

1 **Biodegradation of plastics from waste electrical and electronic**
2 **equipment by greater wax moth larvae (*Galleria mellonella*),**

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12 **ABSTRACT**

13 Biodegradation of insect larva is a potential method to treat waste electrical and
14 electronic equipment (WEEE) plastics, but the information about feeding selectivity of
15 insect larva to WEEE plastics is lacking. Two kinds of fresh and waste plastics were
16 fed to *Galleria mellonella* (*G. mellonella*) larvae including rigid polyurethane (RPU, a
17 thermosetting plastics), polystyrene (PS, a thermoplastic), waste RPU (WRPU) and
18 waste polystyrene (WPS). The larvae both preferred significantly fresh plastics to waste
19 ones. The possible reason was that WEEE plastics contain more metals and chemical
20 additives than fresh plastics. WRPU, WPS plastics and the corresponding frass were
21 analyzed to explore degradation mechanisms through scanning electron microscope
22 (SEM) and Fourier transform infrared (FT-IR). The results of SEM and FT-IR revealed
23 the destruction of physical structure integrities and the changes in functional groups in
24 plastics, implying the occurrence of biodegradation. A typical hard block engineering
25 plastic, waste high impact polystyrene (WHIPS), was fed to *G. mellonella* larvae before
26 and after grounded into powders, and the larvae showed great apparent modification
27 selectivity. In addition, six common fresh foamed plastics were fed to the larvae using
28 different combinations, showing obvious feeding selectivity and preference. It possibly
29 attributed to different physical properties and chemical structures of plastics. The
30 consumption of WRPU plastics by the *G. mellonella* larvae was higher than that by the
31 *Tenebrio molitor* (*T. molitor*) larvae, implying that different insect larvae have different
32 ingesting capability to plastics. The study provides new insights into the selectivity of
33 insect larva for biodegrading WEEE plastics.

34

35 **Keywords:** Waste Electrical and Electronic Equipment Plastics; Insect Larva;

36 Biodegradation; Feeding selectivity; *G. mellonella*

37 **Introduction**

38 Waste electrical and electronic equipment (WEEE) is a mixture of materials and
39 components of scrapped electrical and electronic appliances, typically for example,
40 obsolete computers and cell phones (Li et al., 2019). According to the statistics, about
41 30 million computers are disposed in USA merely and 10 million cell phones are
42 discarded in Europe on a yearly basis. On a worldwide scale, an estimation of 50 million
43 tons of WEEE is produced annually (Luhar et al., 2019). Presently, research about
44 WEEE mainly focuses on its metal recycling. However, plastics materials lack due
45 attention. They constitute approximately 17% of WEEE stream, mainly including
46 acrylonitrile-butadiene-styrene (ABS), high impact polystyrene (HIPS), polyurethane
47 (PU), phenolic resin (PF), polyethylene (PE), polystyrene (PS), polypropylene (PP) and
48 polyvinyl chloride (PVC) plastics (Yang et al., 2013). Taking typical application
49 examples, HIPS plastics usually serve as casing, housing, and PU plastics can serve as
50 insulation. Final disposal of WEEE plastics is reaching similar levels to municipal solid
51 waste plastics (Wu et al., 2020, Shen et al., 2018). A lack of proper management, WEEE
52 plastics cause environmental and health problems. Some plastics related chemicals
53 occur in the food chain, then enter the human body. Hazardous elements in WEEE
54 plastics, including heavy metals and chemical additives, such as lead (Pb), mercury (Hg)
55 and flame (or fire) retardants, will have a negative impact on human health. They may
56 damage physiological system, cause cancer and adverse hormonal effects (Petridis et
57 al., 2017, Charitopoulou et al., 2020).

58 As byproducts of the booming technology, huge volume of environmentally

59 hazardous WEEE plastics produced globally. How to dispose them has become an
60 important and urgent issue (Akram et al., 2019). Compared to common methods of
61 processing WEEE plastics (landfill, incineration, physical recycling and chemical
62 recovery), biodegradation is a more noteworthy way to fight against plastic pollution
63 for its low costs and environmentally friendly impacts (Azubuike et al., 2016). There
64 are many studies on the biodegradation of plastics by microorganisms, including
65 bacteria and fungi (Magnin et al., 2019). Microorganisms utilize plastics as carbon
66 and/or nitrogen sources (Sheth et al., 2019). A fungus, *Aspergillus fumigatus* Strain S45,
67 inoculated on specific tween 20 agar plate at 30 °C, was able to reduce weight of PU
68 films which exposure to UV radiation as pre-processing by 15%-20% (about 37.5 - 50
69 mg) in 28 days (Osman et al., 2018). A *Paenibacillus* sp. can also degrade PE after
70 being cultured with Brain Heart Infusion (BHI), minimum salt medium (MSM) and
71 BHI agar plates at 37°C for 4 weeks. Cultured bacteria and PE films which pretreated
72 with ethanol and bleach incubated in an incubator shaker for 3 months, then about 30%
73 of PE (about 4.5 mg) was degraded (Bardaji et al., 2019). Summarily, the results about
74 the plastics microbial degradation were gained through the simulated environment of
75 the laboratory. Because the conditions required for microbial degradation of plastics are
76 complex, specific culture medium, growth temperature, and pretreatment requirements
77 for plastics are needed (Magnin et al.,2020, Yang et al., 2018). This is not conducive to
78 the application of microorganisms to the actual treatment of waste plastics.

79 Insect is increasing to use for the degradation of plastics. Insect has better
80 processing effect and stronger applicability than microorganisms (Yang et al., 2015).

81 Moreover, insect growth conditions are not as strict as those of microorganisms, making
82 it more suitable for practical applications. The larvae of *Tenebrio molitor* (*T. molitor*)
83 have been proven to ingest and biodegrade PS and PE to CO₂ and lower molecular
84 weight compounds within their gut (Brandon et al., 2018, Wu et al., 2018). Yang et.al
85 (Yang et al., 2018) reported that the PS and bran were fed to *T. molitor* larvae in the 32-
86 day culture test at 20~30 °C, and up to 84 % of PS (about 1.5 g) was consumed. In
87 another research, 47.7% of the ingested Styrofoam carbon was converted into CO₂ in
88 16 days under similar reaction conditions (Brandon et al., 2018). Moreover, the larvae
89 of *Galleria mellonella* (*G. mellonella*) are also capable of chewing and ingesting PE
90 films (Yang et al., 2014, Zhang et al., 2020). The larval homogenate smeared on PE
91 films caused mass loss of 13% (about 92mg) in 14 hours (Bombelli et al., 2017). The
92 plastics used in the above studied are all new thermoplastic raw material, which are not
93 waste, not added catalyst or additives and linear low-density such as PE and PS. The
94 performance of different insects degrading plastic is also compared (Yang et al., 2021,
95 Peng et al., 2019, Billen et al., 2020).

96 However, there are many impurities in plastics from WEEE, such as metals, freon
97 and polycyclic aromatic hydrocarbons (PAHs) in waste refrigerator plastics (Mao et al.,
98 2020, Zhang et al., 2020, Dement'ev et al., 1991). This makes WEEE plastics more
99 hazardous and more treatable than ordinary fresh plastics. Therefore, this paper wants
100 to explore the difference between WEEE and fresh plastics during degradation of *G.*
101 *mellonella* larvae. In addition, a variety of plastics coexist in the actual environment.
102 There are still many types of plastics in WEEE that lack research on degradation of *G.*

103 *mellonella* larvae to co-exist plastics, such as rigid thermosetting PU, hard lumpy
104 engineering plastics HIPS as well as PVC and PF plastics which account for a large
105 proportion. Their physical and chemical properties such as density and functional group
106 are very different which may affect the degradation of these plastics by *G. mellonella*
107 larvae. When insects face different plastics, there may be a certain preference and
108 selectivity under the action of many factors. As far as we know, there is little
109 information about how insect larvae deal with different plastics in WEEE. In addition,
110 the performance of different insects in the face of the same WEEE plastics is also worth
111 exploring.

112 In this study, *G. mellonella* larvae were fed with nine types of plastics commonly
113 found in WEEE, including three waste plastics, i.e. waste HIPS (WHIPS), waste rigid
114 PU (WRPU), waste PS (WPS), and six fresh plastics, i.e. rigid polyurethane (RPU), PF,
115 PE, PS, PP, PVC. Selection considers representative, these plastics basically include the
116 most content plastics in WEEE. The influences of plastics and insect types on
117 degradation, feeding selectivity of insect larvae were investigated and analyzed. X-ray
118 fluorescence spectrometer (XRF) was performed to analyze the element composition
119 between the fresh and WEEE plastics. Scanning electron microscope (SEM) and
120 Fourier transform infrared (FT-IR) were performed to characterize the changes in
121 morphology and functional groups of the plastic feedstock and *G. mellonella* larvae's
122 frass. Besides, as the most typical thermosetting WEEE plastics, WRPU plastic was
123 selected to compare the differences between *G. mellonella*. and *T. molitor* larvae.

124 **2. Materials and Methods**

125 **2.1 Materials**

126 The WRPU, WHIPS and WPS plastics (Fig. S1a, b, c) were gained from an
127 environmental protection company in Shanghai, China. WRPU is insulation material
128 from the waste refrigerator. WHIPS is engineering plastic from the shell of a WEEE.
129 WPS is insulation material from a WEEE. WRPU and WPS are also foamed plastics.
130 Fresh RPU, PF, PE, PS, PP and PVC plastics were uniformly selected to compare
131 feeding selectivity of insect larvae to different plastics. Fresh plastics were all foamed
132 plastics and purchased from a plastic supplier in Shanghai, China. The plastics were cut
133 into cubes with a side length of 1 cm, typically like PS (Fig. S1d).

134 *G. mellonella* larvae and *T. molitor* larvae (Fig. S1e, f) were purchased from a
135 breeding farm in Yancheng, Jiangsu, China. *G. mellonella* was mature larvae of about
136 6th instar, with high activity, 25-30 mm in length, and 310-370 mg / worm mass. *T.*
137 *molitor* was mature larvae of about 5-6th instar, with high activity, 20-25 mm in length,
138 and 55-110 mg / worm mass. All larvae were experienced starvation treatment for 3
139 days before the experiment.

140 **2.2 Methods**

141 **2.2.1 Plastics feeding Insect Larvae tests**

142 According to the pre-experiment of *G. mellonella* larvae ingesting plastics, 2.0 g
143 of WHIPS, WPS, WRPU, PS, RPU plastics and honeycomb (HC) as the only carbon
144 source were fed to 40 *G. mellonella* larvae in incubators of 30 °C and full darkness for

145 7 days, respectively. Remaining mass of plastics was weighed, the generated frass was
146 collected and survival rate of the larvae was measured every day.

147 In the experiment exploring the feeding selectivity of *G. mellonella* larvae to six
148 fresh foamed plastics, 40 *G. mellonella* larvae were as a group. Each group was fed
149 with 30 pieces of different kinds of fresh foamed plastics, including RPU, PS, PP, PVC,
150 PF and PE. One kind (30 pieces for each), two (15 pieces for each), three (10 pieces for
151 each) or six kinds (5 pieces for each) of plastics were combined differently. Other
152 experimental condition was the same as the above.

153 *G. mellonella* and *T. molitor* larvae were selected to compare the behavior of
154 feeding WRPU. According to the same total weight, 40 *G. mellonella* (or 100 *T. molitor*)
155 larvae were as a group, respectively. Each group was cultured for 20 days, the situation
156 of the larvae was observed and recorded every 2-4 days. Other experimental condition
157 was the same as the above.

158 **2.2.2 Analysis and Characterization Methods**

159 Fresh and WEEE plastics were measured by XRF (XRF-1800, Shimadzu Ltd,
160 Kyoto, Japan), to explore the element composition. SEM and FT-IR were used to
161 characterize the changes in surface topography and functional groups of the feedstock
162 and frass of WEEE plastics, to explore the effect of ingestion on the plastic properties.
163 The samples were crushed, sieved through 200 mesh, and then dried at 80°C for 24
164 hours before analysis. SEM images were determined using a JSM-6700F microscope
165 (Royal Dutch Philips Electronics Ltd, Eindhoven, Netherlands) at 30 volts. FT-IR

166 spectra were gained using a NICOLET 6700 FT-IR Spectrometer (Thermo Fisher
167 Scientific Corporation, Massachusetts, USA) over the range of 400-4000 cm^{-1} ,
168 resolution of 4 cm^{-1} and scanning step of 1 cm^{-1} . All tests were repeated in triplicate.

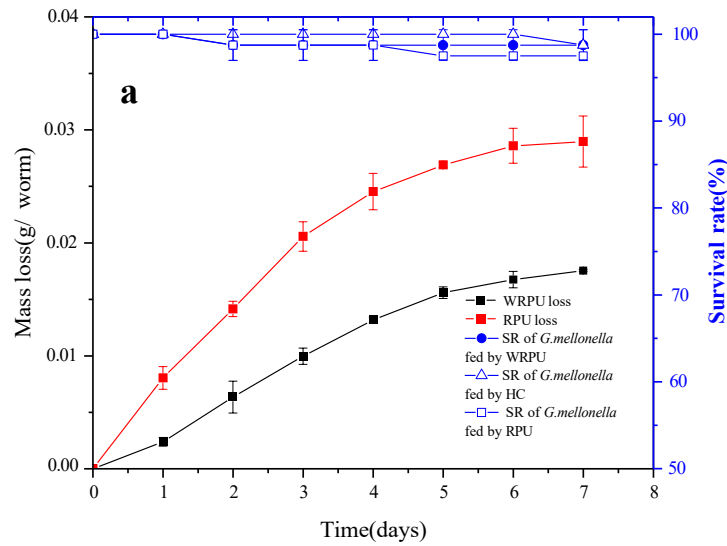
169 **3. Results and Discussion**

170 **3.1 Feeding Selectivity of *G. mellonella* Larvae to Kinds of Fresh and WEEE** 171 **Plastics**

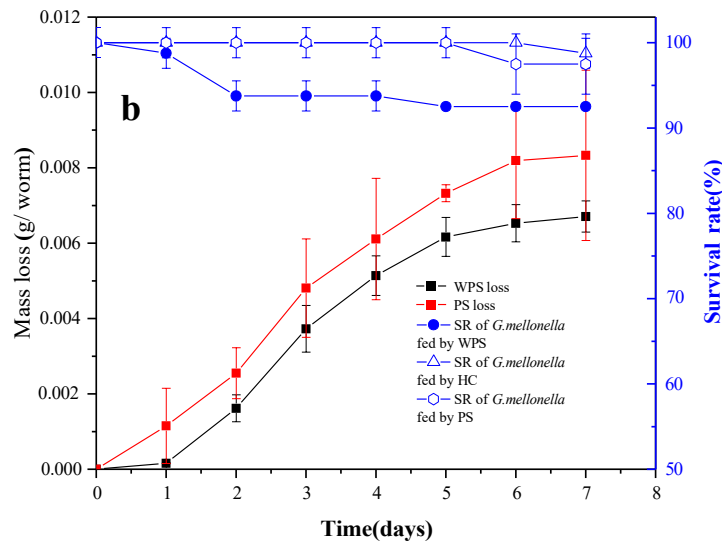
172 Some research on insects degrading plastics focused on PS (Peng et al., 2019,
173 Billen et al., 2020). As the representative of thermosetting plastics and thermoplastics,
174 RPU, PS, WRPU and WPS foamed plastics from WEEE were selected as the sole
175 carbon source respectively, to investigate the difference and selectivity of *G. mellonella*
176 larvae being fed fresh and WEEE plastics.

177 The consumption and survival rate of RPU, WRPU, PS, WPS plastics and HC by
178 *G. mellonella* larvae are measured (Fig.1). After one week, the consumption of RPU
179 and WRPU plastics are about 1.16 g and 0.70 g, respectively. And the average
180 consumption of RPU and WRPU plastics are about 4.14 mg / worm / d and 2.53 mg /
181 worm / d, respectively (Fig. 1a). The consumption of PS and WPS plastics are about
182 0.33g and 0.27 g, respectively. And the average consumption of PS and WPS plastics
183 are about 1.18mg / worm / d and 0.96 mg / worm / d, respectively (Fig. 1b). The
184 consumption of *G. mellonella* larvae to the WRPU and WPS plastics is lower than the
185 corresponding fresh plastics, implying that the larvae prefer the fresh plastics to the
186 WEEE plastics. There are presumably two reasons for the difference. One reason is that

187 waste plastics have strong smell and migration of heavy metals, etc (Mao et al., 2020,
 188 Liu et al., 2019). The other reason is the contents of metals and additives in waste and
 189 fresh plastics are different. As shown in Table S1a, there is chlorine in WRPU, which
 190 is probably from refrigerator foaming agent-Freon (Dement'ev et al., 1991). The metal
 191 contents in WPS is much higher than that in PS (Mao et al., 2020) (Table S1b). The
 192 strong smell and metals migrating to the surface can reduce the appetite of larvae
 193 ingesting waste plastics. The chlorine and metals can poison the gut microbes in the
 194 larvae (Caravelli et al., 2004, Jacquioda et al., 2018).



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197 Fig. 1. Consumption and survival rate of *Galleria mellonella* larvae fed with HC, RPU
198 and WRPU (a) or PS and WPS (b). HC, honeycomb; RPU, rigid polyurethane; PS,
199 polystyrene; WRPU, waste rigid polyurethane; WPS, waste polystyrene.

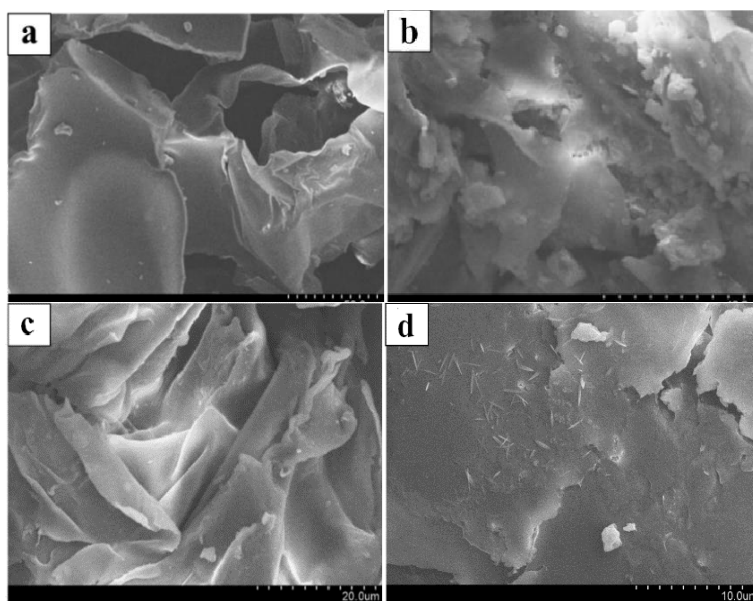
200 Moreover, the consumption of *G. mellonella* larvae to the WPS and PS are much
201 lower than that of the WRPU and RPU. The SR of the WRPU group is similar to that
202 of the RPU and HC group (Fig. 1a), and the SR of the WPS group is significantly lower
203 than that of the PS and HC group (Fig. 1b). It is inferred that compared with WPS and
204 PS, WRPU and RPU contains not only carbon source but also nitrogen and less metals
205 (Peng et al., 2019). The metal contents may have a greater impact on the *G. mellonella*
206 larvae feeding on WPS. But the SR of plastic-fed *G. mellonella* larvae all remains above
207 90% in one week, suggesting that the SR of *G. mellonella* larvae is not significantly
208 affected by WEEE plastics in the experiment period.

209 **3.2 Analysis of the Frass Produced by *G. mellonella* Larvae Ingesting WEEE** 210 **Plastics**

211 After the WRPU and WPS plastics were ingested, it deserves to investigate
212 whether they are biodegraded by *G. mellonella* larvae according to the physical and
213 chemical changes of the feedstock and frass of WEEE plastics. Fig. S2a shows that the
214 plastics produce holes and even fragments after being gnawed by the larvae, implying
215 the *G. mellonella* larvae have a strong gnawing effect on the block-shaped WRPU
216 plastics. SEM images of Fig.2 observe that microscopic morphology changes from
217 honeycomb-like cellular structure of the WRPU plastics (Fig. 2a) to eroded sheet-like
218 structure of the WRPU frass which has generated many small fragment (Fig. 2b). It

219 indicates that *G. mellonella* larvae destroy the physical structure of WRPU plastics and
220220 produce an erosion effect in the intestine.

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222

223 Fig. 2. SEM of WRPU feedstock (a) and frass (b), and WPS feedstock (c) and frass (d).

224 WRPU, waste rigid polyurethane; WPS, waste polystyrene.

225 Fig. S2b shows that the WPS plastics are less damaged than the WRPU plastics.

226 The result seems to confirm the finding in Fig.1 that the consumption of *G. mellonella*

227 larvae to the WPS is lower than that of the WRPU. SEM images of Fig.2 observe that

228 microscopic morphology changes from sheet-like wrinkled structure of the WPS

229 plastics (Fig. 2c) to flatter, cracks and small needle-like particles structure of the WPS

230 frass through the function of the intestine of *G. mellonella* larvae. (Fig. 2d). These

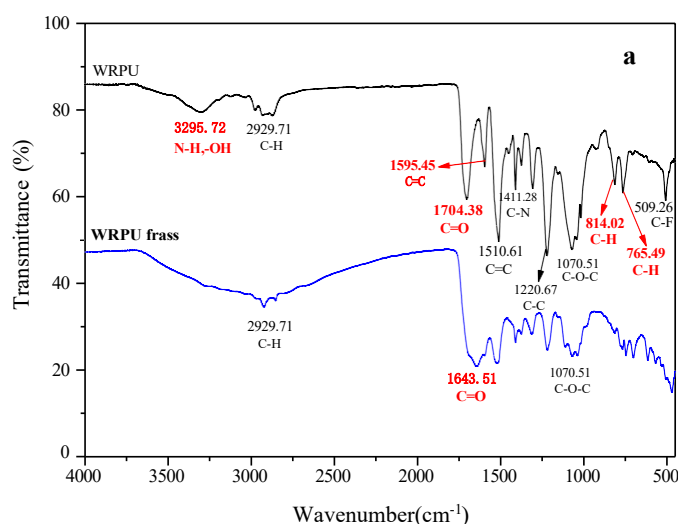
231 phenomena indicate that the feeding process of *G. mellonella* larvae on WRPU and

232 WPS plastics is to gnaw the plastic into granules and then enter the intestine for

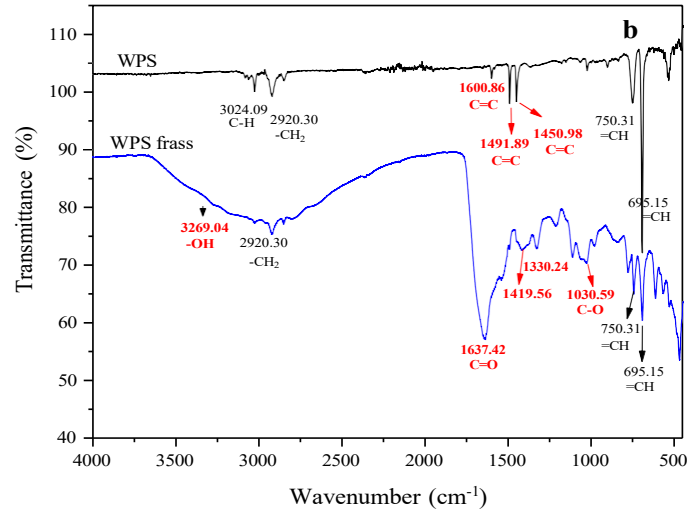
233 digestion. However, the products of digestion-frass have different forms.

234 Subsequently, chemical structures of the WRPU, WPS plastics and the

235 corresponding frass were analyzed by FT-IR. Fig. 3a shows the spectra of WRPU frass
 236 have the significant changes in three regions: 3250-3330, 1550-1710 and 500-900 cm⁻¹
 237 ¹. These changes indicate that some functional groups bonds are decomposed and
 238 transformed to the new bonds. *G. mellonella* larvae mainly acts on the hard segments
 239 of WRPU mainly composed of urethane bonds (Oprea et al., 2010). The absorption
 240 peaks of 3296 cm⁻¹ gradually broaden until they merge with those at 2930 cm⁻¹ to form
 241 the wider absorption peak. This can be presumably attributed to the H- bonded N-H and
 242 -OH of the biodegradation products. The increase in the hydroxyl suggests a possibility
 243 of conversion from hydrophobicity to hydrophilicity of polyurethane (Oprea et al.,
 244 2018). The rupture of the urethane bond in the main chain of WRPU at 1704 cm⁻¹, the
 245 formation of hydrogen bonded carbonyl at 1644 cm⁻¹, and disappearance of C=C and
 246 C-H peaks of benzene rings at 1595, 814 and 765 cm⁻¹, all indicate that WRPU has been
 247 not only ingested, but also degraded. In addition, no significant change occurs in ether
 248 bonds (1071 cm⁻¹), because the soft segment of WRPU dominated by the ether bond is
 249 more bio-resistant.



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Fig. 3. FT-IR spectra of WRPU feedstock and frass (a), and WPS feedstock and frass (b). WRPU, waste rigid polyurethane; WPS, waste polystyrene.

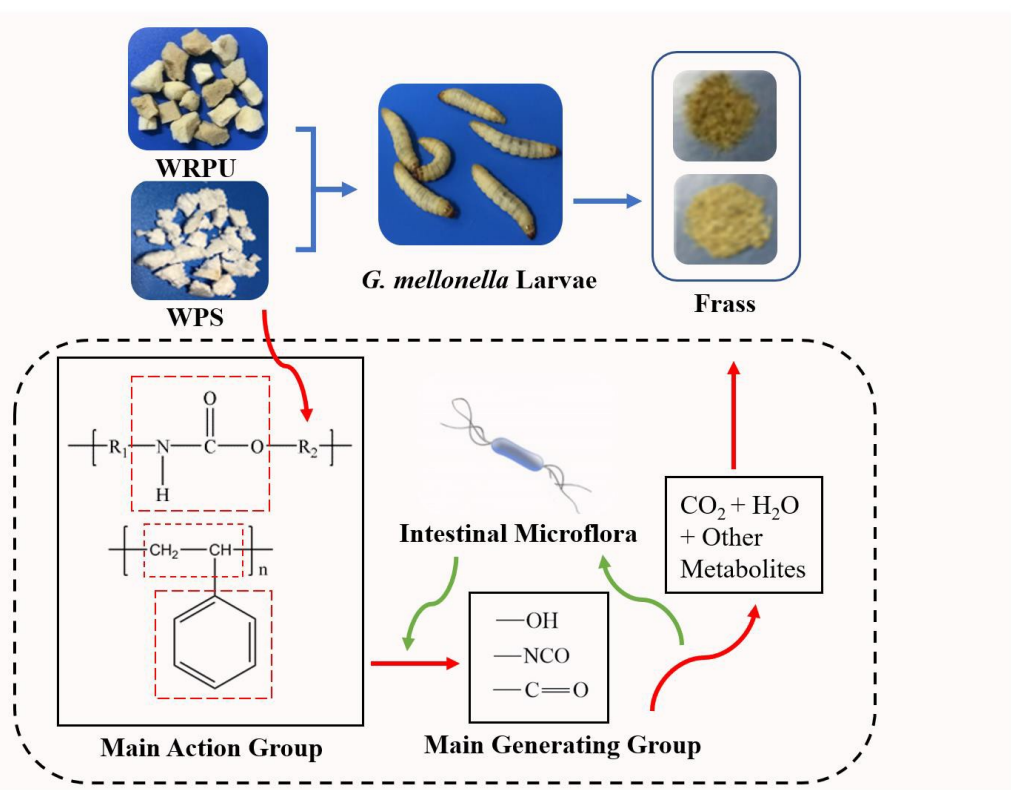
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254 FT-IR spectra of the WPS frass have significant changes in two regions: 2500-
 255 3500 and 1400-1710 cm^{-1} , compared with the WPS (Fig. 3b). Like Fig. 3a, the changes
 256 in 3269 cm^{-1} absorption peaks representing the WPS benzene ring (C=C stretch, 1400-
 257 1710 cm^{-1}) are dampened in the frass samples, indicating the ring cleavage (Yang et al.,
 258 2018). Further evidence of degradation is the observed decrease in intensities of peaks
 259 characteristic for WPS and the appearance of -C=O- (1637 cm^{-1}) (Yang et al., 2014,
 260 Sekhar et al., 2016). The broadening of peaks at 2500-3500 cm^{-1} is associated with the
 261 hydrogen bond of hydroxyl groups and/or carboxylic acid groups, suggesting a shift
 262 from hydrophobic to more hydrophilic surface properties (Lou et al., 2020).

263

In other words, *G. mellonella* larvae changes not only the physical structure but
 264 also chemical structure and composition of the WRPU and WPS plastics. As shown in
 265 Fig. 4, the biodegradation process of WRPU and WPS by *G. mellonella* larvae
 266 undergoes the hydrolysis or oxidation. According to the analysis of chemical groups,
 267 the urethane bonds in WRPU are broken due to the hydrolysis. However, the carbon

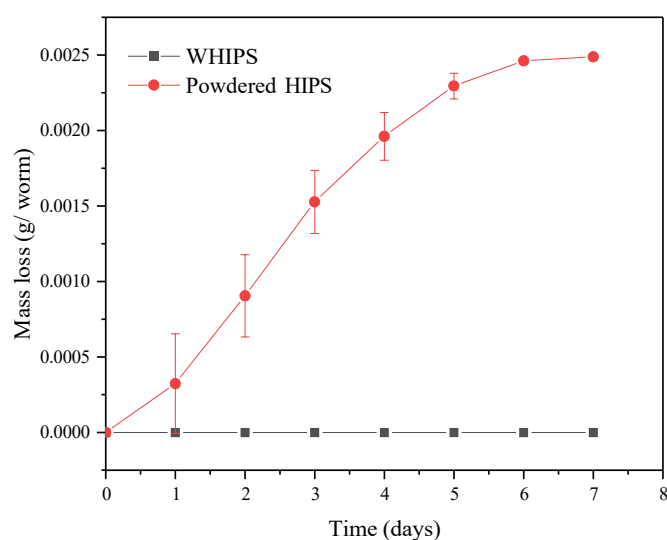
268 chain of WPS is mainly attacked, and oxygen-containing functional groups are
 269 generated due to the oxidation reaction. Depolymerization of high molecular weight
 270 substances occurs, and low molecular weight products are formed, which may be
 271 further assimilated or mineralized. The different degradation products of the two WEEE
 272 plastics are attributed to the differences in not only chemical composition but also
 273 structure of WRPU and WPS. The molecular configuration of the WRPU is a cross-
 274 linked body structure, while the molecular structure of the WPS is linear (Yuan et al.,
 275 2020).



276
 277 Fig. 4. Mechanism diagram of *G. mellonella* larvae biodegrading WRPU and WPS.
 278 WRPU, waste rigid polyurethane; WPS, waste polystyrene.

279 **3.3 Feeding Selectivity of *G. mellonella* Larvae to Apparent Modification of WEEE**
 280 **Plastics**

281 Most EEE plastics are lumpy and hard, such as ABS for TV shell and HIPS for
282 printer shell. So we investigated the effect of changing the appearance of WEEE
283 plastics-WHIPS on the feeding selectivity of *G. mellonella* larvae. Fig. S3a shows that
284 there are no signs of damage on the appearance of the lumpy WHIPS plastics. However,
285 after the lumpy WHIPS plastics are physically modified to grind into powders (Fig.
286 S3b), the powders are basically ingested by *G. mellonella* larvae and the corresponding
287 frass are generated (Fig. S3c). The result reflects that the *G. mellonella* larvae cannot
288 ingest WHIPS hard lumpy engineering plastics, but have a feeding behavior to the
289 powdered WHIPS plastics (Fig. 5). In fact, grinding WHIPS into powder only changes
290 its particle size and does not change its chemical properties. The above results indicate
291 that the appearance of plastics have a great effect on *G. mellonella* larvae ingesting the
292 plastics.



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294 Fig. 5. Consumption of *G. mellonella* larvae to the WHIPS and powdered WHIPS.
295 WHIPS, waste high impact polystyrene.

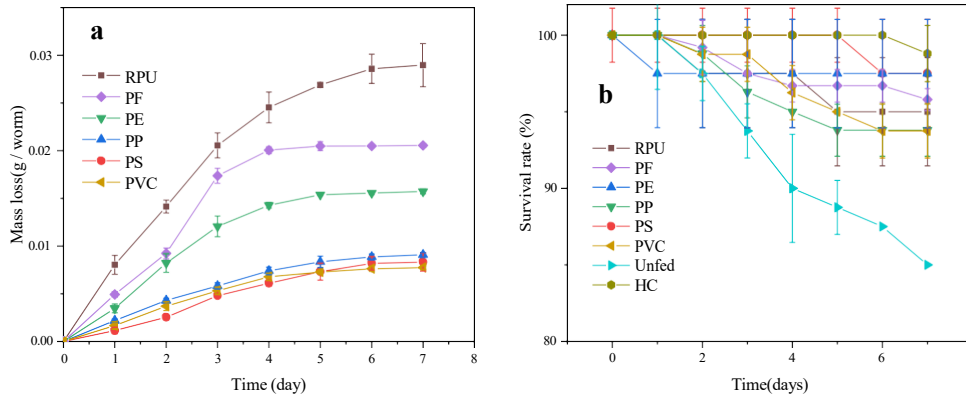
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297 **3.4 Feeding Selectivity of *G. mellonella* Larvae to Different Plastic Combinations**

298 In the actual environment, a variety of plastics usually coexist. Therefore, it
299 deserves to investigate feeding selectivity of *G. mellonella* larvae in the coexistence of
300 multiple plastics. In the study, fresh foamed plastics is collected to eliminate the
301 interference of plastic form and residual substances in WEEE.

302 [Fig. 6a](#) shows the consumption of *G. mellonella* larvae to the plastics fed solely.
303 The result presents a decreasing order as following: RPU>PF>PE>PP>PS>PVC. Lou
304 et al. ([Lou et al., 2020](#)) also reported that the consumption of the larvae to PE is higher
305 than PS. [Table S2](#) outlines the physical and chemical properties of these plastics.
306 Minimal mass loss of PVC in all of the plastics is possibly resulted from its excessive
307 density (291 kg/m³) preventing the larvae from breaking it. Poor abrasion resistance
308 and toughness of PF and RPU make them easy to decompose ([Yang et al., 2018](#)).
309 Compared with PF and RPU, PE, PP, and PS have relatively higher abrasion resistance
310 and toughness, causing their much lower mass loss. Research reported that the chemical
311 structures and functional groups of the plastics also affect their degradation ([Magnin et](#)
312 [al.,2020](#)) . Another reason that PE, PP, and PS are difficult to be biodegraded is their
313 stable carbon chain structure and simplified chemical groups. However, RPU and PF
314 contain urethane groups and hydroxyl groups, respectively, that are easily
315 biodegradable, supplying another explanation why the larvae prefer them ([Tang et al.,](#)
316 [2020](#), [Liu et al., 2019](#)). In addition, the survival rate (SR) of *G. mellonella* larvae in

317 each group remain above 90% (Fig. 6b), indicating that *G. mellonella* larvae can feed a
 318 variety of plastics, and the plastics have no obvious toxic effect on them in the
 319 experiment period.

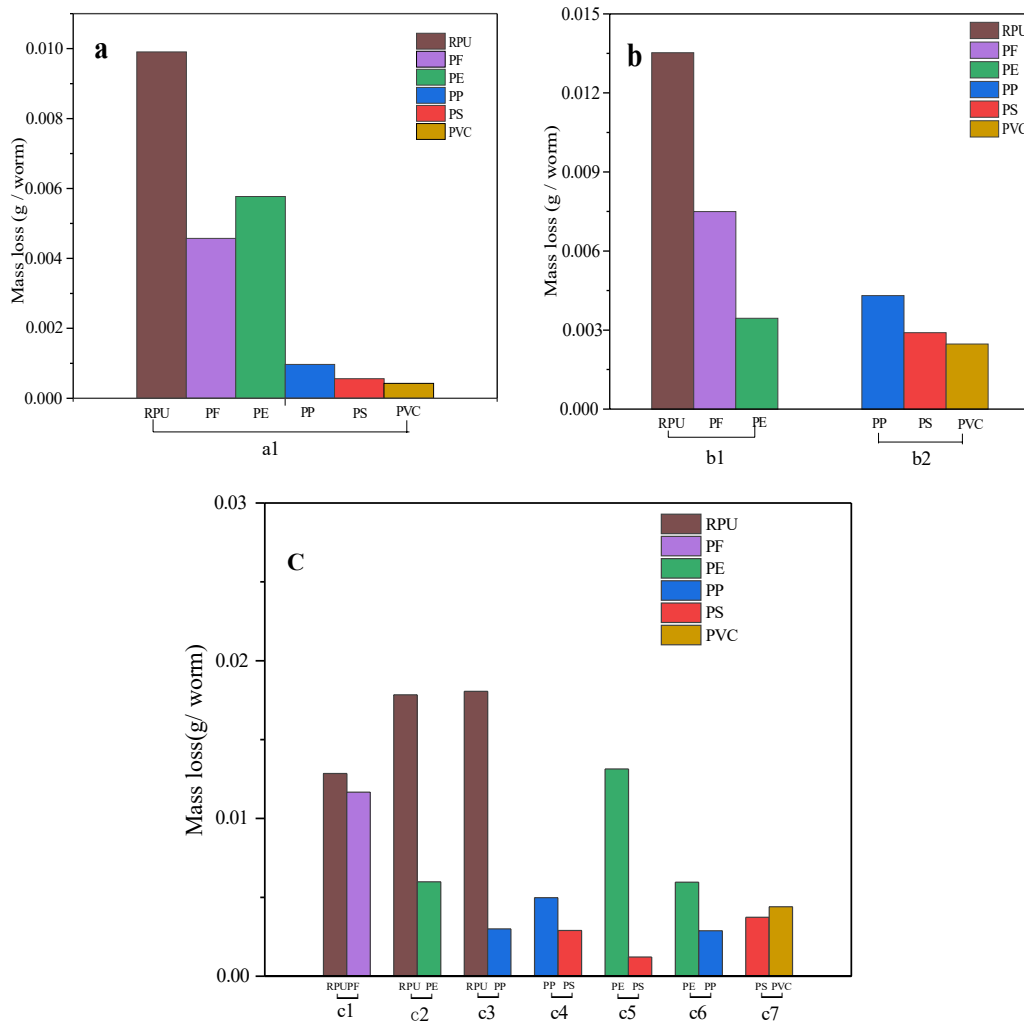


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321 Fig. 6. Consumption (a) and survival rate (b) of *G. mellonella* larvae fed with fresh
 322 RPU, PS, PP, PE, PF, PVC plastics, HC and unfed, respectively. RPU, rigid
 323 polyurethane; PS, polystyrene; PP, polypropylene; PE, polyethylene; PF, phenolic resin;
 324 PVC, polyvinyl chloride; HC, honeycomb; unfed, without any food.

325 The consumption of *G. mellonella* larvae to the plastics in different plastic
 326 combinations is shown in Fig. 7. Interestingly, no matter how the six fresh foamed
 327 plastics were combined to feed the *G. mellonella* larvae such as a group of six (Fig. 7a),
 328 three (Fig. 7b) or two kinds of plastics (Fig. 7c), they select to ingest some of each
 329 plastics instead of one plastics. Even if the plastics (PS or PE) was mixed with beeswax
 330 or bran as reported in the literature (Lou et al., 2020), the *G. mellonella* larvae did not
 331 only select beeswax or bran to ingest. And the consumption order of *G. mellonella*
 332 larvae to the plastics in different combinations is similar to that of fed by one kind of
 333 plastic alone (Fig. 6). The result indicates that there is no difference in the feeding
 334 selectivity of *G. mellonella* larvae in different plastic combinations. In addition,

335 although the consumption of *G. mellonella* larvae to each plastics in the different
 336 combination groups is less than that of the plastics fed alone, the total consumption of
 337 *G. mellonella* larvae is similar to the individual consumption of the plastics that the
 338 larvae prefer. It is different from the literature (Lou et al., 2020) reported that the
 339 consumption of *G. mellonella* larvae in the combination of the plastics and beeswax or
 340 bran was lower than that in only the plastics (PS or PE). The food consumption of *G.*
 341 *mellonella* larvae is limited, when beeswax or bran exist, the larvae prefer them and eat
 342 less plastics.



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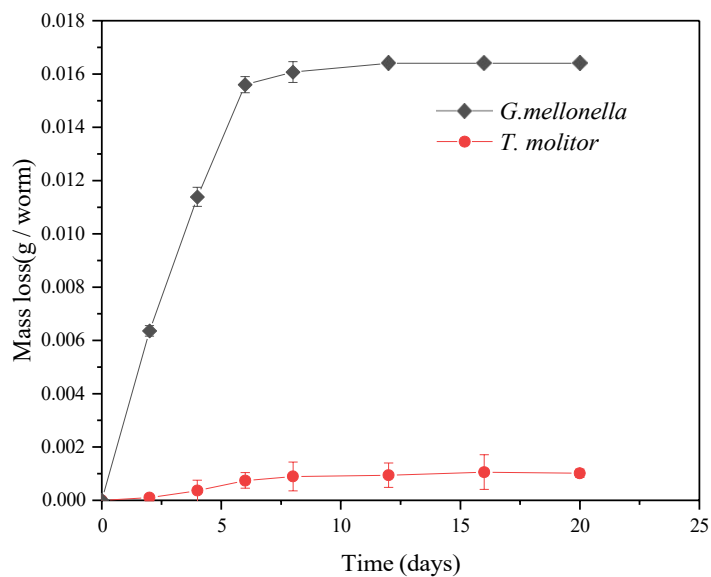
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346 Fig. 7. Feeding preferences of *G. mellonella* larvae co-fed with six kinds (a), three kinds

347 (b), two kinds (c) of different foamed plastics.

348 3.5 Selectivity of Two Insect Larva (*G. mellonella* vs *T. molitor*) being fed WRPU

349 The above results show that *G. mellonella* larvae show feeding selectivity to
350 different plastics. Meanwhile, it deserves to investigate the difference in two insect
351 larvae feeding to the plastics. After 8 days, the WRPU in the group of *G. mellonella*
352 larvae possesses many holes and even fragment (Fig. S4a), but in the group of *T. molitor*
353 larvae is not obviously damaged (Fig. S4b). As shown in Fig. 8, the plastic loss in the
354 *G. mellonella* larvae group is 0.66 g, while in the *T. molitor* larvae group is only 0.1 g
355 after 20 days. The result also indicates that the feeding activity of *G. mellonella* larvae
356 is higher than that of *T. molitor* larvae. It is no accident that *G. mellonella* larvae has a
357 better effect on ingesting WRPU plastics than the *T. molitor* larvae. The literature
358 (Billen et al., 2020) also showed that the consumption of *G. mellonella* larvae to the PS
359 or PE is higher than that of *T. molitor* larvae (mealworms).



360360

361 Fig. 8. WRPU consumption of *G. mellonella* and *T. molitor* larvae. WRPU, waste rigid

362 polyurethane.

363 The possible reason is that *G. mellonella* larvae has larger volume, high activity,
364 high food intake and well-developed gut system (Cassone et al., 2020, Wojda, 2017).

365 These factors will affect the survival and diet of insects (Matyja et al., 2020). The mass
366 loss of plastics in the *G. mellonella* larvae group increases rapidly during the first 8

367 days, and tends to stable after 8 days. Because the larvae of *G. mellonella* in the early

368 stage are very active and ingest a large amount of plastics, and then gradually pupate

369369 and no longer ingest after 8 days.

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371 **Conclusions**

372 In this study, the selectivity of *G. mellonella* larvae to the WEEE plastics in the

373 biodegradation process was explored. Fresh and powder plastics were preferred to

374 ingest by *G. mellonella* larvae due to less metals and additives than WEEE plastics. The

375 consumption of *G. mellonella* larvae to the plastics presents the decreasing order as

376 following: RPU>PF> PE>PP>PS>PVC regardless of feeding alone or in combination.

377 SEM and FT-IR analyses of WRPU and WPS frass complement and confirm the

378 biodegrading of the plastics. Simultaneously, *G. mellonella* larvae to WRPU showed

379 higher feeding activity than that of *T. molitor* larvae.

380 All the results indicate that the consumption of *G. mellonella* larvae to WEEE

381 plastics is lower than that of fresh plastics, but the larvae can ingest a variety of WEEE

382 plastics gnawed suitably. *G. mellonella* larvae has different degradation effects on

383 different WEEE plastics, and the degrading effect is related to the chemical and physical

384 properties of plastics. Different insect larvae have different capacity to degrade WEEE
385 plastics. The possible reason is that the intestinal microorganism of the *G. mellonella*
386 larvae have different effects on different plastics. The microorganisms preferring
387 different plastics might be screened out from the intestinal microflora of the insects.
388 Therefore, the mechanism of the action of insects on plastics needs to further investigate
389 through analyzing systemically the functional characteristics of the microbe.

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