

# **Strategies of Resource Allocation by Diatoms under Dynamic Light**

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PhD by Research

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the degree of Doctor of Philosophy

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## Certificate of original authorship

I, Nerissa Lynn Fisher, declare that this thesis is submitted in fulfillment of the requirements for the award of Doctorate of Philosophy in the Faculty of Science at the University of Technology Sydney.

This thesis is wholly my own work unless otherwise referenced or acknowledged.

In addition, I certify that all information sources and literature used are indicated in the thesis.

This document has not been submitted for qualifications at any other academic institution.

This thesis is supported by the Australian Government Research Training Program.

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Date: 20<sup>th</sup> of May 2021

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### Chapter 1

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## Chapter 2

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### Chapter 3

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## Chapter 4

**Figure 4.1.** (A) PCA of relative metabolite abundances for constant light treatments (24:0 L:D) at high (yellow circles), medium (green circles) and low (blue circles) intensities. (B) PLS-DA of relative metabolite abundances for high and low constant (high and low) and pulse (high – orange triangles, low – purple triangles) light treatments. Light intensities for high, medium and low are 200, 60 and 5 μmol photons

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## Chapter 5

**Figure 5.1.** Schematic of diatom responses to high light from species found in open ocean (*T. oceanica*) and estuary (*T. weissflogii*) in which coastal diatoms exhibit an intermediate response. The main flow of energy involves light energy harvested at photosystem II (PSII) where excitons are passed along the linear electron transport chain to PSI where NADPH is generated to fuel, in addition to ATP, the Calvin-Benson-Bassham (CBB) cycle. Products generated from the CBB cycle enter the cytoplasm (yellow box) to glycolysis which feeds into the tricarboxylic acid cycle (TCA) cycle in the mitochondria (red box) to build macromolecules to support growth. Alternatively, gluconeogenesis diverts energy away from TCA cycle to build carbon reserves. Sources of energy dissipation from high light before reaching PSII are the yield of constitutive losses via fluorescence and heat (YNO) and the yield of regulated thermal dissipation via nonphotochemical quenching (YNPQ) Once photolysis occurs at the PSII reaction centre, electrons can enter processes of light-dependent respiration (LDR) via oxidase activity within the chloroplast (green box) which can be a way to dissipate excess electrons or generate additional ATP. Additionally, mitochondrial respiration can supplement ATP demands via the catabolism of carbon molecules to supply to the CBB cycle or assimilation of nutrients (i.e. nitrogen, N). Nitrogen is particularly essential to build pigments and proteins that are fundamental components of the nonphotochemical quenching mechanism. The arrow thickness correlates to the

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**Figure 5.2.** Development of energy budget models using cellular currencies – carbon (green background), oxygen (blue background) and fluorescence (yellow background) – from (A) historical, (B) current to (C) proposed future models. Historical energy budget models typically include two cellular currencies and a separate biofractionation of macromolecules. Current energy budgets account for the three cellular currencies but does not include that deeper carbon insight gained from metabolomics. Future energy budgets models could integrate all cellular currencies including the information gained from further partitioning of carbon molecules (e.g. metabolites). Such comprehensive energy budgets will provide more accurate accounting of energy that ultimately is retained in biomass under various environmental stressors. Data adapted from (A) Fisher & Halsey (2016) and (B) Chapters 2 and 3 where sub-fractionations within cellular currencies are measurements that were collected from diatoms in response to light.

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### Chapter 2

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### Chapter 3

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## Chapter 2

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**S3.3 Table.** Rates of oxygen production and consumption. MIMS analysis of gross oxygen production ( $GP_{O_2}$ ), light dependent respiration (LDR), dark respiration ( $R_{\text{DARK}}$ ), and net oxygen production ( $Net_{O_2}$ ) for *T. weissflogii* and *T. oceanica* grown under a sinusoidal light regime with a maximum of 400  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  ( $Sine_{\mu}$ ) over a 12:12 L:D cycle. Rates were also collected from a transient exposure to high light that was 3x  $Sine_{\mu}$  ( $Sine_{HL}$ ) at the sample time. Samples were collected during the photoperiod at 1, 3, 6, 9 and 11 hours. Data averaged from 3 independent replicates. Values in parentheses are SE of the mean. For direct comparison with oxygen consumption/production rates from Chapter 2 see S2.1 Table.

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**S4.1 Table.** Cell densities ( $\text{mL}^{-1}$ ), volume of culture (mL) concentrated and preserved for metabolite extraction and the total cells ( $\text{mL}^{-1}$ ) extracted for metabolomics analysis of technical replicates for three constant (high, medium, low) and two pulse (high, low) light treatments.

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## List of Abbreviations

AA	Amino acids
ADP	Adenosine diphosphate
ANOVA	Analysis of variance
AOX	Mitochondrial alternative oxidase
Ar	Argon
ATP	Adenosine triphosphate
BCAA	Branched chain amino acid
$\beta$ -Car	Beta carotene
C	Carbon
CBB cycle	Calvin-Bassham-Benson cycle
CCM	Carbon concentrating mechanism
CEF	Cyclic electron flow
Chl	Chlorophyll
CI	Confidence intervals
CO <sub>2</sub>	Carbon dioxide
Cyt <i>b<sub>6</sub>f</i>	Cytochrome <i>b<sub>6</sub>f</i>
DCMU	3-(3,4-dichlorophenyl)-1,1-dimethylurea
Dd	Diadinoxanthin
DHA	Docosahexaenoic acid
DOC	Dissolved organic carbon
DPS	De-epoxidation state
Dt	Diatoxanthin
E	Irradiance
EGT	Endosymbiotic gene transfer
ETC	Electron transport chain
ETR/ETR <sub>PSII</sub>	Electron transport rate from PSII
FC	Fold change
Fd	Ferredoxin
FDP	Flavodiiron protein
FDR	False discovery rate
FLC	Fluorescence light response curve
FNR	Fd-NADP <sup>+</sup> oxidoreductase
FRRf	Fast repetition rate fluorometry
Fuc	Fucoxanthin
GAP	Glyceraldehyde phosphate
GC-MS	Gas chromatography mass spectrometry
GDC	Glycine decarboxylase p-protein
GF/F	Glass fiber filter
GP <sub>C</sub>	Gross carbon production
GP <sub>O<sub>2</sub></sub>	Gross oxygen production
GPP	Gross primary production
H <sup>+</sup>	proton
HC	High constant
HgCl <sub>2</sub>	Mercuric chloride
HL	High light
HP	High pulse
HPLC	High-performance liquid chromatography

Ig	Growth irradiance
KCN	Potassium cyanide
KEGG	Kyoto encyclopedia of genes and genomes
KW	Kurskal-Wallis
L:D	Light:Dark
LC	Low constant
LC-MS	Liquid chromatography mass spectrometry
LDR	Light dependent respiration
LED	Light emitting diode
LEDR	Light enhanced dark respiration
LEF	Linear electron flow
LHC	Light harvesting complex
LHCX	Light harvesting complex proteins
LIFT	Light induced fluorescence transient
LL	Low light
LP	Low pulse
MAP	Mehler ascorbate peroxidase
MC	Medium constant
MeOH	Methanol
MIMS	Membrane inlet mass spectrophotometry
MOX	Midstream oxidase
MT	Multiple turnover
N/N <sub>2</sub>	Nitrogen/nitrogen gas
NADPH	Nicotinamide adenine dinucleotide phosphate
NetO <sub>2</sub>	Net oxygen production
NP <sub>C</sub>	Net carbon production
NPP	Net primary production
NPQ	Nonphotochemical quenching
NPQ <sub>NSV</sub>	Stern-Volmer nonphotochemical quenching
NR	Nitrate reductase
O <sub>2</sub>	Oxygen
OAA	Oxaloacetate
OEC	Oxygen evolving complex
PAM	Pulse amplitude modulated
PAR	Photosynthetically active radiation
PC	Plastocyanin
PC	Pyruvate carboxylase
PCA	Principle component analysis
PE curve	Photosynthesis-irradiance curve
PEP	Phosphoenolpyruvate
PEPCK/PCK1	Phosphoenolpyruvate carboxykinase
PETC	Photosynthetic electron transport chain
Pex	Peroxin
PFD	Photon flux density
PGA	Phosphoglyceric acid
PGR5	Proton gradient regulation protein
PLS-DA	Partial least squares discriminant analysis
PK	Pyruvate kinase

POC	Particulate organic carbon
PON	Particulate organic nitrogen
PP	Primary productivity
PPDK	Pyruvate-phosphate dikinase
PQ	Plastoquinone
PSI/II	Photosystem I/II
PTOX	Plastid/plastoquinol terminal oxidase
PTS	Peroxisomal targeting signals
QMS	Quadrupole mass spectrometer
R <sub>DARK</sub>	'Dark' (mitochondrial) respiration
R <sub>TOTAL</sub>	Total respiration
RC (I/II)	Reaction centre (of PSI/PSII)
RNA	Ribonucleic acid
RuBisCO	Ribulose-1,5-bisphosphate carboxylase
S	Sulfur
SAM	Significance analysis of microarrays
SCF	Spectral correction factor
SE	Standard error
SHAM	Salicylhydroxamic acid
SHMT	Serine hydroxymethyltransferase
Sine <sub>HL</sub>	Sinusoidal high light
Sine <sub>μ</sub>	Sinusoidal growth irradiance
ST	Single turnover
TCA cycle	Tricarboxylic acid cycle
XC	Xanthophyll cycle
YNPQ	Yield of regulated nonphotochemical quenching
YNO	Yield of constitutive losses via fluorescence and heat
YII	Yield of photochemical conversion at PSII

## List of Symbols

$\mu$	Specific growth rate ( $d^{-1}$ )
[1-C]	Photochemical quenching
[1-Q]	Nonphotochemical quenching
$\alpha$	Alpha; light limiting slope
$\sigma_{PSII}$	Functional absorption cross section of PSII
$e^-$	Electron
$\phi$	Quantum yield
$E_k$	Light saturation index
$E_{k,YII}$	Light saturation index of photochemical conversion at PSII
$F'$	Steady-state fluorescence at any point
$F_o$	Minimum fluorescence from dark acclimated sample
$F_m (F_m')$	Maximum fluorescence from dark acclimated sample
$F_v (F_v')$	Variable fluorescence from dark acclimated sample
$F_v/F_m$	Maximum photochemical efficiency
$F_o'$	Minimum fluorescence under actinic light
$F_m'$	Maximum fluorescence under actinic light
$F_v'$	Variable fluorescence under actinic light
$F_v'/F_m'$	Maximum PSII efficiency under actinic light
$k_{PI}$	Photo-inactivation rate constant
$k_{REC}$	Recovery rate constant
$P_{max}$	Photosynthetic maximum



## Thesis Abstract

Diatoms are the evolutionarily youngest phytoplankton group and considered to be the most productive across diverse ocean, coastal and freshwater environments. Based on their evolutionary history in diverse environments, diatoms have acquired unique diverse mechanisms to cope with fluctuating availability of resources required for cellular maintenance and growth. Yet how these mechanisms actually operate to moderate metabolic functioning by the energetic tracking of light energy to carbon capture – commonly measured as “emergent signatures” or photosynthesis rates via fluorescence, O<sub>2</sub> evolution and/or CO<sub>2</sub> uptake – remains somewhat of a black box.

This thesis addresses the response of diatoms to light, with particular emphasis on the gaps in current energy budgets that quantify trade-offs in O<sub>2</sub> evolution and carbon-assimilation. An initial assessment of a variety of diatom species revealed distinct categories of photo-protective capacities (i.e. nonphotochemical quenching) that correlated with ecological niche, i.e. taxa originating from estuarine, coastal and open ocean environments. Low capacity to dissipate light energy via nonphotochemical quenching by open ocean diatoms was compensated for by an upregulation of midstream oxidase activity highlighting a key trade-off between light harvesting and light utilization strategies. Diurnal monitoring of diatoms with divergent photo-protective capacities further revealed species-specific dynamic respiratory trends, whereby diatoms with high nonphotochemical quenching capacity exhibited more dynamic R<sub>DARK</sub> while diatoms with low nonphotochemical quenching capacity exhibited more dynamic light-dependent respiration (LDR). Fluorescence-derived measures of photoacclimation ( $E_{k,YII}$ ) were found to be significantly correlated to oxygen cycling and carbon retained as biomass. Subsequent metabolomic profiling provided deeper insight into these processes via the underlying light-driven metabolite

reorganisation. Using the model coastal diatom (*T. pseudonana*), high light metabolic profiles were reflective of pathways that support higher growth rate (e.g. glycolysis and TCA cycle) compared to low light metabolic profiles associated with carbon conserving pathways (e.g. gluconeogenesis and glyoxylate cycle).

Together these outcomes uncovered previously hidden dynamics of energy processing by diatoms – including dynamic respiration rates between taxa and with time of day, which also mapped differences in inherent metabolic pathways as well as “emergent” metabolic signatures (e.g. fluorescence, O<sub>2</sub> and CO<sub>2</sub> measures of primary productivity). Combining information from cellular currencies (fluorescence, oxygen and carbon) thus provides a more robust mechanistic understanding of metabolic processes. This thesis has created a foundation for future research to compile more comprehensive energy budgets and a framework for improved estimates of primary productivity models.

## Thesis Structure and Declaration of Contribution

This thesis is comprised of three data chapters (Chapters 2 - 4) with each constructed around an independent experiment. All data chapters have been written in the form of a journal manuscript for peer-review. At the time of thesis submission, Chapter 2 has been published and Chapters 3 and 4 are in final draft for submission and ready for submission/peer review pending availability of funds. Each data chapter introduction is exhaustive thus, to avoid information redundancy, the thesis general introduction (Chapter 1) has focussed on topics not covered in as much depth within data chapter introductions to better develop the outlined aims of the thesis.

**Chapter 2:** Fisher N.L., Campbell D.A., Hughes D.J., U., Halsey, K.H., Ralph, P.J. and Suggett D.J. 2020. Divergence of energy strategies amongst diatoms. Published in *PLOS ONE*.

NLF, KHH, PJR, DJS designed the experiment. NLF conducted the experiment. NLF collected all samples and processed data for FRRf and MIMS measurements. UK performed HPLC data analysis. DAC created R script to process photo-inactivation and repair rates and provided figures. NLF wrote manuscript first draft. DAC, DJH, KHH, PJR, DJS provided substantial critical contributions and edits.

**Chapter 3:** Fisher N.L., Halsey K.H., Hughes D.J., Argyle P., Ralph P.J. and Suggett D.J. Contrasting dynamics of light-dependent and -independent respiration from two

*Thalassiosira* diatoms under diurnal light. Thesis chapter. Intended Journal, *Journal of Phycology*

NLF conceptualised and designed the experiment. KHH, DJH, DJS, NLF finalised the experimental design. NLF conducted the experiment. NLF collected all samples and processed data. NLF wrote manuscript first draft. PA assisted with statistical analyses. KHH, DJH, PJR, DJS provided substantial critical contributions and edits.

**Chapter 4:** Fisher N.L., Halsey K.H., Suggett D.J., Pombrol M., Ralph P.J., Lutz A., Sogin E.M. and Matthews J.L. Light-dependent metabolic phenotype of the model diatom *Thalassiosira pseudonana*. Submitted for peer review to *Journal of Experimental Botany*

NLF, KHH, PJR, DJS conceptualised and designed the experiment. NLF conducted the experiment. NLF collected and extracted all samples. AL provided protocols for metabolite extraction and initial processing of metabolomics samples. EMS processed metabolomics samples and resources associated with analysis. JLM identified metabolites from processed data and provided guidance for analyzing metabolomics data. NLF analysed data and created metabolic pathways. MP and KHH provided transcriptomics data. NLF wrote manuscript first draft. KHH made substantial contributions to the written manuscript. JLM, KHH, PJR, DJS, AL, EMS provided critical contributions and edits.