effectiveness of cross-taxon surrogacy in marine environments have yielded variable results and a 25 significant qualification to the outcomes of tests that have demonstrated surrogacy is the near 26 absence of tests for its persistence through time. This study tested for cross-taxon surrogacy and its 27 persistence through time for three surrogates (crustaceans, molluscs, polychaetes). We used data on 28 biodiversity variables and species assemblages of the surrogates and their targets that had been 29 collected during multiple sampling events over 2.5 yr in two habitats (seagrass, unvegetated 30 sediment) in a large bay in south-eastern Australia. We tested surrogacy by fitting a series of linear 31 models using generalized least squares for biodiversity variables and by Mantel tests of dissimilarity 32 matrices of species assemblages. We also tested whether the type of data transformation affected 33 Mantel tests. We found that each of the groups were effective surrogates for some but not all 34 biodiversity variables (with molluscs or polychaetes being effective surrogates for species richness in 35 both habitats), that none of the groups were effective surrogates for species assemblages, and that the outcomes of Mantel tests of dissimilarity matrices of surrogates and their targets were 36 37 unaffected by the data transformation used. We conclude that while our results for surrogacy for 38 biodiversity variables are promising the inconsistent results from other studies argues for caution 39 about their application beyond the area and context in which they were assessed. The lack of 40 evidence that we found for surrogates of species assemblages, and similar lack of evidence from 41 other studies, suggests that assemblage-level surrogates are unlikely to be useful in biodiversity 42 surveys.

43

- **Keywords** Biodiversity assessment, Conservation planning, Ecological indicators, Jervis Bay,
- *Posidonia*, Species richness

46 Introduction

47 In the absence of detailed information on the distribution of biodiversity, assessment or 48 conservation planning, some form of surrogacy is typically used (Margules and Pressey 2000). A 49 surrogate represents spatial and temporal variation in biodiversity and when used in conservation 50 planning (as a proxy for detailed information on a biodiversity conservation target) leads to 51 adequate representation of the target in reserves (Sutcliffe et al. 2015). The need for a surrogate 52 approach to conservation planning in the marine realm arises from the limited information on the 53 extent and distribution of biodiversity and its patterns of temporal variation, and the logistical (e.g. 54 time, finances) and technical (e.g. incomplete taxonomy) constraints to obtaining detailed 55 information on biodiversity in a planning region in the typical time scales of conservation decisionmaking (Fisher et al. 2011, Menegotto and Rangel, 2018, Valesini et al. 2018). 56

57 The ability of a group of well-known taxa to represent variation in other poorly known taxa is 58 called cross-taxon surrogacy. Surrogacy is a pragmatic approach to conservation planning in the face 59 of limited information on biodiversity and the pressures to make planning decisions (Grantham et al. 60 2009). Studies of cross-taxon surrogacy performance have tested for spatial congruence between 61 putative surrogates and their targets in patterns of species richness, species composition, or sets of 62 representative reserves. A number of comprehensive reviews of these studies have found variable 63 evidence for surrogate effectiveness and cautioned against the general application of the cross-64 taxon surrogacy approach in terrestrial (Rodrigues and Brooks 2007, Eglington et al. 2012, Westgate et al. 2014, Oberprieler et al. 2019) and freshwater aquatic ecosystems (Allen et al. 1999, Heino 65 66 2010, Velghe and Gregory-Eaves 2013).

Tests of the effectiveness of cross-taxon surrogacy in marine environments have similarly
yielded variable results. Molluscs on coastal, intertidal rocky shores and in estuarine habitats
effectively represented spatial variation in species richness of other organisms (Gladstone 2002,
Smith 2005, Shokri et al. 2009), and simulated conservation reserves planned using molluscs

71 performed significantly better than random selection of sites at representing other species 72 (Gladstone 2002, Shokri et al. 2009). The species richness of polychaetes predicted the species 73 richness of other groups in coastal Europe (Olsgard et al. 2003) and in a single estuary in Australia 74 (Shokri et al. 2009). The species richness of corals and fishes was correlated at a trans-oceanic scale 75 (Hughes et al. 2002) but not at the scale of a single bay (Beger et al. 2003). Spatial patterns in the 76 species richness and assemblages of algae, macro-invertebrates and fishes on coastal sub-tidal reefs 77 in south-east Australia are uncorrelated and none of the putative surrogates performed better than 78 random selection in representing their target groups in simulated reserve networks (Gladstone and 79 Owen 2005). On the shelf seabed of the Great Barrier Reef there was little congruency amongst 80 different assemblages of different groups of organisms (Sutcliffe et al. 2012). Spatial variations in 81 assemblages of macro- and meiofauna inhabiting soft-sediments in a bay in Brazil were significantly 82 correlated, suggesting one group was an effective surrogate of the other (Corte et al. 2017). Rarity 83 values of a range of phyla from subtidal rocky reefs in Brittany (western France) were poorly 84 correlated, indicating no potential for cross-taxon surrogacy (Leroy et al. 2017). Ciliated protozoans 85 are suitable surrogates for the diversity of the eukaryotic microperiphyton community (Xu et al. 2015). 86

87 This variability in the existence of surrogacy is also emphasised when the outcomes of studies on the same group of organisms are compared. Polychaetes have been shown to be useful 88 89 surrogates by some studies (Olsgard et al. 2003, Magierowski and Johnson 2006, Shokri et al. 2009) 90 but not others (Olsgard and Somerfield 2000). Molluscs have been shown to be useful surrogates by 91 some studies (Gladstone 2002, Smith 2005) but not others (Olsgard and Somerfield 2000, 92 Magierowski and Johnson 2006). And, while some studies concluded that crustaceans were suitable 93 surrogates (Magierowski and Johnson 2006) others found no evidence of surrogacy (Olsgard and 94 Somerfield 2000, Smith 2005).

95 The lack of a clear and consistent outcome from the different tests of cross-taxon surrogacy 96 can be attributed to a number of interacting causes: the range of spatial scales and habitats that 97 have been used; the existence (or non-existence) of an environmental gradient in the study area; 98 differences in the ecologies and life histories of surrogates and their targets; latitudinal differences 99 among the published studies; variation in sampling effort (i.e. size of sampling unit and number of 100 replicate samples); differences in the units of assessment (i.e. species or assemblages); and the use 101 of different variables and statistical analyses to assess surrogate effectiveness and decide on its 102 presence or absence (Hess et al. 2006, Lawler and White 2008, Grantham et al. 2010, Westgate et al. 103 2014, 2017).

104 Notwithstanding these possible explanations for the lack of consistent evidence for cross-105 taxon surrogacy, a significant qualification to the conclusions of studies that have demonstrated 106 cross-taxon surrogacy is the near-absence of tests for temporal consistency (Mellin et al. 2011, 107 Westgate et al. 2014). A significant correlation between a putative surrogate and its target groups 108 based on one period of sampling could be a one-off result (e.g. a spurious correlation arising from 109 incomplete sampling, Neeson et al. (2013)) and not representative of long-term or underlying 110 relationships among taxa. If so, this could be a major limitation to the application of this form of 111 surrogacy in biodiversity assessment, marine conservation planning, and environmental assessment. 112 In addition, tests for the existence of cross-taxon surrogacy based on multivariate assemblages 113 typically use correlations of similarity matrices with some form of data transformation prior to 114 analysis. Transformation of species' abundances in multivariate data sets is done to reduce the 115 influence of numerically dominant species or erratic differences between sampled locations (Clarke 116 and Gorley 2015). Tests of surrogates of multivariate assemblages have typically used only a single 117 data transformation e.g. presence-absence (Beger et al. 2003), log(X+1) (Corte et al. 2017), square root (Olsgard and Somerfield 2000), or fourth root (Magierowski and Johnson 2006). Given the 118 119 findings that other types of analyses of multivariate assemblages are affected by the choice of data

transformation (Anderson et al. 2005) it is important to understand whether the type of datatransformation affects analyses of surrogacy effectiveness.

122 Therefore, the aim of this study was to test for cross-taxon surrogacy and its persistence through 123 time. We used data on three groups of marine invertebrates (crustaceans, molluscs, polychaetes) 124 that had been collected during multiple sampling events over 2.5 yr. In order to improve the 125 generality of the results we undertook the study in two habitats (seagrass, unvegetated sediment) at 126 the same location. Specifically, we tested the following null hypotheses: (1) biodiversity variables of 127 the surrogates are not related to the same variables in their target through time and in different 128 habitats; (2) pairwise patterns among sites of assemblage dissimilarity for a surrogate and its target 129 are not correlated; and (3) the size of the correlations between the dissimilarity matrices of a 130 surrogate and its target are unaffected by the type of data transformation used.

131

132 Materials and methods

133 Study area

134 This study was undertaken in Jervis Bay, a marine embayment in southern New South Wales (NSW), 135 Australia (35°08'S, 150°45'E) (Fig. 1). At depths <10 m the predominant habitats are either beds of the seagrass Posidonia australis or rocky reefs and at depths >10 m the predominant habitat is 136 137 unvegetated sandy mud. Jervis Bay is currently managed by two marine protected areas: the NSW 138 State-managed Jervis Bay Marine Park, and the Commonwealth of Australia-managed Booderee 139 National Park. Much of the surrounding terrestrial area is within State national parks with limited 140 residential development and no major rivers flowing into the Bay. At the time of this study, Jervis 141 Bay was regarded as relatively pristine. Water temperatures during the study varied between 14-142 15°C (July-August) and 23-24.5°C (January-February) (CSIRO 1993). As the sampling sites spanned the

extent of Jervis Bay, the spatial extent of this study was the area of the Bay i.e. 102 km² (Hutchings
and Jacoby 1994).

145 Field sampling

Four sites in beds of the seagrass *P. australis* and six sites of sandy/muddy substratum that was unvegetated (hereafter called unvegetated sediment) that spanned the breadth of Jervis Bay were sampled (Fig. 1). The seagrass sites were in depths of 2-6 m (mean±SE=4.0±0.9 m) and the unvegetated sediment sites were in depths of 12-20 m (14.7±1.7 m). Each site was sampled on nine occasions between February 1989 and June 1991, with intervals of 2-5 mo between successive sampling events.

152 Samples from unvegetated sediment sites were collected by Smith-McIntyre grab (sample 153 area 0.06 m^2 and sample volume 4.7 L) deployed from a boat. At each site, five replicate samples 154 were collected from an area of several hundred square metres by allowing the boat to drift. All grab 155 samples were collected on the same day. Samples at the seagrass sites were collected by SCUBA 156 divers using hand-operated corers. The corers had an internal diameter of 0.11 m and sampled an 157 area of 0.0095 m² and a volume of 0.95 L of sediment (when pushed into the sediment to a depth of 0.10 m). At each site four replicate cores were collected from each of two plots (2 m x 2 m) that 158 159 were approximately 50 m apart, at each sampling event. The vegetated sites were sampled over a 160 number of days during the same week as the grab samples were collected and all field sampling in 161 each sampling event was completed within one week. For consistency in the field collection the 162 same two divers collected all seagrass samples throughout the study, the same crew operated the 163 grabs, and the same people sorted samples in the lab. Different sampling techniques were used in 164 the two habitats because the seagrass beds were too shallow for effective deployment of a grab, and the unvegetated sediment sites were too deep for repetitive diving over a limited number of 165 166 days. The data were analysed separately for each habitat, and interpretations limited to the 167 temporal variation in cross-taxon surrogacy within each habitat rather than a comparison between

the habitats, to avoid confounding because of the different methods and physical scales of samplingin the two habitats.

170 In the field, the collected samples were put into bags made of 1.0 mm mesh, quickly washed 171 to remove sediment then immediately placed in a polydrum containing 7% neutralised seawater 172 formalin and Biebricht Scarlet (to stain all living organisms) and gently agitated. After 4-5 d the mesh 173 bags were removed and carefully washed under a running tap to remove all the formalin and 174 remaining sediment and then the contents of each bag placed into 70% alcohol. Samples were then 175 sorted in the laboratory under a dissecting microscope and identified to species with a reference 176 collection being deposited at the Australian Museum.

177 Data analyses

178 Analyses were done for two data sets: four sampling events (that spanned 10 mo), and nine 179 sampling events (that spanned 28 mo). In the test of four sampling events the surrogates tested 180 were crustaceans, molluscs, and polychaetes and their target groups were, respectively, molluscs + 181 polychaetes, crustaceans + polychaetes, and crustaceans + molluscs. In the test of nine sampling 182 events the surrogate was molluscs and the target was polychaetes (data for crustaceans were 183 unavailable), and we recognise that because it is a bivariate relationship the results are 184 interchangeable (i.e. polychaetes as a surrogate). The following biodiversity variables were 185 calculated for each surrogate and its target group: species richness; total number of individuals; 186 Margalef's index of species richness, which accounts for the numbers of individuals in a sample; 187 Pielou's evenness index, which measures how equitability individuals are distributed among the 188 species in a sample; and Shannon-Wiener diversity index (calculated to log base e) (Magurran 2003). 189 Variables were calculated using PRIMER 7 software (PRIMER-E, Plymouth).

In the analyses that follow correlations ≥0.7 were used as evidence of strong relationships
between surrogates and their targets, with the surrogate being suitable for biodiversity surveys and

conservation planning (Vellend et al. 2008, Heino 2010). In the tests of the biodiversity variables the relevant correlation was the partial correlations ($_{p}r$) after taking time into account, and in the tests of assemblages, the relevant correlation was the Mantel correlation coefficient (r_{M}).

195 To test the null hypothesis that biodiversity variables of the surrogates are not related to the 196 same variables in their target through time we fit a series of linear models using generalized least 197 squares in R (R Core Team 2019). Each model determined whether an individual taxon (crustaceans, 198 molluscs, polychaetes) was an effective biodiversity surrogate for the other two taxa combined. For 199 example, the species richness of crustaceans (the surrogate) was modelled in relation to the 200 combined species richness of molluscs and polychaetes. Separate models to examine the strength of 201 such taxon surrogacy in each habitat (seagrass, unvegetated sediment) were built for each of the 202 five biodiversity variables. The mean values of each variable at each site at each time were used. To 203 account for repeated measurements over time at the same sites, we used the nlme package 204 (Pinheiro et al. 2019) and the function corAR1 to specify a temporal autocorrelation structure of 205 order one in the models (Mangiafico 2016). After including time as a potential source of variation in 206 the models in this way, we were able to determine the strength of each taxon as a surrogate over 207 and above any influence of time through the use of partial correlation coefficients ($_{p}r$). We used a 208 likelihood ratio (LR) test via the nagelkerke function (Mangiafico 2016) to assess statistical 209 significance and to calculate pseudo R^2 values (from which pr values were calculated) for each taxon 210 as a surrogate (Crawley 2012). The LR tests assessed the significance of the change in deviance (χ^2) 211 when the full model (surrogate and time) was compared with a reduced model (time only). All 212 models were inspected for normality of residuals and homogeneity of variances, with species 213 richness log-transformed in seagrass habitat to meet model assumptions.

For the assessment of multivariate assemblages of surrogates and their targets, the mean abundance of each species at each site was used (n=4 seagrass sites, n=6 unvegetated sediment sites). Patterns of assemblage dissimilarity among sites were visualised by non-metric MDS

217 ordination plots, based on Bray-Curtis dissimilarity matrices of square-root transformed abundance 218 data, for each pair of surrogate and target for each sampling event, in each habitat. Analyses were 219 done with PRIMER 7 software (PRIMER-e, Quest Research Ltd). The null hypothesis that the pairwise 220 patterns among sites of assemblage dissimilarity for a surrogate and its target are not correlated was 221 tested by Mantel test (Heino 2010, Ilg and Oertli 2016, Yong et al. 2018). Mantel correlation 222 coefficients ($R_{\rm M}$) were calculated from the Bray-Curtis dissimilarity matrices of the surrogate and its 223 target. Surrogate-target relationships with $R_{M} \ge 0.7$ were further investigated by partial Mantel 224 correlation coefficients (pR_M), using a third matrix of pairwise physical distances between sites (see 225 also Su et al. 2004, Padial et al. 2012, Ilg and Oertli 2016), to account for the possibility that the 226 correlations were confounded by the different pairwise distances among sites (Fig. 1). Mantel tests 227 were conducted with PC-Ord v 7.08 (McCune and Mefford 2018). The statistical significance of the 228 pR_M values is not reported because the small number of maximum possible permutations of the 229 dissimilarity matrices did not allow for meaningful estimates of P-values (Manly 1997).

230 Prior to the above analyses, separate dissimilarity matrices were constructed in which the 231 abundance data of each species were transformed to square-root, log(X+1), or presence-absence to 232 account for the possible effect of transformation on the Mantel test (Olsgard et al. 1997, Anderson 233 et al. 2005, Mueller et al. 2013). One-way analysis of variance (ANOVA) was used to test the null 234 hypothesis that mean R_M did not differ among data transformations for each of the data sets of four 235 and nine sampling events in each habitat. Data transformation was analysed as a fixed factor with 3 236 levels (square-root, log(X+1), presence-absence). The replicates for each level were the set of R_{M^-} 237 values for all times, in each habitat. The assumption of equality of variances was tested prior to 238 ANOVA by Levene's test.

239 Results

240 Biodiversity

241 The data set for the four sampling events included (i) crustaceans: 154 species (981 individuals) in 242 seagrass and 185 species (16,391 individuals) in unvegetated sediment; (ii) molluscs: 72 species 243 (1,779 individuals) in seagrass and 110 species (4,300 individuals) in unvegetated sediment; and (iii) 244 polychaetes: 85 species (1,666 individuals) in seagrass and 141 species (8,351 individuals) in 245 unvegetated sediment. The data set for the nine sampling events included (i) molluscs: 97 species 246 (3,335 individuals) in seagrass and 178 species (9,537 individuals) in unvegetated sediment; and (ii) 247 polychaetes: 105 species (2,815 individuals) in seagrass and 166 species (15,248 individuals) in 248 unvegetated sediment.

249 Biodiversity variables

250 Seagrass

251 In the set of four sampling events each of the three surrogates showed statistically significant 252 relationships with their targets for most biodiversity variables through time (Table 1, Fig 2). The only 253 exceptions were for the surrogate crustaceans (Margalef's index), molluscs (total number of 254 individuals, Pielou's evenness index) and polychaetes (Pielou's evenness index). The surrogate 255 relationships that exceeded the r=0.7 threshold were crustaceans (total number of individuals 256 $_{p}$ r=0.89), molluscs (species richness $_{p}$ r=0.79, Shannon-Weiner diversity index $_{p}$ r=0.81), and 257 polychaetes (species richness $_{p}$ r=0.73, Shannon-Weiner diversity index $_{p}$ r=0.76). For the test of 258 molluscs as a surrogate of polychaetes in the set of nine sampling events, relationships between 259 surrogate and target were statistically significant for most biodiversity variables (with the exception 260 of total number of individuals); however, none of the tests exceeded the r=0.7 threshold (Table 1, 261 Fig 3). The largest correlation was for species richness (pr=0.60).

262 Unvegetated sediment

In the set of four sampling events there was a statistically significant relationship between each
surrogate and its target through time for most biodiversity variables, except for crustaceans (Pielou's
evenness index, Shannon-Weiner diversity index), molluscs (Pielou's evenness index, Shannon-

Weiner diversity index), and polychaetes (Shannon-Weiner diversity index) (Fig. 3, Table 2). The surrogate-target relationships that exceeded the r=0.7 threshold included crustaceans (species richness pr=0.75, total number of individuals pr=0.75), molluscs (species richness pr=0.89, total number of individuals pr=0.82), and polychaetes (species richness pr=0.73). In the set of nine sampling events the relationship between the surrogate (molluscs) and target (polychaetes) was statistically significant for all biodiversity variables except Pielou's evenness index and Shannon-Weiner diversity index, and none of the tests had pr of at least 0.7 (Fig.4, Table 2).

273 Assemblages

Mean $R_{\rm M}$ values between surrogates and targets did not significantly differ among the three different data transformations (square-root, log(X+1), presence-absence) in both seagrass and unvegetated sediment (Fig. 5). The error bars indicate that $R_{\rm M}$ values varied among the sampling events for all transformations, and in some sampling events $R_{\rm M}$ values were negative. When this occurred, the $R_{\rm M}$ values were negative for all transformations. As a result of the lack of significant differences among data transformations, only analyses of the square root-transformed data are presented in the following section.

281 Seagrass

282 nMDS ordination plots of surrogates and targets that showed the spread of sites according to 283 relative dissimilarity of assemblages showed few concordant pattens for each of the four (Fig 6) or 284 nine (Fig 7) sampling events. For example, in event 1 there was a near-equidistant spread of all sites 285 for crustaceans (the putative surrogate) but a distinct cluster of sites 1 and 2 for the target (the 286 assemblage of molluscs and polychaetes). There was a similar pattern of difference in event 4 287 between surrogate (molluscs) and target (crustaceans and polychaetes). In event 7 (Fig 7) Sites 3 and 288 4 clustered close together for the mollusc assemblage but were widely separated for the polychaete 289 assemblage. There were few examples of concordant patterns of dissimilarity among sites of the

surrogate and target in the four sampling events (e.g. polychaetes as surrogate (event 1), molluscs as
surrogate (event 3)) or nine sampling events (e.g. event 4).

292 Only one test of surrogacy in the four sampling events had a Mantel correlation coefficient 293 $(R_{\rm M}) \ge 0.7$: polychaetes in event 1 ($R_{\rm M}=0.83$, Table 3). This correlation decreased (but still exceeded 294 the 0.7 threshold) when the effect of distance between the sites was controlled for (pR_M =0.71). 295 Values of $R_{\rm M}$ changed considerably among sampling events for each of the surrogates tested, 296 including changing from positive to negative values. In the test of nine sampling events $R_{\rm M}$ for 297 molluscs and polychaetes was high for only one sampling event (event 9) and the value of $R_{\rm M}$ did not 298 alter when the distances between sites was controlled for. Otherwise, values of R_M changed 299 considerably from one sampling event to the next.

300 Unvegetated sediment

In unvegetated sediment there were distinct differences between surrogates and targets in the arrangement of the sites in the nMDS ordination plots for the set of four sampling events (Fig 8 e.g. event 1 with crustaceans as the surrogate, and event 3 with polychaetes as the surrogate) and the set of nine sampling events (Fig 9 e.g. events 1, 5 and 8). The arrangements of sites for the surrogate and target appeared to be similar for molluscs as the surrogate in event 4. There were no obvious examples of similar arrangements of sites for molluscs or polychaetes in any of the nine sampling events.

Results of the Mantel tests showed a high value of $R_{\rm M}$ (0.78) for only one test of surrogacy in the four sampling events: molluscs in event 1 (Table 4). This value increased slightly when the distances between sites were controlled for (p $R_{\rm M}$ =0.79). Other Mantel tests varied considerably for each of the surrogates in each of the sampling events and none approached 0.7. Only one Mantel test returned a high value in the set of nine sampling events for molluscs and polychaetes: 0.84 in event 9. This increased to p $R_{\rm M}$ =0.88 when distances among sites were controlled for.

314 Discussion

315 Performance of surrogates

316 Previous research has revealed no consistency in the performance of cross-taxon surrogates in 317 different habitats, spatial scales, and among different groups of fauna (Westgate et al. 2014), and 318 highlighted the lack of understanding about the persistence of cross-taxon surrogacy through time (Mellin et al. 2011, Westgate et al. 2017). We used data sets of marine invertebrates that were 319 320 collected at regular intervals for periods of 10 and 28 mo to test for the existence of cross-taxon 321 surrogacy and its persistence through time, and we tested for it in two habitats (seagrass, 322 unvegetated sediment). We assessed cross-taxon surrogacy by using generalised linear models (that 323 included time as a factor) to test for relationships between surrogates and their targets for several 324 biodiversity variables and by Mantel correlations to test for relationships between the dissimilarity 325 matrices of surrogates and their targets. Based on a threshold correlation of ≥0.7 as evidence of 326 surrogacy, we found that: (i) each of the tested surrogates had a strong relationship with its target 327 that persisted through time for one or more biodiversity variables in each habitat; (ii) there was no 328 consistent evidence for surrogacy in the species assemblages of either seagrass or unvegetated 329 sediment; and (iii) data transformation did not affect the size of Mantel correlation coefficients.

330 We defined *a priori* the evidence needed to reject the null hypothesis of no relationship 331 between a surrogate and its target, which was the magnitude of the correlation coefficient and its 332 persistence through time. An r-value of \geq 0.70 indicates that a substantial proportion of the variation 333 in the target could be explained by variation in the surrogate (Lovell et al. 2007, Vellend et al. 2008, 334 Heino 2010). A correlation of $r \ge 0.70$ after time has been taken into account, or in a majority of 335 sampling events, indicates that the correlation between surrogate and target is unlikely to be a 336 statistical anomaly (Neeson et al. 2013). Reasons for rejecting cross-taxon surrogacy in other studies 337 have included the non-significance of correlations (Beger et al. 2003, Gladstone and Owen 2007), 338 significant but small correlations (e.g. < 0.30 by Hirst 2008), or a combination of non-significance and

339 significant but small correlations (Karakassis et al. 2006, Leroy et al. 2017). Similarly, surrogacy has 340 been accepted as proven for a range of magnitudes of correlation coefficients (Gladstone 2002, 341 Olsgard et al. 2003, Smith 2005, Magierowski and Johnson 2006, Shokri et al. 2009, Corte et al. 342 2017). While the use of a standard criterion (e.g. $r \ge 0.70$) for accepting the existence of cross-taxon 343 surrogacy would potentially facilitate the application of surrogacy in conservation planning and 344 comparisons among studies, it would need to be established that the criterion was independent of 345 habitat, spatial scale, sampling effort, latitude, and the diversity of the putative surrogate group and 346 its targets.

347 Notwithstanding differences in the criteria used to decide whether or not surrogacy exists, 348 the results of this study show that conclusions about a group's performance as a cross-taxon 349 surrogate are not transferable. We found that crustaceans, molluscs and polychaetes were suitable 350 surrogates for some biodiversity variables in both seagrass and unvegetated sediment. While some 351 other studies also found that polychaetes (Olsgard and Somerfield 2000, Olsgard et al. 2003, 352 Magierowski and Johnson 2006, Shokri et al. 2009), molluscs (Gladstone 2002, Smith 2005), and 353 crustaceans (Magierowski and Johnson 2006) were effective surrogates, others have concluded that 354 polychaetes (Olsgard and Somerfield 2000), crustaceans (Olsgard and Somerfield 2000, Smith 2005) 355 and molluscs (Olsgard and Somerfield 2000, Magierowski and Johnson 2006) were unsuitable as 356 cross-taxon surrogates. Whilst some of these differences can be attributed to the different ways 357 surrogate effectiveness was judged, these studies were also conducted in different environments, 358 over different time scales, in different biogeographical regions, and with faunas of different 359 diversities. This further reinforces the caution expressed by other authors about the application of 360 conclusions about surrogates beyond the area and context in which they have been assessed (Mellin 361 et al. 2011, Westgate et al. 2014).

For one-third of the studies reviewed by Mellin et al. (2011) there was no relationship
between surrogates and their targets, and when there was a relationship the predictive power was

364 weak. Cross-taxon surrogates are expected to perform well when the surrogate and target co-vary 365 spatially and temporally and this may be more likely to occur at smaller spatial scales, when there is 366 a strong ecological or disturbance gradient that favours a surrogate, when the surrogate has a 367 diversity of life histories and ecologies that overlap those of the target group of species, or in low 368 complexity environments (Gaston and Williams 1996, Olsgard and Somerfield 2000, Mellin et al. 369 2011). Mellin et al. (2011) found that marine habitat type was the best predictor of surrogate 370 effectiveness, with low complexity marine habitats such as soft bottoms being best. We found, for 371 biodiversity variables, a similar number of acceptable surrogacy relationships in unvegetated 372 sediment and seagrass (a more complex marine habitat). We also found no difference between 373 these habitats in the lack of surrogacy relationships for species assemblages.

374 Each of the tested surrogates represented a variety of ecological roles and life histories that 375 we expected to overlap with the roles and life histories of their targets and to therefore be suitable 376 as surrogates. For example, the polychaetes in our data set were diverse and represented a range of 377 families, life cycles (from a few months to several years), reproductive strategies (from breeding 378 once then dying, to those which breed annually over several years), larval phases and durations 379 (including long-lived pelagic larvae, a very short larval phase, no larval phase), and a range of feeding 380 strategies (carnivores, filter-feeders, herbivores, opportunistic) (Hutchings 1998, Beesley et al. 2000, 381 Rouse and Pleijel 2006, Jumars et al. 2015). The molluscs were similarly diverse in ecological roles 382 and life histories: the bivalves include suspension feeders, deposit feeders, microcarnivores and 383 some obtain their nutrition via bacteria or zooxanthellae. Gastropods are equally diverse with 384 carnivores and grazers and some with symbioses, others are suspension feeders, parasites, 385 coprophages, and life cycles ranged from annual species to those that lived for several years (Brusca 386 et al. 2016, Lindberg et al. 2004, Ponder and Lindberg 2008). The absence of surrogacy in the test of 387 nine sampling events, which were based on a surrogate (molluscs) and a single target (polychaetes), 388 could be due to smaller degree of overlap in the above features compared to the test of a surrogate 389 and the combined set of two target groups.

390 The lack of evidence for surrogacy in the species assemblages of both habitats mirrors 391 results from other studies that have reported correlations that, while statistically significant, were 392 small in marine (Karakassis et al. 2006, Hirst 2008), aquatic (Heino 2010, Padial et al. 2012, Ilg and 393 Oertli 2016) and terrestrial environments (Irwin et al. 2014, Yong et al. 2018). These conclusions 394 have been consistent across a variety of methods used to test surrogacy of species assemblages, 395 including Mantel tests, Procrustes analysis, and RELATE tests (Beger et al. 2003, Hirst 2008, Heino 396 2010, Padial et al. 2012, Corte et al. 2017). A possible explanation for this poor performance of 397 assemblage-level surrogates in marine environments is environmental variation that differentially 398 affected the range of species comprising the surrogate and the target groups. For example: spatial 399 heterogeneity in the features of each habitat that differed among the sampled sites and influenced 400 the invertebrate biodiversity (Bell et al. 1988, Macfarlane and Booth 2001, Edgar and Barrett 2002, 401 Radke et al. 2011), temporal variation in the features of each habitat that influenced invertebrate 402 biodiversity (Duarte 1989, Ysebaert and Herman 2002, van der Wal et al. 2017), or the absence of a 403 strong ecological gradient in Jervis Bay among the sites sampled within each habitat (Przeslawski et 404 al. 2009, Clark et al. 2015).

405 Temporal variation in surrogacy

406 There are examples in the published literature of cross-taxon surrogacy studies that used 407 biodiversity data from a single sampling event, assessed surrogacy in the same ways as this study, 408 and concluded that surrogacy had been demonstrated (Gladstone 2002, Olsgard et al. 2003, Smith 409 2005, Shokri et al. 2009) or was absent (Beger et al. 2003, Hirst 2008, Sutcliffe et al. 2012). There are 410 fewer examples of studies that have directly tested for the persistence of surrogacy through time. 411 Magierowski and Johnson (2006) found changes through time in the magnitude of the goodness of 412 fit between some putative surrogates (molluscs, echinoderms) and their targets, and no changes for some other putative surrogates (for assemblages of macroinvertebrates inhabiting artificial kelp 413 414 holdfasts) over a total study time of 13 mo. Corte et al. (2017) found significant and large

415 correlations between assemblages of macro- and meiofauna in each of four time periods (spanning 416 11 mo), and concluded surrogacy was present. Olsgard and Somerfield (2000) tested cross-taxon 417 surrogacy in three different years that spanned six years and found that the correlations between 418 some putative surrogates and their targets varied dramatically among the three years (from small to 419 large correlation coefficients), while others were consistently large, showed less variation, and were 420 therefore suitable surrogates. Other approaches to testing surrogacy have integrated the influence 421 of time by pooling multiple samples that had been collected through time and analysing a larger 422 single data set and have concluded that surrogacy existed (Olsgard et al. 2003, Xu et al. 2015) or was 423 absent (Gladstone and Owen 2007). While we applied criteria of a specific and strong correlation 424 coefficient and its persistence through time, such threshold criteria have not been used in other 425 marine studies that assessed surrogacy through time.

426 The limited understanding of temporal variation in the existence or absence of surrogacy has 427 been highlighted by other authors (Mellin et al. 2011, Westgate et al. 2017). We used persistence 428 through time as one criterion for evidence of surrogacy. We found that for some surrogates and 429 some biodiversity variables, strong correlations persisted through time. We also found that 430 surrogacy relationships in species assemblages did not persist through time. Our results highlight the 431 importance of sampling at multiple times to assess surrogacy performance. A single sampling event 432 done at the time of sampling event 1 would have concluded that polychaetes (in seagrass) and 433 molluscs (in unvegetated sediment) were suitable surrogates for the species assemblages of the 434 target groups crustaceans and molluscs, and crustaceans and polychaetes respectively. A strong 435 correlation between a surrogate and its target would be expected to persist through time if both 436 groups varied in the same way in most or all the sampled sites.

437 Effect of data transformation

438 Sampled invertebrate assemblages typically include a few species with much greater abundances
439 that may vary differently through time (e.g. Morrisey et al. 1992, Ysebaert and Herman 2002,

440 Marshall et al. 2018) and therefore some form of data transformation is required. While tools are 441 available to assist decisions about the transformation to use (e.g. Clarke et al. 2014, Clarke and 442 Gorley 2015), the decision in a test related to a biodiversity assessment or conservation objective 443 should depend on the hypothesis being tested or the objective of the assessment or conservation 444 planning (e.g. conservation of species' occurrences, or representative assemblages) and is therefore 445 ultimately an a priori decision. Tests of multivariate cross-taxon surrogacy have typically used only a 446 single data transformation including presence-absence (Beger et al. 2003), log(X+1) (Corte et al. 447 2017), square root (Olsgard and Somerfield 2000), and fourth root (Magierowski and Johnson 2006). 448 Other types of analyses of multivariate assemblages are affected by the choice of data 449 transformation (Anderson et al. 2005) and it is therefore important to understand the effect of 450 transformation on analyses of surrogacy effectiveness. Shokri et al. (2009) compared two 451 transformations and found only slight differences in the magnitudes of correlation coefficients that 452 did not alter their conclusions about surrogate effectiveness. However, if they had used the same 453 threshold criterion of $r \ge 0.70$ their conclusions would have been influenced by the type of 454 transformations. We showed in this study that the magnitude of the correlation coefficients, and 455 therefore the decision about the existence of surrogacy, was not influenced by the type of 456 transformation used.

457 Conclusions

Along with this study, studies of the usefulness of cross-taxon surrogacy in biodiversity assessments have yielded variable results. Our study, which also incorporated a test of temporal consistency, found that if the objective is to assess species richness in seagrass or unvegetated sediment then molluscs or polychaetes would be suitable surrogates. Considering the two habitats separately, crustaceans or molluscs would be suitable surrogates in seagrass, and molluscs or polychaetes would be suitable surrogates in unvegetated sediment. None of the surrogates we tested were suitable as surrogates for species assemblages. Our results highlighted the importance of testing surrogacy at

465 multiple times. We showed that conclusions about the performance of a group as a surrogate are 466 not transferable, and while our results for surrogacy for biodiversity variables are promising the inconsistent results from other studies argues for caution about their application beyond the area 467 468 and context in which they were assessed. While studies of surrogates of species assemblages have 469 applied a range of data transformations prior to creation of dissimilarity matrices, and results of 470 ecological studies are affected by the type of data transformation used, we found that different data 471 transformations did not significantly affect the outcomes of Mantel tests, and authors should adopt 472 the transformation relevant to the characteristics of sampled assemblage and/or the objective of the 473 research. The lack of support that we found for surrogates of species assemblages in both habitats, and evidence from other studies also showing weak correlations, suggests that surrogates for 474 475 assemblages are unlikely to be useful in biodiversity surveys.

476 References

- 477 Allen AP, Whittier TR, Larsen DP, Kaufmann PR, O'Connor RJ, Hughes RM, Stemberger RS, Dixit SS,
- 478 Brinkhurst RO, Herlihy AT, Paulsen SG (1999) Concordance of taxonomic composition patterns across
- 479 multiple lake assemblages: effects of scale, body size, and land use. Can J Fish Aq Sci 56:2029-2040
- 480 Anderson MJ, Connell SD, Gillanders BM, Diebel CE, Blom WM, Saunders JE, Landers TJ (2005)
- 481 Relationships between taxonomic resolution and spatial scales of multivariate variation. J Anim Ecol
 482 74:636–646
- 483 Anderson T, Brooke B, Radke L, McArthur M, Hughes M (2009) Mapping and characterising soft-
- 484 sediment habitats and evaluating physical variables as surrogates of biodiversity in Jervis Bay, NSW.
- 485 Geoscience Australia Record 2009/10. Geoscience Australia, Canberra
- 486 Beesley P.L, Ross GJB, Glasby CJ (eds) (2000) Polychaetes & Allies: The Southern Synthesis. Fauna of
- 487 Australia. Vol. 4A Polychaeta, Myzostomida, Pogonophora, Echiura, Sipuncula. CSIRO Publishing,
- 488 Melbourne
- 489 Beger M, Jones GP, Munday PL (2003) Conservation of coral reef biodiversity: a comparison of
- 490 reserve selection procedures for corals and fishes. Biol Conserv 111:53–62
- 491 Bell JD, Steffe AS, Westoby M (1988) Location of seagrass beds in estuaries: effects on associated
- 492 fish and decapods. J Exp Mar Biol Ecol 122:127–146
- 493 Brusca RC, Lindberg DR, Ponder WF (2016) Phylum Mollusca. pp 453–-530 In: Brusca RC, Moore W,
- 494 Shuster SM. Invertebrates. Sinauer Associates, Sunderland
- 495 Clark GF, Kelaher BP, Dafforn KA, Coleman MA, Knott NA, Marzinelli EM, Johnston EL (2015) What
- 496 does impacted look like? High diversity and abundance of epibiota in modified estuaries. Environ
- 497 Pollut 196:12–20
- 498 Clarke KR, Gorley RN (2015) PRIMER v7: User manual/tutorial. PRIMER-E, Plymouth

- 499 Corte GN, Checon HH, Fonseca G, Vieira DC, Gallucci F, Di Domenico M, Amaral ACZ (2017) Cross-
- taxon congruence in benthic communities: Searching for surrogates in marine sediments. Ecol Indic
 78:173–182
- 502 Crawley MJ (2009) The R Book. Second edition. John Wiley & Sons Ltd, Chichester, West Sussex,
- 503 England, United Kingdom
- 504 Crossman S, Li O (2015) Surface hydrology polygons (regional). Geoscience Australia, Canberra
- 505 CSIRO (1993) Jervis Bay Marine Ecological Studies Final Report. CSIRO Division of Fisheries, Jervis Bay
- 506 Duarte CM (1989) Temporal biomass variability and production/biomass relationships of seagrass
- 507 communities. Mar Ecol Progr Series 51:269–276
- 508 Edgar GJ, Barrett NS (2002) Benthic macrofauna in Tasmanian estuaries: scales of distribution and
- relationships with environmental variables. J Exp Mar Biol Ecol 270:1–24
- 510 Eglington SM, Noble DG, Fuller RJ (2012) A meta-analysis of spatial relationships in species richness
- across taxa: Birds as indicators of wider biodiversity in temperate regions. J Nat Conserv 20:301-309
- 512 Fisher R, Knowlton N, Brainard RE, Caley MJ (2011) Differences among Major Taxa in the Extent of
- 513 Ecological Knowledge across Four Major Ecosystems. PLoS ONE 6:e26556
- 514 Gaston KJ, Williams PH (1996) Spatial patterns in taxonomic diversity In: Gaston KJ (Ed.), Biodiversity.
- 515 A Biology of Numbers and Difference, Blackwell Science, Oxford, pp 202–229
- 516 Gladstone W (2002) The potential value of indicator groups in the selection of marine reserves. Biol
- 517 Conserv 104:211–220
- 518 Gladstone W, Owen V (2005) The potential value of surrogates for the selection and design of
- 519 marine reserves for biodiversity and fisheries pp 224-226 In: Day JC, Senior J, Monk S, Neal W (eds)

- 520 First International Marine Protected Areas Congress, 23-27 October 2005, Conference Proceedings
- 521 (IMPAC1, Geelong)
- 522 Grantham HS, Wilson KA, Moilanen A, Rebelo T, Possingham HP (2009) Delaying conservation
- 523 actions for improved knowledge: how long should we wait? Ecol Lett 12:293–301
- 524 Grantham HS, Pressey RL, Wells JA, Beattie AJ (2010) Effectiveness of biodiversity surrogates for
- 525 conservation planning: different measures of effectiveness generate a kaleidoscope of variation.
- 526 PLoS ONE 5, e11430
- 527 Heino J (2010) Are indicator groups and cross-taxon congruence useful for predicting biodiversity in
- 528 aquatic ecosystems? Ecol Indic 10:112-117
- 529 Hirst AJ (2008) Surrogate measures for assessing cryptic faunal biodiversity on macroalgal-
- 530 dominated subtidal reefs. Biol Conserv 141:211–220
- 531 Hughes TP, Bellwood DR, Connolly SR (2002) Biodiversity hotspots, centres of endemicity, and the
- 532 conservation of coral reefs. Ecology Letters 5:775–784
- 533 Hutchings P (1998) Biodiversity and functioning of polychaetes in benthic sediments. Biodivers
- 534 Conserv 7:1133–1145
- 535 Hutchings PA, Jacoby C (1994) Temporal and spatial patterns in the distribution of infaunal
- 536 polychaetes in Jervis Bay, New South Wales. Memoir Natl Hist 441–452
- 537 Ilg C, Oertli B (2017) Effectiveness of amphibians as biodiversity surrogates in pond conservation.
- 538 Conserv Biol 31:437-445
- Irwin S, Pedley SM, Coote L, Dietzsch AC, Wilson MW, Oxbrough A, Sweeney O, Moore KM, Martin R,
- 540 Kelly DL, Mitchell FJ (2014) The value of plantation forests for plant, invertebrate and bird diversity
- and the potential for cross-taxon surrogacy. Biodivers Conserv 23:697-714

542

- 544 Jumars PA, Dorgan KM, Lindsay SM (2015) Diet of worms emended: an update of polychaete feeding
- 545 guilds. Ann Rev Mar Sci 7:497–520
- 546 Karakassis I, Machias A, Pitta P, Papadopoulou KN, Smith CJ, Apostolaki ET, Giannoulaki M,
- 547 Koutsoubas D, Somarakis S (2006) Cross-community congruence of patterns in a marine ecosystem:
- 548 Do the parts reflect the whole?. Mar Ecol Prog Ser 310:47–54
- 549 Leroy B, Gallon R, Feunteun E, Robuchon M, Ysnel F (2017) Cross-taxon congruence in the rarity of
- subtidal rocky marine assemblages: No taxonomic shortcut for conservation monitoring. Ecol Indic
- 551 77:239–249
- Lindberg DR, Ponder WF, Haszprunar G (2004) The Mollusca: relationships and patterns from their
- 553 first half-billion years. pp 252–278 In: Cracraft J, Donoghue MJ. Assembling the Tree of Life. Oxford
- 554 University Press, New York
- 555 Lovell S, Hamer M, Slotow R, Herbert D (2007) Assessment of congruency across invertebrate taxa
- and taxonomic levels to identify potential surrogates. Biol Conserv 139:113–125
- 557 MacFarlane GR, Booth DJ (2001) Estuarine macrobenthic community structure in the Hawkesbury
- 558 River, Australia: Relationships with sediment physicochemical and anthropogenic parameters.
- 559 Environ Monit Assess 72: 51–78
- 560 Magierowski RH, Johnson CR (2006) Robustness of surrogates of biodiversity in marine benthic
- 561 communities. Ecol App 16:2264–2275
- 562 Magurran AE (2003) Measuring biological diversity. Blackwell, Oxford

- 563 Mangiafico SS (2016) Summary and Analysis of Extension Program Evaluation in R, version 1.9.0.
- 564 rcompanion.org/handbook/
- 565 Margules CR, Pressey RL (2000) Systematic conservation planning. Nature 405:243–253
- 566 Marshall JE, Bucher DJ, Smith SD (2018) Patterns of infaunal macromollusc assemblages in a
- 567 subtropical marine park: implications for management. Mar Freshwater Res 69:502–513
- 568 McCune B, Mefford MJ (2018) PC-ORD. Multivariate Analysis of Ecological Data. Version 7.08. Wild
- 569 Blueberry Media, Corvallis, Oregon, USA
- 570 Mellin C, Delean S, Caley J, Edgar G, Meekan M, Pitcher R, Przeslawski R, Williams A, Bradshaw C
- 571 (2011) Effectiveness of biological surrogates for predicting patterns of marine biodiversity: a global
- 572 meta-analysis. PLoS One 6:e20141
- 573 Menegotto A, Rangel TF (2018) Mapping knowledge gaps in marine diversity reveals a latitudinal
- 574 gradient of missing species richness. Nature Comm 9:4713
- 575 Morrisey DJ, Underwood AJ, Howitt L, Stark JS (1992) Temporal variation in soft-sediment benthos. J
- 576 Exp Mar Biol Ecol 164:233–245
- 577 Mueller M, Pander J, Geist J (2013) Taxonomic sufficiency in freshwater ecosystems: effects of
- 578 taxonomic resolution, functional traits, and data transformation. Freshw Sci 32:762–778
- 579 Neeson TM, Van Rijn I, Mandelik Y (2013) How taxonomic diversity, community structure, and
- sample size determine the reliability of higher taxon surrogates. Ecol App 23:1216–1225
- 581 Oberprieler SK, Andersen AN, Gillespie GR, Einoder LD (2019) Vertebrates are poor umbrellas for
- 582 invertebrates: cross-taxon congruence in an Australian tropical savanna. Ecosphere 10:e02755.

- 583 Olsgard F, Somerfield PJ, Carr MR (1997) Relationships between taxonomic resolution and data
- transformations in analyses of a macrobenthic community along an established pollution gradient.
- 585 Mar Ecol Prog Ser 149:173–181
- 586 Olsgard F, Somerfield PJ (2000) Surrogates in marine benthic investigations-which taxonomic unit to
- 587 target? J Aq Eco Stress Rec 7:25–42
- 588 Olsgard F, Brattegard T, Holthe T (2003) Polychaetes as surrogates for marine biodiversity: lower
- 589 taxonomic resolution and indicator groups. Biodivers Conserv 12:1033–1049
- 590 Padial AA, Declerck SA, De Meester LUC, Bonecker CC, Lansac-Tôha FA, Rodrigues LC, Takeda A, Train
- 591 S, Velho LF, Bini LM (2012) Evidence against the use of surrogates for biomonitoring of Neotropical
- 592 floodplains. Freshw Biol 57:2411-2423
- 593 Pinheiro J, Bates D, DebRoy S, Sarkar D, R Core Team (2017) nlme: linear and Nonlinear mixed effects
- 594 models. https://CRAN.R-project.org/package=nlme
- 595 Ponder WF, Lindberg DR (Ed) (2008) Phylogeny and evolution of the mollusca. University of
- 596 California Press, Berkeley
- 597 Przeslawski R, Radke L, Hughes M (2009) Temporal and finescale variation in the biogeochemistry of
- 598 Jervis Bay. Geoscience Australia, Record 2009/12. Geoscience Australia, Canberra
- 599 Quinn GP, Keough MJ (2002) Experimental design and data analysis for biologists. Cambridge
- 600 University Press, Cambridge
- 601 R Core Team (2019) R: a language and environment for statistical computing. R Foundation for
- 602 Statistical Computing, Vienna, Austria
- 603 Radke LC, Huang Z, Przeslawski R, Webster IT, McArthur MA, Anderson TJ, Siwabessy PJ, Brooke BP
- 604 (2011) Including biogeochemical factors and a temporal component in benthic habitat maps:
- 605 influences on infaunal diversity in a temperate embayment. Mar Freshwater Res 62:1432–1448

- 606 Rodrigues AS, Brooks TM (2007) Shortcuts for biodiversity conservation planning: the effectiveness
- 607 of surrogates. Ann Rev Ecol Evol Syst 38:713-737
- 608 Rouse G, Pleijel F (2006) Reproductive biology and phylogeny of Annelida. Science Publishers, Enfield
- 609 Shokri MR, Gladstone W, Kepert A (2009) Annelids, arthropods or molluscs are suitable as surrogate
- 610 taxa for selecting conservation reserves in estuaries. Biodivers Conserv 18:1117–1130
- 611 Smith SD. (2005) Rapid assessment of invertebrate biodiversity on rocky shores: where there's a
- 612 whelk there's a way. Biodivers Conserv 14:3565-3576
- 613 Su JC, Debinski DM, Jakubauskas ME, Kindscher K (2004) Beyond species richness: community
- similarity as a measure of cross-taxon congruence for coarse-filter conservation. Conserv Biol
- 615 18:167-173
- 616 Sutcliffe R, Pitcher CR, Caley MJ, Possingham HP (2012) Biological surrogacy in tropical seabed
- 617 assemblages fails. Ecol App 22:1762–1771
- 618 Sutcliffe PR, Klein C.J, Pitcher CR, Possingham HP (2015) The effectiveness of marine reserve systems
- 619 constructed using different surrogates of biodiversity. Conserv Biol 29:657–667
- 620 Valesini FJ, Wildsmith MD, Tweedley JR (2018) Predicting estuarine faunal assemblages using
- 621 enduring environmental surrogates, with applications in systematic conservation planning. Ocean
- 622 Coast Manage 165:80–98
- 623 van der Wal D, Lambert GI, Ysebaert T, Plancke YM, Herman PM (2017) Hydrodynamic conditioning
- 624 of diversity and functional traits in subtidal estuarine macrozoobenthic communities. Estuar Coast
- 625 Shelf Sci 197:80–92
- 626 Velghe K, Gregory-Eaves I (2013) Body size is a significant predictor of congruency in species richness
- 627 patterns: a meta-analysis of aquatic studies. PLoS ONE 8:e57019

- Vellend M, Lilley PL, Starzomski BM (2008) Using subsets of species in biodiversity surveys. J App Ecol
 45:161-169
- 630 Westgate MJ, Barton PS, Lane PW, Lindenmayer DB (2014) Global meta-analysis reveals low
- 631 consistency of biodiversity congruence relationships. Nature Comm 5:3899
- 632 Westgate MJ, Tulloch AI, Barton PS, Pierson JC, Lindenmayer DB (2017) Optimal taxonomic groups
- 633 for biodiversity assessment: a meta-analytic approach. Ecography 40:539–548
- 634 Xu G, Wang Z, Yang Z, Xu H (2015) Congruency analysis of biofilm-dwelling ciliates as a surrogate of
- 635 eukaryotic microperiphyton for marine bioassessment. Mar Poll Bull 101:600–604
- 636 Yong DL, Barton PS, Ikin K, Evans MJ, Crane M, Okada S, Cunningham SA, Lindenmayer DB (2018)
- 637 Cross-taxonomic surrogates for biodiversity conservation in human-modified landscapes–A multi-
- 638 taxa approach. Biol Conserv 224:336-346
- 639 Ysebaert T, Herman PM (2002) Spatial and temporal variation in benthic macrofauna and
- relationships with environmental variables in an estuarine, intertidal soft-sediment environment.
- 641 Mar Ecol Prog Ser 244:105–124



Fig. 1 The study area in Jervis Bay, Australia, showing relative positions of sites in seagrass and unvegetated sediment habitats (source of polygon: Crossman and Li 2015)



Fig 2 Seagrass: Relationships between surrogates (X-axes) and targets (Y-axes) for univariate measures of biodiversity (s species richness, n total no. individuals, d Margalef's index, j Pielou's evenness index, h Shannon-Wiener diversity index). Points are mean values from each site at each sampling event. Coloured symbols indicate a significant relationship between surrogate and target and grey symbols indicate a non-significant relationship.



Fig 3 Unvegetated sediment: Relationships between surrogates (X-axes) and targets (Y-axes) for univariate measures of biodiversity (s species richness, n total no. individuals, d Margalef's index, j Pielou's evenness index, h Shannon-Wiener diversity index). Points are mean values from each site at each sampling event. Coloured symbols indicate a significant relationship between surrogate and target and grey symbols indicate a non-significant relationship.



Fig 4 Relationships between surrogate (X-axes) and target (Y-axes) for univariate measures of biodiversity (s species richness, n total no. individuals, d Margalef's index, j Pielou's evenness index, h Shannon-Wiener diversity index) in seagrass (vegetated) and unvegetated sediment (unvegetated). Points are mean values from each site (n=4 seagrass sites, n=6 unvegetated sediment sites) at each sampling event (n=9). Black symbols indicate a significant relationship between surrogate and target and grey symbols indicate a non-significant relationship.



Fig. 5 The effect of data transformation (square root, $\bigcirc \log (X+1)$, presence-absence) on the magnitude of the Mantel correlation coefficient (R_M) between the Bray-Curtis dissimilarity matrices of surrogates and their targets. Values shown are mean R_M -values (± standard error) for (A) seagrass for 4 sampling events (n=12), (B) seagrass for 9 sampling events (n=9), (C) unvegetated sediment for 4 sampling events (n=12), and (D) unvegetated sediment for 9 sampling events (n=9). Results of one-way ANOVA testing for significant differences among mean R_M -values are also shown (Levene's tests done prior to ANOVAs were all non-significant)

Event	Surrogat	e: C	Targ	get: M+P		Surroga	ate: M		Targe	et: C+P	Su	irrogate: P	Та	rget: C+	Μ
1	4	2D Stre 3	3	2D Stre	3			2D Stre	4	1 ^{2D Stre}	4	2D Stra 1 2	4	1	2D Stre 2
	1	2	4	1-	21			4	3	2	3	2		3	
2	4	2D Stre	3	2D Stre	3			2D Stre	1	2D Stre 4	1	2 2D Stre		2	2D Stre
	3	2	2	4 2		1	4		2	3	3	4	1	4	3
3	4	2D Stre 1	3	2D Stre 4			1	2D Stre	1	2D Stre	3	2D Stre	4		2D Stre
	2	3		2	3	:	2	4	2 4	3	42	1	21		3
4		3 2D Stré	1	2D Stre			1	2D Stre	3	2D Stre	1	2D Stre 4			2D Stre 2
	41			4	3			4		4	2		4	1	з
		2	3	2		2			2		5	2			5

Fig 6. Seagrass: nMDS ordination plots (based on Bray-Curtis dissimilarity matrices of square-root transformed abundances) comparing dissimilarities among sites (numbered 1-4) for assemblages of surrogates and their targets in each of four sampling events. C Crustaceans, M Molluscs, P Polychaetes.

	Mol	luscs						Pol	ych	aete	es		
				2D Stre		4					1		2D Stre
3											-		
-1				4									2
21													
				2D Stre			3						2D Stre
3						1			2				
				2		-							_
		4											4
	1					3	3						
				2D Stre									2D Stre
			1			3							
3						4	2						1
		2		4		•	2						
				2D Stre									2D Stre
		1					1						
3													4
				4		_							
	2					3					2		
3				2D Stre									2D Stre
	1							3		1			
				2				•					
						4							2
4				25.61									
	1			2D Stre		4							1
2	2		ſ									_	
	•		۷									2	
	4							3	8				
	3 21 3 3 3 3 4	3 21 3 1 3 3 2 3 1 3 1 4 1 1 4 1 1 4 1		$ \begin{array}{ccccccccccccccccccccccccccccccccc$	MONUUSCS 2D Stress 3 4 21 2D Stress 3 2 4 1 3 2 4 2 3 2 4 2	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	NOTUBES 2D Strid 4 3 4 1 21 4 1 3 2 4 3 3 2 3 3 3 2 4 3 4 1 3 3 3 2 4 3 3 2 4 3 3 2 4 3 3 2 5 1 3 2 4 3 3 2 5 4 2 3 2 5 4 3 3 2 5 4 3 3 2 5 4 3 4 2 5 4 4 4 3 2 5 5 4 4 3 2 5 4 5 5 4 3 2 5 5 5 5 3 2 5 5 5 5	NONIDECS 2D Strip 4 3 4 4 21 4 3 21 4 3 3 2D Strip 3 2 2D Strip 1 3 2D Strip 3 4 1 3 3 2 4 1 2D Strip 3 3 2 4 3 2 4 3 2 4 3 2 4 3 2D Strip 1 3 2 4 3 2D Strip 1 3 2D Strip 3 4 3 3 2 3 3 3 2D Strip 4 3 2D Strip 4 3 2D Strip 4 3 2 4 3 2 4 3 2 4	NOTICISCS 2D Strict 4 3 4 4 21 4 3 21 4 3 3 2D Strict 2 4 1 3 2 4 3 3 2 4 1 2D Strict 3 3 2 4 2 1 2D Strict 1 3 3 2 4 2 3 2 4 3 3 2 4 3 3 2 5 1 3 2 5 1 3 2 4 3 1 2 3 3 4 3 2 5 4 3 3 3 4 3 4 3 4 3 4 4 3 2 4 4 3 2 4 3 4 3 3 <th>Nonusci 20 stri 4 3 4 4 21 4 3 21 4 3 3 20 stri 3 4 1 3 3 2 4 1 20 stri 3 3 2 4 2 1 20 stri 1 1 3 2 4 2 3 2 4 3 1 3 2 3 3 1 3 2 3 3 1 3 2 3 3 1 3 2 2 3 3 1 3 2 2 3 3 1 4 2 3 1 3 1 4 1 2 4 1 1 3 2 1 3 1 1 4 1 2 1 1 3 1 4<!--</th--><th>Nonluscs 2D Stric 4 1 3 4 1 1 1 21 4 1 1 1 1 3 20 Stric 1 1 1 1 1 3 20 Stric 1 3 1</th><th>MONUUSCS 20 Stric 4 1 3 4 1 1 21 4 3 3 21 4 1 3 3 20 Stric 1 2 3 20 Stric 1 3 4 1 3 3 3 3 2 4 3 3 3 2 4 3 3 3 2 4 3 2 3 2 4 3 2 3 2 4 3 2 3 2 2 3 2 3 3 2 2 3 2 3 4 1 2 3 2 3 4 2 3 2 4 3 4 1 2 3 2 4 3 2 4 2 2 2 4 2 2 3 3 2</th></th>	Nonusci 20 stri 4 3 4 4 21 4 3 21 4 3 3 20 stri 3 4 1 3 3 2 4 1 20 stri 3 3 2 4 2 1 20 stri 1 1 3 2 4 2 3 2 4 3 1 3 2 3 3 1 3 2 3 3 1 3 2 3 3 1 3 2 2 3 3 1 3 2 2 3 3 1 4 2 3 1 3 1 4 1 2 4 1 1 3 2 1 3 1 1 4 1 2 1 1 3 1 4 </th <th>Nonluscs 2D Stric 4 1 3 4 1 1 1 21 4 1 1 1 1 3 20 Stric 1 1 1 1 1 3 20 Stric 1 3 1</th> <th>MONUUSCS 20 Stric 4 1 3 4 1 1 21 4 3 3 21 4 1 3 3 20 Stric 1 2 3 20 Stric 1 3 4 1 3 3 3 3 2 4 3 3 3 2 4 3 3 3 2 4 3 2 3 2 4 3 2 3 2 4 3 2 3 2 2 3 2 3 3 2 2 3 2 3 4 1 2 3 2 3 4 2 3 2 4 3 4 1 2 3 2 4 3 2 4 2 2 2 4 2 2 3 3 2</th>	Nonluscs 2D Stric 4 1 3 4 1 1 1 21 4 1 1 1 1 3 20 Stric 1 1 1 1 1 3 20 Stric 1 3 1	MONUUSCS 20 Stric 4 1 3 4 1 1 21 4 3 3 21 4 1 3 3 20 Stric 1 2 3 20 Stric 1 3 4 1 3 3 3 3 2 4 3 3 3 2 4 3 3 3 2 4 3 2 3 2 4 3 2 3 2 4 3 2 3 2 2 3 2 3 3 2 2 3 2 3 4 1 2 3 2 3 4 2 3 2 4 3 4 1 2 3 2 4 3 2 4 2 2 2 4 2 2 3 3 2

Event		Molluscs			Polychaetes	
7		2	2D Stre		2	2D Stre
	1	3	4	4		1
8	1		2D Stre 2	1	3	2D Stre 2
			3		4	
9	4	3 1	2D Stre 2	4		2D Stre 2 31

Fig 7. Seagrass: nMDS ordination plots (based on Bray-Curtis dissimilarity matrices of square-root transformed abundances) comparing dissimilarities among sites (numbered 1-4) for assemblages of molluscs and polychaetes in each of nine sampling events.

Event	Surrogate: C	Target: M+P	Surrogate: M	Target: C+P	Surrogate: P	Target: C+M
1	^{2D Stre} 2 4 6 5 3	20 Strd 4 1 3 2 6 5	1 20 Stre	\$ 20 Stress 5 1 2	4 ^{2D Stra} 3 5 2 6	5 2D Stress 6 3 4 1 2
2	6 ^{2D Stress} 5 2 4 1 3	5 4 ^{2D Stress} 6 1 3	1 20 Stre 3 5	4 20 Strest 5 1 6 3 2	4 2D Stress 62 3	5 20 Stre 4 2 1 3
3	20 Stress 3 5 2 6	5 20 Stres 62 143	6 ^{2D Stress}	3 20 Stress 4 1 5 6 2	5 20 Stress 3 4 2 3 1	4 ^{2D Stress} 4 ^{2D Stress} 5 2
4	3 20 Stress 1 4 5 2 6	3 20 Stree 4 1 5 2 6	3 20 Stress 1 4 6 2 5	3 4 5 1 2 6	2 ⁶ 2D Stress 1 4 3 5	1 2 2D Stress 1 6 3 4 5

Fig 8. Unvegetated sediment: nMDS ordination plots (based on Bray-Curtis dissimilarity matrices of square-root transformed abundances) comparing dissimilarities among sites (numbered 1-6) for assemblages of surrogates and their targets in each of four sampling events. C Crustaceans, M Molluscs, P Polychaetes



Event		Molluscs		F	Polychaetes	
7	1		2D Stress		6	2D Stress
	3	4	5	2	1 3	5
		6				
	2			4		
8			6 ^{2D Stree}			2D Stre
		5			4	5
		2 4		2	4	
					•	6
		3				
9			2D Stre	5	; c	2D Stress
	1				O	
	3	6 5	2	4		2
	4			8	k	

Fig 9. Unvegetated sediment: nMDS ordination plots (based on Bray-Curtis dissimilarity matrices of square-root transformed abundances) comparing dissimilarities among sites (numbered 1-6) for assemblages of molluscs and polychaetes in each of nine sampling events

Table 1 Seagrass: Results of generalized least squares analyses to fit linear models betweensurrogates and targets for each biodiversity variable across four and nine sampling events.Biodiversity variables tested were species richness (s), total number of individuals (n), d Margalef'sindex, j Pielou's evenness index, and h Shannon-Wiener diversity index

Surrogate	Target	Response variable	χ²	Df	Р	_p r ²	_p r
Crustaceans	Molluscs + Polychaetes	S	9.04	1	0.003	0.4	0.63
		n	51.56	1	<0.0001	0.8	0.89
		d	2.09	1	0.10	0.07	0.26
		j	24.52	1	<0.0001	0.36	0.60
		h	4.12	1	0.04	0.17	0.41
Molluscs	Crustaceans + Polychaetes	S	25.04	1	<0.0001	0.62	0.79
		n	3.54	1	0.06	0.21	0.46
		d	11.52	1	0.0007	0.37	0.61
		j	1.75	1	0.20	0.14	0.37
		h	53.33	1	< 0.0001	0.65	0.81
Polychaetes	Crustaceans + Molluscs	S	17.51	1	<0.0001	0.54	0.73
		n	5.21	1	0.02	0.3	0.55
		d	9.23	1	0.002	0.26	0.51
		j	0.66	1	0.40	0.08	0.28
		h	34.23	1	<0.0001	0.58	0.76

(a) Four sampling events

Surrogate	Target	Response variable	χ²	Df	Р	_p r ²	_p r
Molluscs	Polychaetes	S	11.50	1	0.0007	0.36	0.60
		n	2.06	1	0.20	0.09	0.30
		d	7.22	1	0.007	0.22	0.47
		j	4.01	1	0.045	0.08	0.28
		h	5.09	1	0.02	0.27	0.52

Table 2 Unvegetated sediment: Results of generalized least squares analyses to fit linear models between surrogates and targets for each biodiversity variable across four and nine sampling events. Biodiversity variables tested were species richness (s), total number of individuals (n), d Margalef's index, j Pielou's evenness index, and h Shannon-Wiener diversity index

Surrogate	Target	Response variable	χ^2	Df	Р	_p r ²	_p r
Crustaceans	Molluscs + Polychaetes	S	28.43	1	<0.0001	0.57	0.75
		n	33.80	1	<0.0001	0.57	0.75
		d	7.78	1	0.005	0.29	0.54
		j	1.33	1	0.20	0.05	0.22
		h	0.61	1	0.40	0.03	0.17
Molluscs	Crustaceans + Polychaetes	S	66.58	1	<0.0001	0.79	0.89
		n	44.83	1	<0.0001	0.68	0.82
		d	15.89	1	<0.0001	0.47	0.69
		j	1.05	1	0.30	0.05	0.22
		h	0.37	1	0.50	0	0
Polychaetes	Crustaceans + Molluscs	S	21.27	1	<0.0001	0.53	0.73
		n	13.53	1	0.0002	0.39	0.62
		d	6.94	1	0.008	0.28	0.53
		j	4.43	1	0.04	0.16	0.40
		h	0.72	1	0.40	0.05	0.22

(a) Four sampling events

Surrogate	Target	Response variable	χ^2	Df	Р	_p r ²	_p r
Molluscs	Polychaetes	S	34.45	1	<0.0001	0.37	0.61
		n	12.67	1	0.0004	0.21	0.46
		d	8.96	1	0.003	0.14	0.37
		j	1.38	1	0.20	0.01	0.10
		h	2.81	1	0.09	0.05	0.22

Table 3 Seagrass: Mantel (R_M) correlation coefficients between dissimilarity matrices of surrogates and targets. Partial Mantel correlation coefficients (pR_M) controlling for the effects of physical distances between sites were calculated for surrogates with $R_M \ge 0.7$. Tests were done on Bray-Curtis dissimilarity matrices with abundance data transformed to square-root

(a)	Four	samp	ling	events
-----	------	------	------	--------

Event	Surrogate	Target	R _M	р <i>R</i> м
1	Crustaceans	Molluscs + Polychaetes	0.49	
	Molluscs	Crustaceans + Polychaetes	0.57	
	Polychaetes	Crustaceans + Molluscs	0.83	0.71
2	Crustaceans	Molluscs + Polychaetes	-0.68	
	Molluscs	Crustaceans + Polychaetes	-0.73	
	Polychaetes	Crustaceans + Molluscs	-0.97	
3	Crustaceans	Molluscs + Polychaetes	-0.42	
	Molluscs	Crustaceans + Polychaetes	0.26	
	Polychaetes	Crustaceans + Molluscs	-0.49	
4	Crustaceans	Molluscs + Polychaetes	0.25	
	Molluscs	Crustaceans + Polychaetes	0.69	
	Polychaetes	Crustaceans + Molluscs	-0.17	

Event	Surrogate	Target	R _M	р <i>R</i> м
1	Molluscs	Polychaetes	0.66	
2	Molluscs	Polychaetes	-0.82	
3	Molluscs	Polychaetes	-0.29	
4	Molluscs	Polychaetes	0.08	
5	Molluscs	Polychaetes	0.57	
6	Molluscs	Polychaetes	0.18	
7	Molluscs	Polychaetes	0.05	
8	Molluscs	Polychaetes	0.17	
9	Molluscs	Polychaetes	0.71	0.71

Table 4 Unvegetated sediment: Mantel (R_M) correlation coefficients between dissimilarity matrices of surrogates and targets. Partial Mantel correlation coefficients (pR_M) controlling for the effects of physical distances between sites were calculated for surrogates with $R_M \ge 0.7$. Tests were done on Bray-Curtis dissimilarity matrices with abundance data transformed to square-root

Event	Surrogate	Target	R _M	р <i>R</i> м
1	Crustaceans	Molluscs + Polychaetes	0.51	
	Molluscs	Crustaceans + Polychaetes	0.78	0.79
	Polychaetes	Crustaceans + Molluscs	0.56	
2	Crustaceans	Molluscs + Polychaetes	0.59	
	Molluscs	Crustaceans + Polychaetes	0.59	
	Polychaetes	Crustaceans + Molluscs	0.66	
3	Crustaceans	Molluscs + Polychaetes	0.54	
	Molluscs	Crustaceans + Polychaetes	0.31	
	Polychaetes	Crustaceans + Molluscs	0.54	
4	Crustaceans	Molluscs + Polychaetes	0.68	
	Molluscs	Crustaceans + Polychaetes	0.62	
	Polychaetes	Crustaceans + Molluscs	0.63	

(a) Four sampling events

Event	Surrogate	Target	R _M	р <i>R</i> м
1	Molluscs	Polychaetes	0.69	
2	Molluscs	Polychaetes	0.56	
3	Molluscs	Polychaetes	0.23	
4	Molluscs	Polychaetes	0.52	
5	Molluscs	Polychaetes	-0.03	
6	Molluscs	Polychaetes	0.1	
7	Molluscs	Polychaetes	-0.5	
8	Molluscs	Polychaetes	0.41	
9	Molluscs	Polychaetes	0.84	0.88