Determining SNPs and demographic variables that impact level one fingerprint pattern

by Andrew Walton

Thesis submitted in fulfilment of the requirements for the degree of

Doctor of Philosophy

under the supervision of Dr Sebastien Moret, Dr Dennis McNevin and Dr Mark Barash

University of Technology Sydney Faculty of Science

April 2022

Certificate of original authorship

Certificate of original authorship

I, Andrew Walton declare that this thesis, is submitted in fulfilment of the requirements for the

award of Doctor of Philosophy, in the Faculty of Science at the University of Technology

Sydney.

This thesis is wholly my own work unless otherwise referenced or acknowledged. In addition, I

certify that all information sources and literature used are indicated in the thesis.

This document has not been submitted for qualifications at any other academic institution.

This research is supported by the Australian Government Research Training Program.

Signature:

Production Note:

Signature removed prior to publication.

Date: 2/4/2022

ii

Abstract

The current forensic use of fingerprints is for identification purposes and requires a reference sample for comparison to any unknown fingermark. Features of individual ridges can be used for identification however in combination they create overall level one fingerprint patterns (arches, loops and whorls). Past studies have indicated that some fingerprint characteristics such as level one pattern may occur at higher frequencies in some biogeographical ancestries (BGAs) and ridge density (the number of ridges within a defined area) may be used to determine the sex of an individual. The frequency at which these patterns and their subclassifications occur is largely unknown in the Australian population as there have been no modern studies utilising statistical analysis. Fingerprint experts would benefit from the publication of this information as it is the first step in building a statistical model that may add probabilities to their opinions on pattern rarity in a court setting. Previously, they may only rely upon their own observations from their experience and studies based on overseas populations.

This research aimed to represent the level one fingerprint pattern and ridge density frequencies of the diverse Australian population. This also provided the opportunity to assess the association of pattern and ridge density with BGA, sex, hands, fingers, and genetic markers. By assessing these associations, a new avenue of investigative potential could be unlocked from fingerprint evidence. For fingermarks that do not return a match it may be possible to predict which hand or finger the mark came from (provided it is not a full set), the ancestry, sex, or genotype of the depositor.

A total of 828 volunteers, 515 people from Sydney donated their fingerprints, DNA and self-declared BGA through a questionnaire and 313 people from Melbourne provided fingerprints with self-declared BGA information. The fingerprints in Sydney were collected via fingerprint scanner and those from Melbourne were provided as ink on card. The fingerprints were then classified using the National Crime Information Center (NCIC) classification system. Goodness of fit tests, multinomial logistic regression and general estimating equations were utilised for association of ancestry, sex, hands, and fingers with the pattern and ridge densities. Associations between fingerprint patterns and genetic markers were investigated for five genetic models.

The goodness of fit and multinomial logistic regression analyses revealed several patterns occurring at significantly higher and lower frequencies than expected for all independent variables. The general estimating equations also showed significant differences amongst ridge densities (radial, ulnar, and proximal positions) for all independent variables. A further

investigation was made into the useability of ridge density in fingerprints of unknown finger or hand. Results showed that proximal and ulnar positions produced dissimilar results to right and left positions, indicating this characteristic would be limited in its usefulness in forensic casework.

In people of European and Middle Eastern biogeographical ancestry over 60 SNPs were significantly associated with fingerprint patterns and ridge densities and four genetic loci were amongst the hundreds of genetic markers that were not quite significant. The four loci included two distinct areas on chromosome six, an area on chromosome one and an area on chromosome 11. Several genetic markers were novel, and several replicated those found in previous studies. The hypothesis that non-coding regions and epigenetic regulation are causative of fingerprint development was tentatively supported. These results provide strong evidence that frequencies of level one fingerprint pattern and ridge density differ between ancestral populations and sex and occur with different frequencies amongst fingers and between hands. Genetic markers may be identified in the future with diverse and increased sample sizes and through DNA phenotyping the prediction of an individual's fingerprints may be possible, allowing the interrogation of a fingerprint database even if there are no physical fingerprints.

This research met the original aims to assess the association of pattern and ridge density with BGA, sex, hands, fingers, and genetic markers in the diverse Australian population. Many results were novel and created potential leads for future investigation. The use of this research by practitioners however would be premature as larger BGA groups are needed for both pattern to BGA and pattern to SNP association studies.

Acknowledgements

Acknowledgements
To,
The volunteers
My supervisors
My friends
My family
The University of Technology Sydney
New South Wales Police Force
Victoria Police
The Department of Home Affairs
Thank you for all the support, without you this research would not have been completed over the last five years.

Table of contents

Ce	ertificat	te of original authorship	ii
A	bstract		iii
Α	cknowl	edgements	v
Tá	able of	contents	vi
Te	erms ar	nd abbreviations	x
Li	st of fig	ures	xii
Li	st of ta	bles	xv
Re	esearch	communication	xviii
	Publica	ations	xviii
	Preser	itations	xviii
1.	Intr	oduction to fingerprint development and DNA foundations	2
	1.1.	Foetal fingerprint development	2
	1.2.	Characteristics of fingerprints	4
	1.2.	1. Fingerprint patterns	4
	1.2	2. Methods of fingerprint classification	10
	1.2.	3. Investigational information available from fingerprints	11
	1.3.	The uniqueness and evidential weight of fingerprints	13
	1.4.	Fingerprint genetics	15
	1.4.	1. Inheritance of fingerprints	16
	1.4	2. Genetics of fingerprint pattern development in humans	18
	1.4.	3. Syndromes with known fingerprint pattern effects	20
	1.4.	4. Fingerprints from an evolutionary standpoint	22
	1.5.	Approach for targeting genes with influence on fingerprint phenotype	23
	1.5.	1. Fingerprint candidate SNPs and selection criteria	24
	1.5	2. Forensic application of STRs, microhaplotypes and mitochondrial DNA	27
	1.5	3. Forensically relevant SNP classes	28
	1.5.	4. Web-based bioinformatical resources for SNP selection	29
	1.5	5. SNP chip platforms for targeted SNP genotyping	30
	1.6.	Forensic Intelligence	32
	1.6.	1. Locard's exchange principle	33
	1.6	2. DNA phenotyping	33
	1.6.	3. Phenotyping provides avenues for investigation	35
	1.6.	4. Eyewitness unreliability	37
	1.7.	Project hypotheses and aims	38

2.	Mat	erials and methods	. 41
	2.1.	Ethics approval	. 41
	2.2.	Samples	. 41
	2.3.	DNA extraction	. 42
	2.4.	DNA quantification	. 42
	2.5.	Candidate genes and SNPs using bioinformatics resources and previous literature	. 43
	2.6.	SNP chip selection and DNA genotyping	. 44
	2.7.	Fingerprint classification	. 46
	2.8.	Statistical analysis	. 46
3.	Rari	ty of fingerprint pattern and its association with BGA and sex	. 48
	3.1.	Introduction	. 48
	3.2.	Volunteers by categories	. 49
	3.3.	Frequency distribution of pattern	. 51
	3.4.	Pattern distribution on the fingers	. 55
	3.5.	Association of pattern using chi-squared (χ^2) with post-hoc analysis	. 57
	3.6.	Association of pattern using multinomial logistic regression (MLR)	. 61
	3.7.	Summarising the multinomial logistic regression results	. 76
4.	Ridg	e density association with BGA and sex	. 81
	4.1.	Introduction	. 82
	4.2.	The effect of pressure on digital fingerprint scans	. 84
	4.3.	The semi-automated ridge count method	. 85
	4.4.	Ridge density results and discussion	. 87
	4.4.	1. Mode, median and mean for sex and BGA ridge density per position	. 87
	4.4	2. General estimating equation analysis of ridge density	. 92
	4.4	, , , , , , , , , , , , , , , , , , , ,	
		parison	
	4.4.	 Ridge density as radial and ulnar positions compared to left and right position 101 	15
	4.5.	Summarising the general estimating equation results	103
5.	Inhe	ritance of fingerprint patterns – A case study	106
	5.1.	Introduction	106
	5.2.	Classification	109
	5.3.	Family structure and demography	109
	5.4.	Pattern frequency, ridge count and ridge tracing	111
	5.5.	Pattern inheritance	116
6.	SNP	association with fingerprint patterns	125

	6.1.	Introduction	125
	6.2.	R packages – SNPassoc and qqman	126
	6.3.	Descriptive analysis	127
	6.3.1	l. Missing SNP data	127
	6.3.2	2. Hardy-Weinberg Equilibrium	128
	6.4.	Association of pattern	129
	6.4.1	I. Multiple finger phenotypes	131
	6.4.2	2. Single finger phenotypes	137
	6.4.3	3. Summary	165
7.	Cond	clusions and future directions	169
	7.1.	Conclusions	169
	7.1.1	Pattern association with BGA and sex	169
	7.1.2	2. Ridge density links with BGA and sex	172
	7.1.3	3. Inheritance of fingerprint patterns	174
	7.1.4	1. SNPs linked to fingerprint pattern	175
	7.2.	Future directions	179
Αį	pendic	es	184
	Append	A xib	184
	Henr	ry classification system	184
	Append	dix B	186
	Vuce	tich classification system	186
	Append	dix C	188
	State	ement of consent presented to each volunteer	188
	Append	dix D	189
	UTS	questionnaire for self-reported ancestry and phenotypic traits	189
	Append	dix E	190
	MLR	results for arches, loops, and whorls	190
	Append	dix F	194
	Wolf	fram Mathematica code for counting ridges	194
	Append	dix G	195
	Ridg	e density GEE results for the ulnar, radial, and proximal positions	195
	Append	H xib	199
	Ridg	e density GEE results for left and right finger position	199
	Append	I xib	202
	R co	de – SNPassoc and qqman packages	202
	Annend	dix I	206

References	322
Manhattan plots for each variable and model	224
Appendix K	224
A list of all significant and suggestive SNPs	206

Terms and abbreviations

BGA Biogeographical Ancestry

SNP Single Nucleotide Polymorphism

USA United States of America

NCIC National Crime Information Center

DNA Deoxyribonucleic Acid

EVC Externally Visible Characteristic

NCIDD National Criminal Investigation DNA Database

ARC Absolute Ridge Count

LOD Logarithm Of Differentiation

QTL Quantitative Trait Locus

ROES Ridges Off the End Syndrome

PCA Principal Component Analysis

FISWG Facial Identification Scientific Working Group

FBI United States Federal Bureau of Investigation

MLR Multinomial Logistic Regression

GLM Generalised Linear Model

GEE General Estimating Equation

qPCR Quantitative Polymerase Chain Reaction

HWE Hardy-Weinberg Equilibrium

AIC Akaike Information Criterion

CHR Chromosome

BP Base Pair

LL Left Little finger

RL Right Little finger

LR Left Ring finger

RR Right Ring finger

LM Left Middle finger

RM Right Middle finger

LI Left Index finger

RI Right Index finger

LT Left Thumb finger

RT Right Thumb finger

EMT Epithelial Mesenchymal Transition

List of figures

Figure 1: Growth of the hand between weeks five and eight of gestation	2
Figure 2: Radiating development of friction ridge pattern	2
Figure 3: Pore units containing individual sweat glands create the epidermal ridges	4
Figure 4: Three general patterns of level one fingerprint	5
Figure 5: An example of an ulnar loop from the ring finger on the left hand	6
Figure 6: An example of a radial loop from the index finger on the left hand	6
Figure 7: An example of a plain whorl on the right thumb	7
Figure 8: An example of a central pocket loop whorl on the right middle finger	7
Figure 9: An example of a double loop whorl on the ring finger of the right hand	7
Figure 10: An example of an accidental loop whorl on the left thumb	7
Figure 11: An example of a plain arch located on the left thumb	8
Figure 12: An example of a tented arch located on the left middle finger	8
Figure 13: Bifurcations and ridge endings – two basic types of minutiae	8
Figure 14: An example of the method used to calculate ridge density	12
Figure 15: An example of the main symptoms of Ridges Off the End Syndrome (ROES)	21
Figure 16: Similarities of fingerprints between species	23
Figure 17: Types of SNPs based on their location and effect	25
Figure 18: Diagram of the oligonucleotide SNP array principle	31
Figure 19: Diagram of the BeadArray principle	32
Figure 20: Biogeographical ancestry of volunteers (N=515) from Sydney	49
Figure 21: Biogeographical ancestry of volunteers (N=316) from Melbourne	49
Figure 22: Combined biogeographical ancestry of volunteers (N=831) combined	49
Figure 23: Sex of volunteers (N=515) from Sydney	50
Figure 24: Sex of volunteers (N=318) from Melbourne	50
Figure 25: Combined sex of volunteers (N=833) combined	50
Figure 26: Occurrence of fingerprint patterns per finger	55
Figure 27: Summarised results of the MLR per pattern and BGA	76
Figure 28: Summarised results of the MLR per pattern and sex	77
Figure 29: Summarised results of the MLR per pattern and hand	77
Figure 30: Summarised results of the MLR per pattern and finger	78
Figure 31: A demonstration in the concept of ridge count	81
Figure 32: Location of ridge density box and direction of counting for the assessment of	
deposition pressure effect	84

Figure 33: An example of (A) radial, (B) ulnar and (C) proximal positioning of ridge densi	ity
boxes (25mm ²) with direction of counting indication by dashed lines	86
Figure 34: A demonstration in the variation of ridge density in the proximal position pe	r
pattern	97
Figure 35: A diagram of the relationship between individuals fingerprinted for the inher	ritance
case study	110
Figure 36: Distribution of level one fingerprint pattern within the two linked extended f	amilies
	111
Figure 37: The percentage frequency of patterns in the extended family and the combin	ned
European BGA from Sydney and Melbourne	112
Figure 38: Ridge count frequency of the family sample	113
Figure 39: Ridge count frequency of the Sydney population	114
Figure 40: Family breakdown of ulnar loop ridge count	114
Figure 41: Examples of inner, meeting, and outer whorls on fingers of the left hand	115
Figure 42: The frequency and locations of whorl pattern subclassifications in the family	group
	115
Figure 43: Frequency and location of whorl pattern subclassifications in the Sydney pop	oulation
	116
Figure 44: The presence of a radial loop on the right index finger	117
Figure 45: The presence of a tented arch on the right index finger	119
Figure 46: A family pedigree displaying the average ridge count of each person	122
Figure 47: Missing SNP data from the genotyped samples	128
Figure 48: Log-additive model of SNP association to the radial loop on index fingers phe	notype
	132
Figure 49: Manhattan plot of SNPs for the "eight or more whorl" phenotype for the log	-
additive model	134
Figure 50: P-values suggesting association to phenotype of at least one arch pattern for	the
codominant model	135
Figure 51: A subsection of chromosome six p-values suggesting association to phenotype	oe of at
least one arch pattern for the codominant model	137
Figure 52: Manhattan plot of SNPs for arches on the left thumb for the log-additive mo	del . 138
Figure 53: Manhattan plot of SNPs for loops on left thumb for the dominant model	140
Figure 54: A subsection of chromosome 6 displaying an area of increased association fo	r loops
on the left thumb in the dominant model	141

Figure 55: Manhattan plot of SNPs for the loops of the right thumb phenotype for the
dominant model
Figure 56: A subsection of chromosome one showing areas of chromosome 1 displaying an
area of increased association for loops on the right thumb in the dominant model 143
Figure 57: Manhattan plot of SNPs for arches on the left index finger for the log-additive model
Figure 58: Manhattan plot of SNPs for whorls on the left index finger phenotype under the
dominant model
Figure 59: Manhattan plot of SNPs for whorls on the left middle finger for the dominant model
Figure 60: Manhattan plot of SNPs for loops on the right middle finger for the log-additive
model
Figure 61: A subsection of chromosome 11 for loops on the right middle finger in a log-additive
model
Figure 62: Manhattan plot of SNPs for loops on the left ring finger for the recessive model . 151
Figure 63: Manhattan plot of SNPs for whorls on the left ring finger for the overdominant
model
Figure 64: Manhattan plot of SNPs for arches on the left ring finger for the log-additive model
Figure 65: Manhattan plot of SNPs for arches on the right ring finger for the log-additive model
Figure 66: Manhattan plot of SNPs for arches on the right ring finger for the recessive model
Figure 67: Manhattan plot of SNPs for loops on the left little finger for the log-additive model
Figure 68: Manhattan plot of SNPs for whorls on the left little finger for the log-additive model
Figure 69: Manhattan plot of SNPs for arches on the left little finger for the log-additive model
Figure 70: Manhattan plot of SNPs for arches on the right little finger in the log-additive model
Figure 71: Manhattan plot of SNPs for arches on the right little finger in the recessive model
164

List of tables

Table 1: NCIC classification system	11
Table 2: Criteria for considered genotyping options	45
Table 3: Frequencies (%) of the three main fingerprint pattern groups in males and female	s . 51
Table 4: Frequencies (%) of the three main fingerprint pattern groups in four BGA groups .	52
Table 5: Frequencies (%) of fingerprint pattern groups in several studies within differing Bo	ЗАs
	53
Table 6: Frequencies of fingerprint pattern groups between males and females	54
Table 7: Frequencies of fingerprint pattern groups between four BGA groups	54
Table 8: Adjusted residuals of the chi-squared analysis for fingerprint pattern association v	vith
sex	58
Table 9: Adjusted residuals of the chi-squared analysis for fingerprint pattern association $oldsymbol{v}$	vith
BGA	58
Table 10: P-values calculated from each individual adjusted residual for sex against patteri	า 60
Table 11: P-values calculated from each individual adjusted residual for BGA against patte	rn 61
Table 12: Multinomial logistic regression significance (p-value) analysing the effect of hand	d,
finger sex and BGA on pattern	62
Table 13: Multiplier for presence of plain arch relative to ulnar loops compared to the thu	mbs
	63
Table 14: A summation of published results on the association of fingerprint patterns with	sex
	63
Table 15: Multiplier for presence of tented arch relative to ulnar loops compared to the	
thumbs	66
Table 16: Multiplier for presence of radial loops relative to ulnar loops compared to the	
thumbs	66
Table 17: Multiplier for presence of double loops whorls relative to ulnar loops compared	to
the thumbs	68
Table 18: A summation of published results on the asymmetrical frequency of fingerprint	
patterns	69
Table 19: Multiplier for presence of plain whorls relative to ulnar loops compared to the	
thumbs	70
Table 20: A summation of published results on the association of fingerprint patterns with	BGA
	75
Table 21: Ridge density of fingers when applying increasing pressure	85

Table 22: The number of observations removed based on quality due to inability to measur	e
ridge density	87
Table 23: Sex ridge density ulnar position	88
Table 24: Sex ridge density radial position	88
Table 25: Sex ridge density proximal position	89
Table 26: BGA ridge density ulnar position	89
Table 27: BGA ridge density radial position	90
Table 28: BGA ridge density proximal position	90
Table 29: A summary of the Bayesian probability outcomes for ridge density according to	
males and females	91
Table 30: Shapiro-Wilk test of normality with observations of skewness and kurtosis for the	!
Sydney data	100
Table 31: Shapiro-Wilk test of normality with observations of skewness and kurtosis for the	;
Melbourne dataset	100
Table 32: Pairwise comparisons of the Kruskal-Wallis test for ridge density positions for the	
Sydney dataset	100
Table 33: Pairwise comparisons of the Kruskal-Wallis test for ridge density positions for the	
Melbourne dataset	101
Table 34: A female's distinct set of mirrored fingerprints within the family sample	112
Table 35: An illustration of X-linked dominant inheritance	118
Table 36: Multiple linear regression results of correlation of average ridge density between	
children and parents	123
Table 37: Multiple linear regression results explaining the variation in children's ridge count	ts
	123
Table 38: Significant SNPs for the log-additive model of radial loops on the index fingers	131
Table 39: Genotypes and occurrence of the phenotype in rs41269369	132
Table 40: Distribution of the phenotype per genotype for rs1339062 in a dominant model	133
Table 41: Frequency of the phenotype per genotype in the recessive model for rs2443426 .	136
Table 42: Frequency of the phenotype per genotype in the recessive model for rs1447177 .	136
Table 43: Frequency of the phenotype per genotype for rs2891225 in the dominant model.	138
Table 44: Phenotype distribution per genotype for rs668502 for the dominant model	142
Table 45: Phenotype distribution per genotype for rs6871490 for the log-additive model	144
Table 46: Phenotype distribution per genotype for rs1109126 for the dominant model	147
Table 47: Phenotype distribution per genotype for rs11192037 for the dominant model	147
Table 48: Frequency of the phenotype per genotype for rs1791810	149

Table 49: Frequency of the phenotype per genotype for rs1607098 150
Table 50: Frequency of the phenotype per genotype for rs1465555 for the recessive model 151
Table 51: Frequency of the phenotype per genotype for the overdominant model for
rs1887263
Table 52: Frequency of the phenotype per genotype for rs114920443 154
Table 53: Frequency of the phenotype per genotype for rs72641095 156
Table 54: Frequency of the phenotype per genotype for rs889472 for the recessive model 157 $$
Table 55: Phenotype distribution per genotype for rs11805515
Table 56: Frequency of the phenotype per genotype in the log-additive model for rs11805515
Table 57: Frequency of the phenotype per genotype for rs77698137 163

Research communication

Publications

Walton AD, Moret S, Gunn P, Barash M. Comment on "Linkage analysis of a model quantitative trait in humans: Finger ridge count shows significant multivariate linkage to 5q14. 1" by Medland *et al.*, "Common Genetic Variants Influence Whorls in Fingerprint Patterns" by Ho *et al.* and "Hot on the Trail of Genes that Shape Our Fingerprints" by Walsh *et al.* Forensic Science International: Genetics. 2018.

Walton A, Moret S, Barash M, Gunn P. (2019). "The frequency of fingerprint patterns separated by ancestry and sex in a general population from Sydney, Australia." Australian Journal of Forensic Sciences: 1-6.

Presentations

ANZFSS 24th International Symposium – September 14th, 2018, Perth – "The frequency of fingerprint patterns separated by ancestry and sex in a general population from Sydney, Australia" – Walton A, Moret S, Barash M, Gunn P.

NSWPF Fingerprint Expert Conference – December 11th, 2019, Parramatta – "Determining the effect of ancestry and sex on fingerprint patterns" – Walton A, Moret S, Barash M, Gunn P.