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Abstract: Hydrogen production from waste activated sludge (WAS) was widely considered and intensively investigated as a promising technology to recover energy from wastewater treatment plants. To date, no efforts have been made on either systematic summarization or critical thinking of the application niche of hydrogen production from WAS treatment. It is therefore time to evaluate whether and how to recover hydrogen in a future paradigm of WAS treatment. In this critical review, the principles and potentials, microorganisms, possible technologies, and process parameters of hydrogen generation were analyzed. Microbial electrolysis cell shows high theoretical hydrogen yield and could utilize a variety of organic compounds as substrates, which is regarded as a prospective technology for hydrogen production. However, the poor organics utilization and rapid consumptions of produced hydrogen hindered hydrogen recovery from WAS. Based on the analysis of the current state of the literatures, the opportunities and challenges of hydrogen production from WAS are rethought, the detailed knowledge gaps and perspective of hydrogen production from WAS were discussed, and the probable solutions of hydrogen recovery from WAS treatment are figured out. To guide the application and development of hydrogen recovery, a more promising avenue through rational integration of the available technologies to form a hybrid process is finally proposed. The integrated operational paradigm of WWTPs could achieve substantial technical, environmental and economic benefits. In addition, how this hybrid process works is illustrated, the challenges of this hybrid process and future efforts to be made in the future are put forward.

Keywords: Hydrogen; waste activated sludge; anaerobic fermentation; hybrid process

1. Introduction

With the development of society, people need more and more energy, which inevitably leads to the shortage of fossil fuels. In addition, the combustion of fossil fuels makes large amounts of greenhouse gas (i.e., carbon dioxide) release in the atmosphere, causing the climate change and global warming. The issues of energy shortage and global warming drive the efforts to seek clean, recyclable, and renewable energy. Compared with methane, hydrogen possesses higher energy yield (i.e., 142.35 kJ/g which is 2.75 folds than that of other hydrocarbons) and generates water rather than greenhouse gases while it is combusted [1]. Therefore, hydrogen is considered as the green energy and widely accepted to be the most promising alternative to fossil fuels

Hydrogen can be produced from valuable resources such as water and fossil fuels, and also from wastes such as sewage and food waste via a series of chemical-physical and biological methods. There are many publications that reported hydrogen production by utilising the chemical-physical methods and biological methods [2]. Currently, most of the hydrogen (>85%) is produced through the pyrolysis of fossil fuels, and the gasification of biomass [3]. Although these chemical-physical methods could obtain high hydrogen yield, they are not sustainable due to high energy consumption [4, 5]. By contrast, biological hydrogen production is a more cost-effective and environmental friendly method to produce hydrogen from varieties of organic wastes (e.g., waste activated sludge) due to the simple operational conditions, steady H₂ yield and low energy consumption [6, 7]. As for different substrates, using wastes to produce hydrogen is an environmentally favorable and economically sustainable way. With the growing energy crisis worldwide, this aspect is becoming more important and pushing forward new attempts employing more wastes.

Waste activated sludge (WAS), which is the main byproduct of municipal wastewater treatment plants (WWTPs), is generated with large amounts annually [8]. For instance, it was documented that 11.2 million metric tons of dry sludge were generated in China while 10 million tons were produced in EU countries [9]. On one hand, treatment and disposal of such massive amount of WAS are costly, accounting for up to 60% of the total operation cost of a WWTP [10]. On the other hand, WAS contains high levels (50~70%) of organic compounds such as protein, carbohydrate, and lipid [11, 12], which makes it an ideal renewable resource. For example, Jiang et al. investigate the physicochemical characteristics of WAS and found that when the volatile suspended solid (VSS) of WAS was 10.81 g/L, WAS contained 14.88 g/L total chemical oxygen demand (TCOD), 9.94 g COD/L total protein, 0.86 g COD/L total carbohydrate, and 0.17 g COD/L lipid and oil [13]. Similar WAS characteristics were also reported in other papers [14, 15]. In addition, it was reported that ~35% of carbon element, ~3.8% of nitrogen element, ~1.6% of phosphorus element, and other trace elements contained in WAS [16].

Many publications showed that various wastes containing high organic substrates could be utilized to produce hydrogen by anaerobic fermentation process [17, 18], indicating that WAS is a potential substrate for hydrogen production. Although WAS is generally treated by the anaerobic digestion to produce methane, several hydrogen producers are found to be present in the digester such as *Clostridium pasteurianum* [19] and *Thermoanaerobacterium* [20, 21]. In fact, hydrogen is observed as an important intermediate in the

70 anaerobic digestion process [22]. Thus, hydrogen production from WAS attracted much attention in the past
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71 decades, by which fossil fuels are saved, greenhouse gas (e.g., CO₂) emission is reduced, WAS is reused and
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42 reduced, and sustainable clean energy H₂ and volatile fatty acids (VFAs) are also obtained. However, practical
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63 application of hydrogen production from WAS has not yet been achieved. On the contrary, some doubts and
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74 debates have been arisen recently about its technical and economic feasible in full-scale situations due to the
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175 low hydrogen yield. There are many challenges in reactor control, system development, and energy recovery.
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176 For example, hydrogen is an intermediate product in the anaerobic digestion, thus the produced hydrogen
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177 would be quickly consumed by hydrogen-utilizing methanogens to produce methane, homoacetogens to produce
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17 acetic acid, or sulfate-reducing bacteria to produce hydrogen sulfide. The highest hydrogen yield from WAS
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19 reported so far has been only 20.30 mg per gram volatile suspended solids [23].
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22 Several review papers were published on hydrogen production using the various wastes such as
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24 biodegradable municipal wastes [24], food waste [25], agriculture waste, wastewater [26], and lignocellulosic
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26 materials [27, 28]. These works were mainly to review the progress of hydrogen production in one aspect or
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28 several aspects. For example, Yang et al. provided a review on fermentative hydrogen production from sewage
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30 sludge but only focus on pretreatment methods and co-fermentation with other substrates [5]. Systematic
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32 summarization and critical thinking of the application niche of hydrogen production from WAS is still lacking.
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34 In addition, many endeavors were dedicated recently to improve hydrogen yield from WAS through enhanced
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36 the disintegration of WAS and suppressed or killed the competitive microorganisms (e.g., methanogens), which
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38 have made great progress [7, 29]. Hence, this review article aimed to comprehensively sum up the knowledge
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40 obtained in this field and critically think the prospect of biohydrogen production from WAS. To
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42 find out whether and how to recover hydrogen in a future paradigm of WAS treatment, the principles and
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44 advances of hydrogen production from WAS were needed to systematically review, its opportunities and
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46 challenges were required to critically re-examine, and the possible solutions were essential to carefully think
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48 about.
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52 Based on reviewing more than 200 publications and critically analyzing the opportunities and challenges
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54 of hydrogen production from WAS, this review aims to offer useful information to remove key barriers that
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56 hinder hydrogen production from WAS to be applied in full-scale situations, and to stimulate more thinking
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58 and discussion of a probable application niche for hydrogen production from WAS. To guide the application
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98 and development of hydrogen recovery, a more promising hybrid process through rational integration of the
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99 available technologies is proposed as an example, and how this hybrid process works is illustrate d and future
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100 efforts to be made in the future is discussed.
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101 **2. The pathways of hydrogen production from waste activated sludge**

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102 Several methods such as direct-biophotolysis, indirect-biophotolysis, photo-fermentation,
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103 dark-fermentation, and microbial electrolysis cell can be theoretically used for hydrogen production [30]. To
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104 date, however, only three approaches, i.e., dark-fermentation, photo-fermentation, and microbial eletrolysis
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105 cell, have been documented to produce hydrogen from WAS (Figure 1). Dark-fermentative hydrogen production
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106 is a process that uses organic matters in either soluble or solid state as electron donors and protons as electron
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107 acceptors to produce hydrogen by strict or facultative anaerobic bacteria. Photo-fermentative hydrogen
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207 production is a process that photo-fermentative bacteria (mainly purple nonsulfur bacteria) use light as energy
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208 and soluble organic matters (mainly small molecule organic acids) as electron donors to produce hydrogen. As
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209 for microbial electrolysis cell, exoelectrogenic microbes at the anode first oxidize organic matters into electrons
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210 and protons. The electrons produced are gathered at the anode and then transferred to the cathode, in which the
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211 electrons transferred are utilized to reduce protons for hydrogen production.
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35 **Figure. 1. Schematic diagram of hydrogen production from dark-fermentation (a) [31, 32],**
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37 **photo-fermentation (b) [33, 34] and microbial electrolysis cells (c) [35, 36].**

39 **2.1. Mechanism of hydrogen production from WAS dark fermentation**

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41 The main compositions of WAS are protein, carbohydrate, and lipids, and they usually locate in either
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437 extracellular polymeric substances or intracellular cells. Due to this characteristic, WAS must undergo
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438 disintegration and hydrolysis processes before the organics in WAS can be further degraded for hydrogen
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419 production. Dark-fermentation is the most intensively studied and widely used technique for hydrogen
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420 production from raw WAS. It is generally thought that the anaerobic digestion of WAS goes through five
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51 successive steps: disintegration, hydrolysis, acidogenesis, acetogenesis and methanogenesis (Figure 1a). As
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53 biosolids, sludge cells need to be disrupted in the disintegration step before the organics in sludge cells are
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523 utilized, and sludge disintegration is ordinarily considered the major rate-limiting step in anaerobic digestion
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524 [11, 22]. The released organics with large molecule weight such as protein and carbohydrate will be subjected
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126 to hydrolysis process, in which soluble organics with large molecule weight are degraded to small molecule
127 weight organics such as amino acids and glucose [37, 38]. Several enzymes including protease and
128 α -glucosidase are involved in this step [39, 40]. After disintegration and hydrolysis, there are several
129 pathways involved in acidogenesis and acetogenesis steps that can directly or indirectly produce hydrogen by
130 a series of hydrogen producers such as *Enterobacter* sp. and *Clostridium* sp. The detailed pathways are
summarized in Figure 2.

Figure 2. The pathways of hydrogen production from sludge dark fermentation [41-46].

134 It can be seen that all the major compositions of WAS (i.e., protein, carbohydrate, and lipids) could be used
135 as substrates for hydrogen production from dark-fermentation. Hydrogen could be directly produced via
136 deamination of amino acids and β -oxidation of long-chain organic acids [47]. Hydrogen could be also generated
137 via two different pathways from the degradation of pyruvate, an important intermediate produced from
138 glycolysis of carbohydrate and deamination of amino acids. Pyruvate is easily degraded to acetyl-CoA via
139 decarboxylate with reduced ferredoxin produced, which gives electrons to protons to generate hydrogen. This
140 is the dominant pathway for hydrogen production by *Clostridium* sp. [48]. The other pathway is formate
141 cleavage, which is generally dominated by facultative anaerobes, such as *Enterobacter* and *Klebsiella* [49, 50].
142 In the process of glucose glycolysis, a large number of NADH would be produced due to the shortage of electron
143 transport chain in fermentative bacteria. NADH and H^+ are usually oxidized into NAD^+ in the stage of
144 acidogenesis to maintain the proper ratio of NADH/ NAD^+ . However, when the oxidation of NADH is lower
145 than the production of NADH, excess NADH and H^+ would inevitably exist. To maintain
146 the normal metabolic activity, fermentative bacteria in this case would produce hydrogen from the oxidation
147 of excess NADH [51]. Besides, in the stage of H_2 -producing acetogenesis, some H_2 -producing acidogens
148 such as *Syntrophomonas wolfei*, and *Syntrophobacter wolinii* could convert propionic acid, butyric acid, ethanol
149 and other organics into acetic acid and hydrogen [52-55].

2.2. Mechanism of hydrogen production from photo fermentation system

150 After dark fermentation, there are a large number of organic compounds such as fatty acids and alcohols
151 left in the dark fermentation liquid, which has been demonstrated to be further bio-converted to hydrogen by
152 photosynthetic bacteria [56]. To recover more hydrogen from WAS, combining dark fermentation with

154 photo fermentation is therefore recommended by researchers [56]. The purple nonsulfur bacteria, which can
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135 acquire electrons from volatile fatty acids to produce hydrogen, are the most extensively studied
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156 photosynthetic bacteria in photo fermentation systems [33]. Organics in such systems are first oxidized and
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157 the electrons generated are transported to the photosystem by ubiquinone. Then electrons are energized by
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158 light and cycled in the photosynthetic electron transport chain, which would produce a proton gradient. The
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159 energized electrons are further transferred to ferredoxin via oxidoreductase and utilized to produce hydrogen by
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160 nitrogenase, with energy and ATP being derived from the proton gradient (Figure 1b) [33, 57].
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161 **2.3. Mechanism of hydrogen production from microbial electrolysis cell**

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162 Microbial electrolysis cells, which directly utilizes electrons generated from substrate degradations by
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203 exoelectrogens to reduce proton for hydrogen production, has also received much attention recently. In such
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264 systems, exoelectrogens at the anode first oxidize substrates to produce electrons, protons (H^+), and carbon
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265 dioxide. The electrons produced are gathered at the anode and then transferred to the cathode through an external
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266 circuit with an applied potential, while the protons move to cathode by diffusing directly through the electrolyte
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287 (single chamber) or membrane (double chamber). The electrons are finally used to reduce protons to produce
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368 hydrogen at cathode [35, 58] (Figure 1c).
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369 There are lots of organic matters that could be used as substrates to produce hydrogen by exoelectrogens,
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370 such as raw sludge [59], protein, organic acids [60], and cellulose [61, 62]. Like dark fermentation, raw sludge
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371 also requires to be disintegrated in the disintegration step before organic matters in sludge cells could be used
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492 by exoelectrogens. Generally, hydrolytic microbes, fermentative acidogenic bacteria, and exoelectrogens are
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173 co-existed in microbial electrolysis cells. On one hand, exoelectrogens
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474 could utilize the products of hydrolytic microbes and fermentative acidogenic bacteria at anode to generate
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475 electrons, thereby prompting the cascade utilization of organics in WAS efficiently through syntrophic
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496 interactions [63]. On the other hand, exoelectrogens would also compete with hydrolytic microbes and
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517 fermentative acidogenic bacteria for the available substrates such as soluble carbohydrate. It is noted that the
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578 electrons from oxidation of organics in dark-fermentation are given to the intermediate products. However,
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579 the electrons produced from exoelectrogens are gathered at the anode and transferred to the cathode, which
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580 would make the oxidation of organics completely to electrons, CO_2 and protons. If all the electrons
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681 transferred to the cathode are used to reduce protons, the hydrogen yield from microbial electrolysis cells
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would be greater than that from dark fermentation. The high theoretical hydrogen yield and the ability to utilize a variety of organic compounds (e.g., volatile fatty acids, sludge fermented liquid and WAS) make microbial electrolysis cell a prospective technology for hydrogen production.

Sludge disintegration is also considered as the rate-limiting step in microbial electrolysis cells, thus some pretreatment methods of sludge were used to prompt the disruption of extracellular polymeric substances and cell envelopes before the sludge entering into the microbial electrolysis cells. It was found that pretreatment methods could increase largely the yield and rate of hydrogen production from microbial electrolysis cells. Besides, adding fermentative bacteria into microbial electrolysis cells was also demonstrated to be an effective strategy to enhance hydrogen production. Wang et al. reported that combined sodium dodecyl sulfate pretreatment and microbial electrolysis cell could get the higher hydrogen yield of 8.5 mg H₂/g VSS, which was ~3.4-fold higher than using microbial electrolysis alone [60].

3. The bottlenecks of hydrogen production from WAS

3.1. The potential of hydrogen production from WAS

WAS shows huge theoretical potential in hydrogen production. The theoretical methane yield of 354 mg-CH₄ can be produced from WAS anaerobic digestion if 1 gram sludge cells (expressed as C₅H₇NO₂) are completely digested [64]. It is reported that 28% methane is produced from the hydrogenotrophic methanogenesis pathway [65], this indicates ~50 mg-H₂ would be consumed theoretically in this process. Besides, two other processes, i.e., homoacetogenesis and sulfate-reducing processes, are known to consume H₂ largely. The theoretical hydrogen yield, therefore, is much greater than 50 mg-H₂ if 1 gram of sludge cells is completely degraded in dark-fermentation process. When 1 gram acetate is used to produce

As mentioned above, however, hydrogen yield reported is always at very low levels even at bench-scale, and this is considered the bottleneck that hinders hydrogen recovery being implemented in field situations. It is documented that the highest hydrogen yields produced from WAS by the anaerobic fermentation or microbial electrolysis cells are only 20.3 mg-H₂/g-VSS[23]. Thus, why is hydrogen yield often at low levels?

3.2. Key factors affecting hydrogen production from WAS

182 would be greater than that from dark fermentation. The high theoretical hydrogen yield and the ability to
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183 utilize a variety of organic compounds (e.g., volatile fatty acids, sludge fermented liquid and WAS) make
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184 microbial electrolysis cell a prospective technology for hydrogen production.
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185 Sludge disintegration is also considered as the rate-limiting step in microbial electrolysis cells, thus some

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207 **3.2. Key factors affecting hydrogen production from WAS**

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208 There are several factors restricting the production of hydrogen from dark fermentation. First of all,
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209 organic matters (e.g., protein, carbohydrate, and lipids) contained in sludge are much underutilized due to the
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210 rigid structure of microbial walls.[37, 67] According to previous report [9], less than 40% of VSS reduction
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211 could be achieved in sludge anaerobic fermentation systems. The released organics are not completely
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212 utilized to produce hydrogen. Apart from a small amount of soluble substrates being used for the growth and
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213 maintenance of anaerobes[68], a large number of soluble organics such as protein, humic substances, volatile
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214 fatty acids, and alcohol are still remained in fermentation liquid. Secondly, as an intermediate product in the
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215 anaerobic digestion, the hydrogen produced is readily consumed by (1) hydrogen-utilizing methanogens to
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216 produce methane [69], (2) acetobacteria to produce acetic acid [70], and (3) sulfate-reducing bacteria to produce
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217 hydrogen sulfide [71], which reduce the final hydrogen yield directly.
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218 Although photo fermentation can in principle degrade soluble organics with small molecule weight (e.g.
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219 volatile fatty acids and alcohols) completely, there still exist some restrictions in practice. To ensure reaction
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220 efficiency, continuous illumination is required due to the low light conversion of purple nonsulfur bacteria.
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221 However, solar illumination is easily influenced by outdoor conditions while artificial illumination is expensive
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222 [72, 73]. The design and construction of photo-bioreactors are difficult and costly due to several limitations such
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223 as big surface-volume ratio and highly transparent [74]. Furthermore, the effluent directly from dark-
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224 fermentation is opaque and contains several toxic matters which would hinder light penetration and inhibit the
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225 activity of nitrogenase [75, 76].
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226 Microbial electrolysis cell is not mature enough to be applied in real world. When WAS is directly used as
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227 substrate, the microbial communities of anodophilic biofilms have not yet been known clearly. Since
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228 exoelectrogens and fermentative microbes such as homoacetogens and methanogens co-existed in microbial
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229 electrolysis cells[77-79], most of the organics released from WAS are metabolized by fermentative microbes
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230 rather than exoelectrogens, which could achieve degradation of complex organics but then lead to the low
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231 coulombic efficiency at the same time [59]. Carbon-based materials (e.g., carbon felt, graphite fibres, and
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232 graphite plates) are usually used as anode but they are conductivity-poor[80]. Noble metals such as platinum
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233 and palladium were also tried to use as cathode due to their outstanding catalytic activity but they are also
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234 costly. It remains a challenge to find proper materials to make anode and cathode [81, 82]. There are two
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235 kinds of reactor configurations (i.e., single chamber and double chambers) usually used in microbial
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236 electrolysis cells [83, 84]. The double chambers use membrane to separate cathode and anode, leading to the
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237 high internal resistance. Although single chamber is without membrane, it makes hydrogen produced at
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238 cathode easily consumed by anodophilic microbial communities [85, 86].

239 **4. Recent efforts to improve hydrogen production from waste activated sludge**

240 **4.1. Recent progress in hydrogen production from WAS dark fermentation**

241 In the past years, many endeavors were dedicated to improving hydrogen yield from WAS (Table 1). In
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242 order to enhance the disintegration of WAS and to suppress or kill the competitive microorganisms (e.g.,
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243 methanogens) in dark fermentative systems, many WAS pretreatment methods such as heating [87], acid [88],
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244 alkaline, and ultrasonic [89, 90] pretreatments were tested. Xiao et al.[91] found that compared with the control,
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245 thermal pretreatment at 121°C for 30 min enhanced soluble chemical oxygen demand from 113.7 to 2442.1 mg/L
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246 and reduced the dry solid from 8.96 to 7.55 g/L due to the acceleration of disruption of floc structure and
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247 microbial cells. As a result, hydrogen yield was improved from 1.46 mL H₂/g VSS to 8.62 mL H₂/g VSS. Cai
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248 et al. [6] found that alkaline pretreatment not only facilitated the disruption of WAS but also suppressed the
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249 activities of hydrogen-consuming microorganisms, which improved hydrogen yield from 9.1 mL H₂/g DS to
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250 16.6 mLH₂/g DS. The summarization of these pretreatments was detailed in a recent review [92]. Hydrogen
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251 yields from anaerobic fermentation of WAS were expressed by a variety of indexes and units such as mL/g VSS,
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252 mL/g-total solids (TS), and mL H₂/g DS, which was reviewed in a recent paper[5].
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253 Manipulating the fermenters was also documented to be effective. Zhao et al.[93] found that compared
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254 with alkaline pretreatment (i.e., initial pH 10) alone, controlling pH in the fermenter at constant pH 10 caused
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255 47.8% higher in hydrogen yield (18.2 versus 26.9 mL H₂/g VSS). It was reported that shortening hydraulic
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256 retention time from 8 h to 6 h benefited to wash out H₂-consuming bacteria (e.g., propionic acid bacteria), as
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257 their growth rate is slower than H₂ producers'[94]. A relative high fermentative temperature facilitated the
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258 hydrolysis of substrates and simplified microbial diversity, which thereby profited for hydrogen accumulation.
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259 Yokoyama et al. observed the activity of H₂-consuming microbes was completely suppressed when the
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260 fermenter was controlled at 75 °Cfor the treatment of cow waste slurry[95].
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261 The characteristics of fermented WAS (e.g., the ratio of carbon to nitrogen and the level of
1 polyhydroxyalkanoates) affected hydrogen yield as well. It was documented that pertinent increases of
262 carbon/nitrogen ratios benefited the conversion of protein while an increase of sludge polyhydroxyalkanoates
3 enhanced the cell solubilization, the hydrolysis process of solubilized substrates, and the soluble protein
263 conversion of non-polyhydroxyalkanoates biomass, thereby promoting the production of hydrogen [96].
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264 Based on these findings, optimizing the composition of the fermented WAS via either promoting the content
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267 Manipulating hydrogen-producers was also helpful to improve hydrogen yields, such as inoculating some
268 specific microbes and using metabolic engineering to modify hydrogen-producers in fermenters. Kotay et
269 al.[101] constructed a microbial consortium by inoculating with three established bacteria (i.e., *Enterobacter*
270 *cloacae* IIT-BT 08, *Citrobacter freundii* IIT-BT L139 and *Bacillus coagulans* IIT-BT S1) and found that
271 hydrogen yield could be improved 1.5-4 times with different ratio of these bacteria. Among these, 1:1:1 v/v ratio
272 of these three bacteria was found to achieve the maximal hydrogen yield of 41.23 ml H₂/g COD_{reduced}. Kim et
273 al. engineered the pentose phosphate pathway to enhance hydrogen production in recombinant *Escherichia coli*
274 and found that hydrogen yield increased by 3.5-fold, i.e., 21.665 mmol H₂/mol glucose
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366 WAS, as the byproduct of wastewater biological treatment, adsorbs and concentrates lots of exogenous
367 pollutants such as heavy metal, nanoparticles and micropollutants [106, 107]. Recently, some studies have
368 investigated the effects of exogenous pollutants on hydrogen production from WAS anaerobic fermentation.
369 Wu et al. found that 20 mg/g TSS poly aluminum chloride (a widely used inorganic coagulant) increased
370 hydrogen accumulation from 20.9 mL to 27.4 mL/VSS. Mechanism analysis shows poly aluminum chloride
371 inhibits the activities of all the microbes related to anaerobic fermentation, and its inhibition to
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288 hydrogen-consuming microorganism was much severer than that to hydrogen producer [108]. Wang et al.
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289 reported that 1403 ± 150 mg/kg TSS triclocarban (an antimicrobial agent) enhanced the activities of acetate
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290 kinase and [FeFe] hydrogenase and restrained the activities of carbon monoxide dehydrogenase and
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291 Coenzyme F₄₂₀, thus improved hydrogen production from 10.1 to 14.2 mL/g VSS during WAS anaerobic
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292 fermentation [109]. Wei et al. found that polyethylene terephthalate microplastics inhibited hydrolysis,
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293 acidogenesis and acetogenesis processes, thus reducing hydrogen production from WAS anaerobic
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294 fermentation [14]. Many exogenous pollutants show dose-dependent impacts on WAS anaerobic fermentation
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295 [110]. The results exhibit that most of the pollutants at low dose could improve the hydrolysis, acidification and
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296 methanogenesis processes while high dose show negative influence due to their toxicity [111, 112]. Ordinarily,
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297 the activities of methanogens are more easily inhibited than those of hydrolytic and acidogenic bacteria due to
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298 the less tolerance of methanogens [113, 114]. However, there are still few studies to investigate the effect of
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299 exogenous pollutants on hydrogen production. More efforts are required to unveil and regulate their influences
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300 on hydrogen production from WAS anaerobic fermentation.
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301 Additionally, to further utilize the organics left in the dark fermentation liquid such as fatty acids and
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302 alcohols, two-stage systems (e.g., dark fermentation-photo fermentation or dark fermentation-microbial
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303 electrolysis cells) were also proposed to improve hydrogen yield [63, 115-117]. For example, it was reported
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304 that a combined process (i.e., dark fermentation-microbial electrolysis cell) was used to recover bioenergy from
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305 WAS and acquired 20.30 mg H₂/g VSS of hydrogen yield while only 1.39 mg H₂/g VSS of hydrogen production
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306 from WAS anaerobic fermentation alone [7, 23]. The combined of dark fermentation and microbial electrolysis
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4.2. Recent progress in hydrogen production from photo fermentation

309 Previous investigations made on photo-fermentative hydrogen production mainly focused on
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optimization of the fermenter construction and operation, metabolic engineering of hydrogen producers, or
wavelength and intensity of light. Glass and polymethyl methacrylate are considered the most suitable
materials for constructing photobioreactor[74]. Nitai et al.[118] reported that the suitable pH and
temperature ranges for photo-fermentative hydrogen production were 6.8-7.5 and 31-36 °C, respectively.
Several metabolic engineering methods were used to manipulate H₂-producer, such as strain improvement by

315 modifying light-harvesting antenna complexes and replacement of nitrogenase with hydrogenase[119].
1
316 Kondo et al.[120] found that strains with a 41%-49% decrease in LH2 (LH: light-harvesting) content by UV
3
317 irradiation produced 50% more hydrogen than untreated strains. Different light wavelengths and intensities
5
318 also affected hydrogen yield. It was reported[121] that the suitable light wavelength and intensity were in
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8
319 the range of 400-1000 nm and 6-10 klux, respectively. The details about the impact of light wavelength and
10
320 intensity on hydrogen production could be found in a recent review[118].
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13
321 In the dark-photo fermentation combination system, however, some pretreatments of the effluent from
15
322 dark fermentation (e.g., dilution, ammonium stripping, and centrifugation) are necessary before the effluent is
17
323 used for hydrogen production through photo-fermentation [122, 123]. Yetis and coworkers[124] reported that
19
324 when the concentration of fermentation medium was at 30% dilution of pretreated sugar refinery wastewater,
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22
325 hydrogen production would be stopped, but a small amounts of hydrogen would be produced (i.e., 130 mL H₂/L)
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326 at 20% dilution.
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27 **4.3. Recent progress in hydrogen production from microbial electrolysis cell**

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327 Efforts made on microbial electrolysis cell mainly focused on optimizations of the three aspects:
32
328 improvement of electrode materials, reactor configuration, and manipulation of microbial community. Electrode
34
329 is one of the most significant part of microbial electrolysis cell. While bacterial activities mainly occur at anode,
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330 hydrogen is produced at cathode, so in microbial electrolysis cell, it is important to find the highly efficient and
37
331 low cost electrode materials (included anode and cathode) to achieve microbial electrolysis cell practical
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332 implication. Based on these principles, there are many new, cheap and effective anode materials (e.g., carbon
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333 nanotubes and graphene[125, 126]) and cathode materials (e.g., carbon nanotube
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334 and biocathode [127, 128]) developed, which are helpful to reduce the cost and improve hydrogen yields. Xie
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335 et al.[129] claimed that graphene-sponge was cheap and easy to fabricate which 1 m² 2 mm-thick
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336 graphene-sponge would only cost \$4 compared to 1 m² carbon cloth cost \$1000[130]. Rozendal et al.[131]
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337 reported that 0.63 m³-H₂ per 1 m³ cathode liquid volume would be produced from the microbial biocathode
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338 compared to 0.08 m³-H₂ produced from the control electrode. In addition, anode surface characteristic is one
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339 of the most important factors to affect microbial attachment and electron transfer, so many treatments such as
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340 heat treatment and acid treatment[132] on anode materials surface have been used to improve the properties of
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342 anode surface. It was reported that carbon mesh heated at 450 °C for 30 min could produce the maximal
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343 power density of 46 W/m³, which produce 3% more power than that using cleaned carbon mesh treated with
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344 acetone in microbial fuel cell[133].
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345 Moreover, microbial electrolysis cell reactor configuration, which is closely relevant to hydrogen
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346 production rate, has made great progress in both the traditional type (i.e., incorporated membranes) and
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347 membrane-free designs. More details about reactor configuration were reviewed elsewhere[134]. In microbial
12
348 electrolysis cell, except exoelectrogens, there also exists methanogens which would consume hydrogen, so
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349 inhibiting the growth and activity of methanogens such as adding methanogen inhibitors [135], using low pH
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350 [136] and temperature [137] are helpful to improve hydrogen yields. Chae et al. [135] reported that adding 2-
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351 bromoethanesulfonate (286 μM) in microbial electrolysis cell could decrease methanogenic electron losses from
21
352 36.4 ± 4.4 to 2.5 ± 0.3%, i.e., CH₄ yield reduced from 145.8 ± 17.4 μmol to 10.2 ± 1.2 μmol, at the same time,
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353 hydrogen efficiency would be improved from 56.1 ± 5.7 to 80.1 ± 6.5%. In addition, harvesting hydrogen
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354 rapidly (e.g., use more efficient materials) is also helpful to enhance hydrogen yields.
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355 **Table.1. Endeavors that have been done to improve hydrogen yields from WAS.**
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357 Similarly to dark-photo fermentation combination system, the dark fermentation- microbial electrolysis
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358 cell two stages system is also constructed to improve hydrogen yields. In this system, scientist have found that
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359 acetate has the best conversion performance in microbial electrolysis cell compared to other dark fermentation
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360 products, such as propionic and butyric, so the ethanol type fermentation could be a more proper pathway in
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361 dark fermentation if this two-stages system is taken[138].
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362 **5. Research gap and perspective**
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363 Although numerous efforts have been performed, hydrogen production from WAS is still far from
52
364 achieving at practical scales due to low hydrogen yield, high energy (chemical) input in WAS pretreatment
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365 and fermenter control, or cost-prohibitive materials in photo-bioreactors (or anodes). In view of the state of the
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366 art relate to hydrogen bioproduction from WAS, further research on the following perspectives is urgently
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367 needed:
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- 368 (1) Many pretreatments were developed to promote sludge disintegration, but the utilization of sludge
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369 organics was still low due to WAS contains lots of refractory organics. Thus, more efforts are required to
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370 improve the biodegradability of WAS.
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371 (2) In addition, the low activities and growth rate of hydrogen-production microorganism limited hydrogen
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372 generate rate and yield. More endeavors are essential to comprehensively study hydrogen-production
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373 microorganisms and improve their activities.
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374 (3) Moreover, although all methods were used to improve hydrogen production, the hydrogen yield from WAS
15
375 dark fermentation was still low due to the low energy conversion efficiency. Hence, more efforts are need
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376 to develop two-stage process (e.g., dark fermentation and MECs) to improve hydrogen recovery.
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21 While continuing improvements in those aspects can be expected with the ongoing researches, it seems some
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23 of the challenges such as unstable performances and relatively high ratio of energy (or capital) input to energy
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25 output might also remain, making hydrogen production from WAS competitively unfavorable. According to the
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27 analyses, one major reason for the current situation is that previous studies mainly focus on the advancement of
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29 hydrogen production technology alone and lack rational integration of some available technologies. For
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31 example, Guo et al.[139] compared different pretreatment methods (e.g., multi-enzyme, heat and microwave
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33 pretreatments) and tried to find the best pretreatment method (heating pretreatment achieved the maximum
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35 hydrogen yield of 15.3 ml H₂/g VSS) but ignored other factors, such as fermenter control and WAS
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37 characteristic. Zhao et al.[93] investigated the effect of fermentation pH on hydrogen production and found the
38
39 optimal constant pH of 10 (In this scenario, the maximal H₂ yield of 26.9 mL H₂/ g VSS could be achieved).
40
41 However, other potential strategies were not considered as well. To further enhance hydrogen
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43 yield in a sustainable way, more appropriate strategies other than advancing hydrogen production technology
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45 alone should be therefore sought. Rational integration of the technologies available in both wastewater
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47 treatment and WAS treatment might be a more feasible avenue.
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52 53 391 **6. A future paradigm to improve hydrogen production** 54 55

56 392 **6.1. The hybrid process for the operation of WWTPs** 57 58

59 It is widely acceptable that WWTPs are not only the places to purify sewage but also the facilities to
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394 recovery energy and resource. The yield of each useful product (e.g., hydrogen) from WWTPs should be
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395 maximized, and meanwhile the operation input should be minimized as far as possible. In addition, the
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396 investment of WWTPs to recovery resource should be controlled at a reasonable level. Based on these
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397 principles, the combining dark fermentation technology with other available technologies including high-rate
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398 activated sludge process, mainstream deammonification, phosphate recovery, free ammonia (FA)-based sludge
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399 pretreatment and microbial electrolysis cells partially or completely might be a solution to accomplish
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400 desirable hydrogen production in sustainable ways. The schematic process of such combined strategy
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401 proposed is shown in Figure 3.
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**Figure.3. The conceptual operation of WWTPs with hybrid technologies for hydrogen and other energy
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In such WWTPs, wastewater is proposed to be cleaned by the adsorption-biodegradation (A/B) process,
which is widely accepted as the most promising process that could achieve desirable nutrient removal and
maximum energy recovery concurrently. Generally, most organic carbon is first adsorbed in the A-stage while
the effluent with a low C/N ratio is further biodegraded in B-stage. In this hybrid system, wastewater is first fed
to by a high-rate activated sludge reactor (called A-stage), where most organics are captured through various
mechanisms such as bio-sorption and storage. Up to now, many methods including chemical, biological, or their
combinations, have been explored for capturing organics at A-stage as much as possible [142, 143]. It is
documented that >80% of organic carbons and >90% of phosphorus would be removed from a high-rate reactor
through controlling solids retention times at 2.0-2.5 days and hydraulic retention times at
0.5-1 day [142]. Then, the effluent from the A-stage, containing low organic matters (i.e., low COD) but high
ammonium, is further treated by mainstream deammonification (called B-stage) [144], while the captured
organics are separated and passed onto hydrogen production units such as anaerobic fermenter for energy
recovery. Compared with the conventional WAS, A-stage sludge is easier to be disrupted [142], which is
beneficial to the subsequent hydrogen production.

To provide more soluble substrates for hydrogen production, pertinent pretreatment of sludge prior to
hydrogen production is usually performed in the sludge treatment line. Although many pretreatment

421 methods were tested, most of them require considerable input of either energy or chemicals, as mentioned
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422 above. This is unsustainable in long-term operation and largely limits the applications in field situations.
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423 Here, FA-based sludge pretreatment may be a solution for sustainable operation. FA, which could be
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424 obtained from anaerobic fermenter effluent directly, is a waste-generated, renewable chemical that can be
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425 produced in situ in WWTPs as a byproduct of sludge treatment (Figure 3). Recent investigations
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426 demonstrated that FA not only facilitated the lysis of sludge cells but also promoted the biodegradability of
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427 organics released[145, 146]. Although FA is reported to cause a strong biocidal impact or inhibition on
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428 broad microbes, its inhibitions to hydrogen consumers were much severer than those to hydrolytic
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429 microorganisms and hydrogen producers, which benefited hydrogen accumulation [147].
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430 After FA pretreatment, the pretreated sludge is suggested to be fed to anaerobic fermenter for the production
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431 of hydrogen and volatile fatty acids. Wang et al. reported that with an increase of FA from 34 to 254 mg/L, the
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432 maximum hydrogen yield from dark fermentation increased from 7.3 to 15.6 mL/g VSS without any addition
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433 of inoculum [147]. Zhang et al. demonstrated that when WAS was pretreated with
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434 237.8 mg/L FA for 3 d, the organics released were largely enhanced, with the soluble COD of 3400 ± 120 mg/L
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435 and volatile fatty acids of 226.9 mg COD/g VSS being 4.5-fold and 2.7-fold that without FA pretreatment,
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436 respectively[148]. Then the fermented WAS is further fed to microbial electrolysis cell unit for secondary
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437 hydrogen production. In such processes, the accumulation stage of soluble organic matters and hydrogen
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438 production stage is separated, which is beneficial for microbial electrolysis cell to generate hydrogen. Although
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439 many components in WAS could use as substrates for hydrogen production in microbial electrolysis cells,
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42
440 volatile fatty acids are the preferred substrates that could achieve higher efficiency and
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441 hydrogen recovery [141]. In addition, the combination of dark fermentation with microbial electrolysis cell
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442 could also achieve cascade utilization of complex organics from WAS through the syntrophic interactions
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443 between fermentative bacteria and exoelectrogens. He et al. confirmed that the positive effects of such
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444 combination could enhance hydrogen production, as compared to microbial electrolysis cell alone[23].
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445 And last, the effluent of dark fermentation-microbial electrolysis cell usually contains 200-600 mg/L
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446 ammonium and 100-300 mg/L phosphorus, which are mainly released from the disintegration of sludge cells
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447 [149, 150]. The phosphorus released can be recovered via either crystallization or precipitant as valuable
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448 products. Phosphorus recovery from WAS, an alternative and renewable resource, is very significant, because
449 the world's main P resource, i.e., phosphate rock, is non-renewable and may exhaust in next 100-250 years
450 [149, 151]. Thus, this could not only fulfill world demand for phosphorus but also compensate for a part of
451 WWTPs costs. In addition, phosphorus recovery from the fermentation liquid could further reduce its loading
452 in mainstream wastewater treatment, which thereby benefits to acquire the cleaner effluent. The effluent of P
453 recovery basin, which contains high levels of ammonium, is partially sent to the sludge pretreatment unit for
454 free ammonia production. By adjusting proper pH and temperature, the desirable free ammonia concentration
455 can be attained easily.

456 **6.2. Benefits of the hybrid process**

457 With a reasonable integration of various available technologies in WWTPs, it could achieve substantial
458 technical, environmental and economic benefits. The hybrid technologies could use a minimal energy input to
459 address some key questions occurred in current WWTPs, in particular, some key challenges faced by hydrogen
460 production from WAS. Firstly, the A-stage could capture much organic matters to improve WAS characteristics.
461 Generally, the destruction of WAS in anaerobic digester is around 35% whereas the corresponding value of A-
462 stage sludge could achieve at 70% [152]. Secondly, FA pretreatment could efficiently break the rigid structure
463 of WAS microbial cell to release more extracellular and intracellular organics for hydrogen production, which
464 is usually thought to be the rate-limiting step. In addition, FA pretreatment could efficiently inhibit hydrogen
465 consumption, i.e., homoacetogenesis, methanogenesis and sulfate-reducing processes. Microbial electrolysis
466 cell could further utilize most of the organics (e.g., volatile fatty acids, proteins and carbohydrates) left in the
467 liquor of anaerobic fermenter to produce hydrogen. All
468 these aspects may enable hydrogen yield from WAS at desirable levels.

469 This hybrid technologies could also achieve considerable environmental benefits. WAS adsorbs and
470 concentrates substantial exogenous pollutants such as heavy metal and micro-pollutants [153]. The improper
471 disposal of sludge would make these toxic and harmful contaminants reenter natural environments, posing
472 the serious threat to the ecological environment. Substantial reduction and reuse of WAS through this hybrid
473 technologies is conducive to the final disposal of sludge and avoids the secondary pollution of the
474 environment. In addition, the extensively use of hydrogen rather than methane as energy reduces carbon

dioxide production, thereby slowing down the greenhouse effect. Moreover, the recovery of P-based fertilizers from WAS reduces the exploitation of phosphate deposits, avoiding environmental damage [15].

In addition, such operational paradigm also has enormous economic benefits. For example, mainstream deammonification could save the extra addition of carbon source which are usually applied in conventional WWTPs to enhance nutrient removal [140]. Free ammonia is renewable and waste-produced, which could be produced *in situ* from sludge fermenter effluent. In comparison, other pretreatments require high chemicals or energy input [147]. Combination of dark fermentation with microbial electrolysis cell could improve hydrogen yields largely from WAS, i.e., recover more energy from sludge and create more values for WWTPs [147]. Moreover, the P-based fertilizers from the effluent of dark fermentation-microbial electrolysis cell through either crystallization or precipitant could enhance the income of WWTPs and offset part of the cost of wastewater treatment [15]. And last, such operation could also enhance sludge reduction, thus decreasing the cost of sludge transport and disposal [154].

Here, the economic benefit of this hybrid strategy with taking a 500 000 population equivalent (i.e., 100 000 cubic meters per day) WWTP was estimated as an example (please see Table S1 for details, Supporting information). In China, a conventional WWTP is often operated without the process of sludge anaerobic digestion, which account for >80% of total WWTPs. In such a WWTP, wastewater treatment would consume \$15000~\$20000 per day (i.e., \$5475000~\$7300000 per year), depending on the organic carbon, nitrogen, and phosphorus available in the wastewater. In addition the disposal of excess sludge would consume about \$10000 per day (i.e., \$3650000 per year). By incorporating these hybrid technologies into the conventional WWTPs, a same size WWTP could save the expense of using extra organic carbon (i.e., about \$547500),

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\$2254000), and also produce more $\sim 2.64 \times 10^5$ kg of hydrogen per year. It is estimated that this combined strategy would save \sim \$3.83 million in a WWTP with a capacity of 10 m³/d per year, as compared with the traditional operation paradigm (without anaerobic digestion process). Considering thousands of the conventional WWTPs existing in the world, this hybrid strategy should have huge economic benefits. However, it should be noted that the technical and economic merits presented here are only indicative, which need to be further clarified at real-world situations in the future.

485 recover about 2.26×10 tons of P-based fertilizers, reduce the portion cost of sludge disposal (i.e., about
475 dioxide production, thereby slowing down the greenhouse effect. Moreover, the recovery of P-based fertilizers
476 from WAS reduces the exploitation of phosphate deposits, avoiding environmental damage [15].

477 In addition, such operational paradigm also has enormous economic benefits. For example, mainstream
478 deammonification could save the extra addition of carbon source which are usually applied in conventional
479 WWTPs to enhance nutrient removal [140]. Free ammonia is renewable and waste-produced, which could be
480 produced *in situ* from sludge fermenter effluent. In comparison, other pretreatments require high chemicals or
481 energy input [147]. Combination of dark fermentation with microbial electrolysis cell could improve hydrogen

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486 \$2254000), and also produce more $\sim 2.64 \times 10^5$ kg of hydrogen per year. It is estimated that this combined
49 strategy would save \sim \$3.83 million in a WWTP with a capacity of 10 m³/d per year, as compared with the
497 traditional operation paradigm (without anaerobic digestion process). Considering thousands of the
498 conventional WWTPs existing in the world, this hybrid strategy should have huge economic benefits.
499 However, it should be noted that the technical and economic merits presented here are only indicative, which
500 need to be further clarified at real-world situations in the future.
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6.3. Challenges of the hybrid process and future directions

Although many endeavors have been done, microbial electrolysis cell has not yet achieved practical application, which is mainly tested at lab-scale. Moreover, some other individual technologies of this hybrid system (e.g., phosphate removal based-high rate activated sludge process) are also not applied in real-world. Hence, scaling up this hybrid system would be therefore inevitable. This requires not only enhancing the size and treatment capacity of this hybrid system to a full-scale level, but also enabling desirable energy recovery. Thus, the biggest challenge for a practical situation is how to scale up the system size and guarantee the performances of each individual simultaneously.

Microbial electrolysis cells, which play important roles in this hybrid system, face some specific limitations in full-scale application. First, in a full-scale reactor, long distance between electrodes and pH gradient would lead to high internal resistances. Second, high capital costs of expensive electrodes and membrane materials also limit its application. Third, there are still lacking comprehensive understanding of microbiology involved in microbial electrolysis cells including fermentative bacteria, exoelectrogens and biocathode microorganisms. By further designing microbial electrolysis cell reactors, developing new materials, deeply understanding and taking control over microbiology are expected to address these challenges.

Apart from improving the performance of individual units, rational integration of individual technologies is also important for achieving full-scale application of such hybrid systems. Mathematical modeling could be useful to understand, design, and operate such complex hybrid system. In addition, on-line monitoring and real-time process control is indispensable in practice. These monitoring parameters such as ammonium and nitrite concentrations in the nitrification reactor, FA concentration in pretreatment unit, and electrode potential in microbial electrolysis cells could use as valuable indicators to manipulate the whole system, ensuring the stability and robustness of such hybrid system.

Finally, it's crucial to acquire government support to develop such emerging technologies. The ignored environmental costs and emerging technologies, which have not yet benefited from application scale, lead to the expensive cost of recover energy and resource from WWTPs, as compared with the use of fossil-fuel energy. More investment in WAS treatment and disposal is required, particularly in the developing world. For

529 example, in China, the investment in WAS treatment and disposal (5.6 billion USD/year) was much less than
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530 that in wastewater treatment (68.8 billion USD/year) [155]. Government must try to put the emerging
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531 technologies, which could save energy even produce energy, into priority position through improvement of
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532 existing regulatory frameworks to contain or enhance the costs of greenhouse gas (e.g., carbon dioxide)
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533 emission and waste treatment and disposal. Government and relevant enterprises should also offer enough
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534 funds and suitable infrastructure to develop these emerging technologies available in real-world situations.
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535 7. Conclusions

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536 In this paper, the principles and potentials, microorganisms, possible technologies, and process parameters of
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537 hydrogen production were reviewed. Besides, both the opportunities and challenges of hydrogen production were
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538 analyzed. Microbial electrolysis cells were regarded as the prospective technology for hydrogen production due to
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539 high theoretical hydrogen yield and the ability of using varieties of organics as substrate. Nonetheless, the low
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540 organics utilization from WAS and the quick depletion of generated H₂ limited hydrogen accumulation. Based on
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541 systematic summarization and critical analysis, a possible paradigm for hydrogen production and wastewater
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542 treatment by rationally integrating dark fermentation with several other treatment technologies, including MEC, high-
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543 rate activated sludge process, mainstream deammonification, phosphate recovery, are presented. This hybrid process
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544 could achieve substantial technical, environmental and economic benefits. In addition, the limitations and future
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545 directions for application of such hybrid processes were discussed. More efforts are required to achieve the
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546 application of this hybrid process in real-world.
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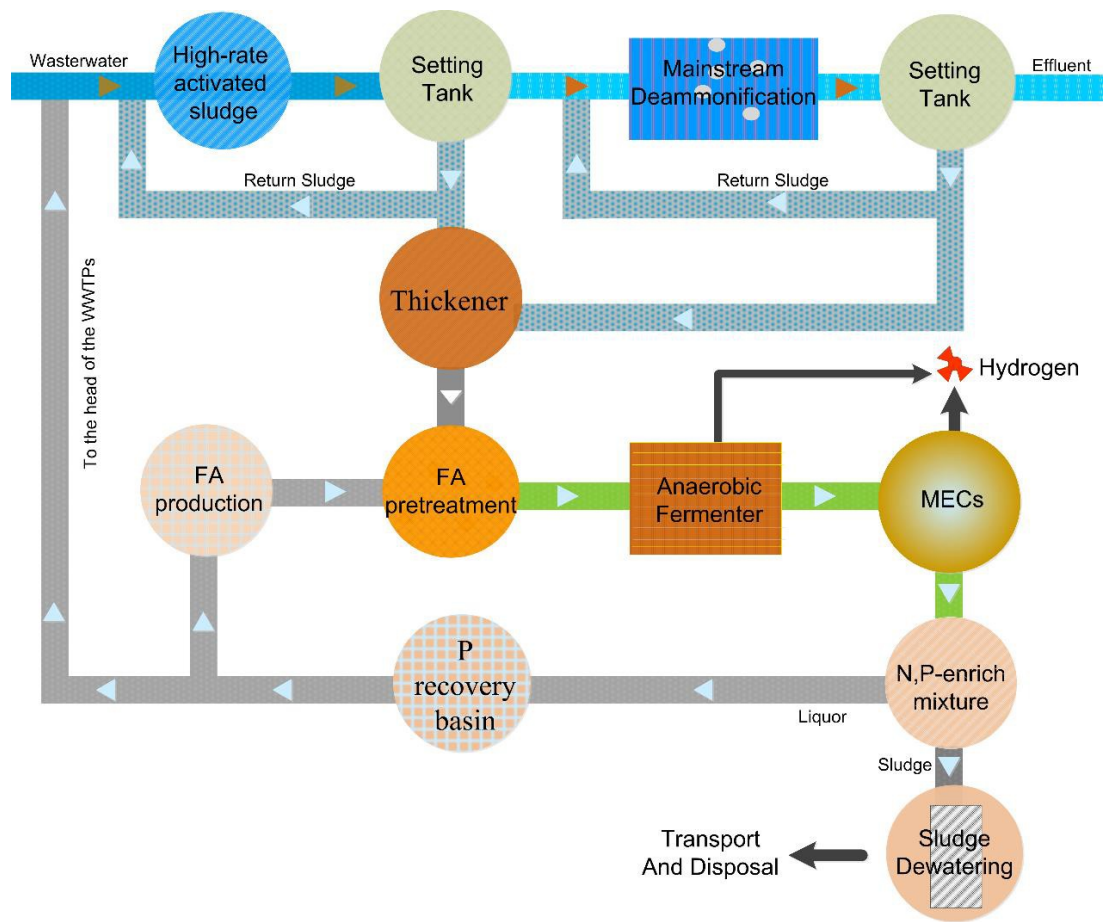
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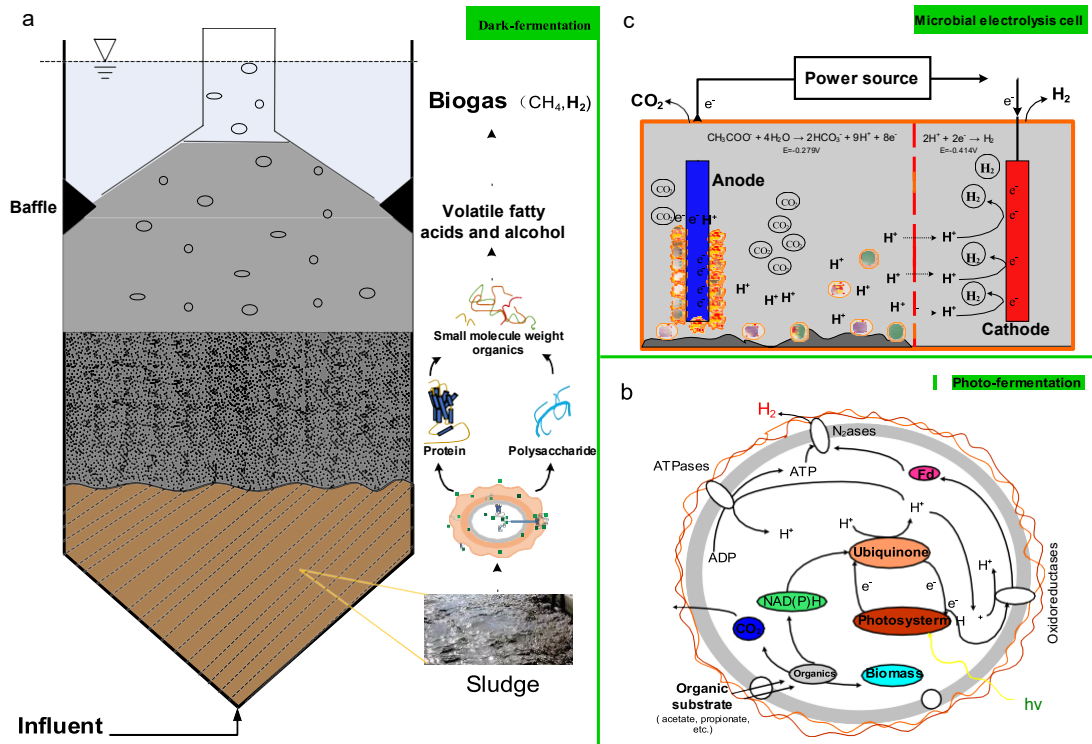


Figure. 1. Schematic diagram of hydrogen production from dark-fermentation (a), photo-fermentation (b) and microbial electrolysis cells (c).

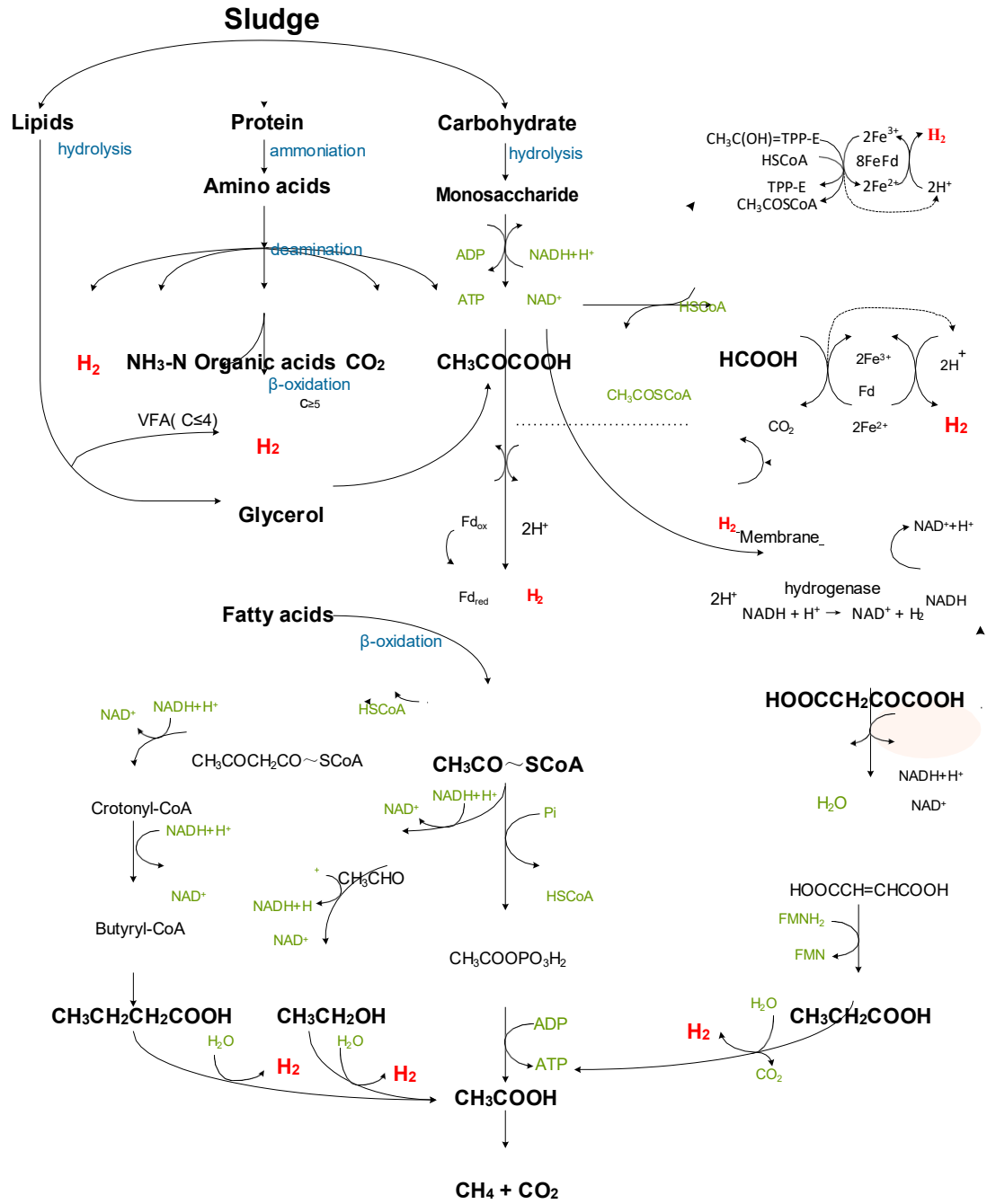


Figure. 2. The pathways of hydrogen production from sludge dark fermentation

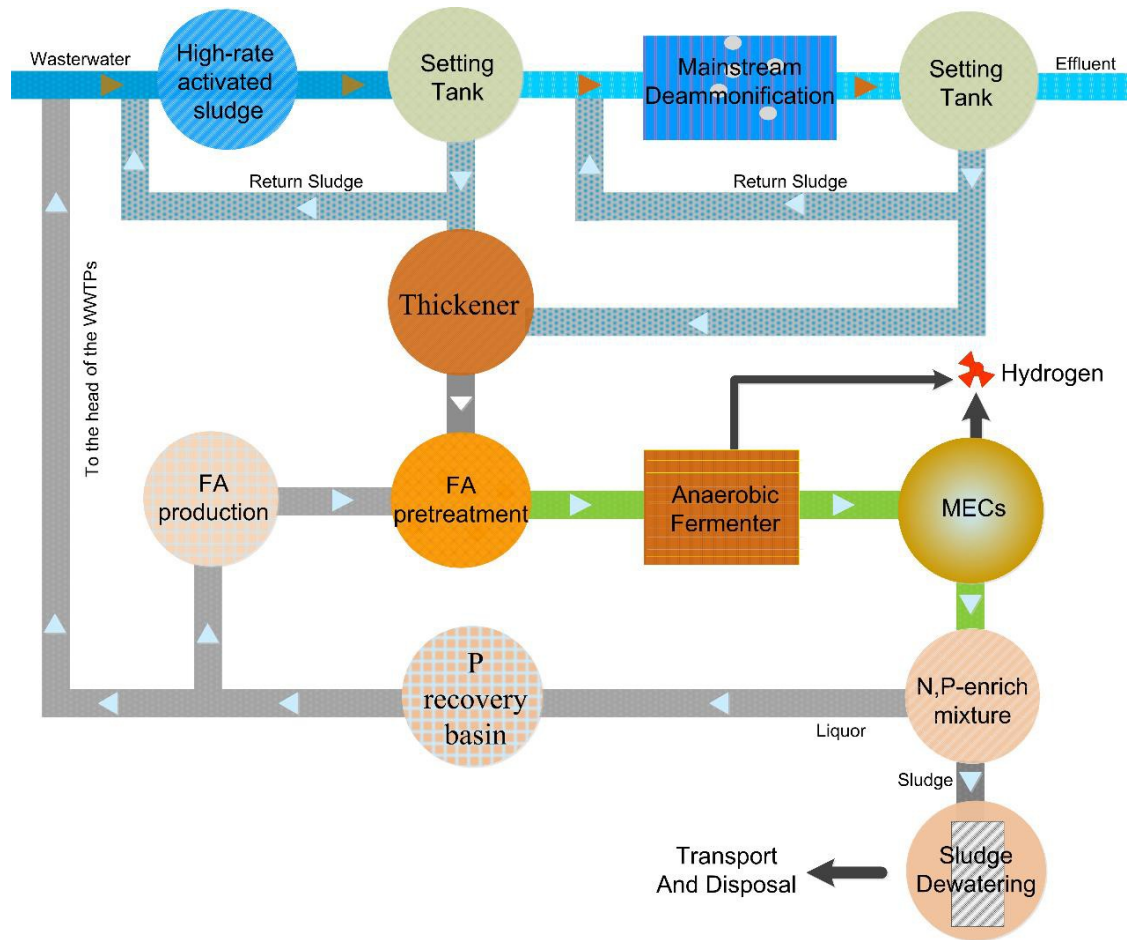


Figure. 3. The conceptual operation of WWTPs with hybrid technologies for hydrogen and other energy recovery.

Table 1. Endeavors that have been done to improve hydrogen yields from WAS

| Type of H ₂ production | Regulating strategy | Brief description of operation | Result | Reference |
|-----------------------------------|--------------------------|---|---|---------------|
| Dark fermentation | Fermenter operation | Alkaline fermentation | Facilitated the disruption of WAS and suppressed the activities of hydrogen consuming microorganisms | [11, 93] |
| Dark fermentation | Fermenter operation | High fermentative temperature | Facilitated the hydrolysis of substrates and simplified microbial diversity | [95] |
| Dark fermentation | Fermenter operation | Shorter hydraulic retention time | Wash out H ₂ -consuming bacteria (e.g., propionic acid bacteria) | [94] |
| Dark fermentation | WAS characteristics | Increased PHA content | Enhanced the cell disintegration, the hydrolysis of organics, and the soluble protein conversion of non-PHA biomass | [22, 96, 98] |
| Dark fermentation | WAS characteristics | Co-fermenting WAS with other carbon-rich organics | Pertinent increases of carbon/nitrogen ratios was helpful to the conversion of protein | [97, 99, 100] |
| Dark fermentation | Pretreatment methods | Thermal pretreatment | Accelerated the disruption of floc structure and microbial cells. | [87, 91] |
| Dark fermentation | Pretreatment methods | Ultrasonic pretreatments | Accelerated sludge solubilization, and enhanced the release of biodegradable organics, | [89, 91] |
| Dark fermentation | Pretreatment methods | Alkaline pretreatment | Enhanced sludge disintegration and inhibited the activity of methanogens | [6] |
| Dark fermentation | Pretreatment methods | Free nitrite acid pretreatment | Promoted the biodegradability of organics released and inhibited activities of hydrogen consumers | [7, 90] |
| Dark fermentation | H ₂ -producer | Metabolic engineering | Modify hydrogen-producers in fermenters and enhanced hydrogen production by 3.5 fold | [102, 103] |
| Dark fermentation | H ₂ -producer | Constructed a microbial consortium | inoculating with three established bacteria and improved hydrogen production by 1.5-4 times | [101] |
| Photo fermentation | Fermenter operation | Temperature optimization | The suitable temperature ranges of photo-fermentative hydrogen generation were 31-36 °C | [118] |

| | | | | |
|-----------------------------|--------------------------|--|---|------------|
| Photo fermentation | Fermenter operation | High transparent materials | Glass and polymethyl methacrylate are considered the suitable materials for constructing photo-bioreactor | [74] |
| Photo fermentation | Fermenter operation | pH optimization | The suitable pH ranges of photo-fermentative hydrogen generation were 6.8-7.5 | [118] |
| Photo fermentation | Light | Light wavelengths and intensities | The suitable light wavelength and intensity were in the range of 400-1000 nm and 6-10 klux, respectively. | [121] |
| Photo fermentation | H ₂ -producer | Strain improvement and replacement of nitrogenase with hydrogenase | Strains with a 41%-49% decrease in LH2 (LH: light-harvesting) content by UV irradiation produced 50% more hydrogen than untreated strains. | [119, 120] |
| Microbial electrolysis cell | Electrode improvement | Anodic material selection | Many new, cheap and effective anode materials (e.g., carbon nanotubes and graphene) were found | [125, 126] |
| Microbial electrolysis cell | Electrode improvement | Cathode material selection | many new, cheap and effective cathode materials (e.g., carbon nanotube and bio-cathode) were developed | [127, 128] |
| Microbial electrolysis cell | Electrode improvement | Anode modification | Many treatments such as heat treatment and acid treatment on anode materials surface have been used to improve the properties of anode surface (e.g., electron transfer). | [132, 133] |
| Microbial electrolysis cell | Reactor configurations | Incorporated membranes | Reduced the possibility of hydrogen diffusing to the anode to be used by electrogenic or methanogenic bacteria, and made the collected hydrogen more pure | [134] |
| Microbial electrolysis cell | Reactor configurations | Membrane-free designs | Lead to both high hydrogen recoveries and production rates | [134] |
| Microbial electrolysis cell | Microbial manipulation | Adding methanogen inhibitors | Added 2-bromoethanesulfonate in MEC enhancing H ₂ production efficiency from 56.1± 5.7 to 80.1± 6.5%. | [135] |