

ORIGINAL RESEARCH ARTICLE

Open Access



Genetic Variants within *NOGGIN*, *COL1A1*, *COL5A1*, and *IGF2* are Associated with Musculoskeletal Injuries in Elite Male Australian Football League Players: A Preliminary Study

Ysabel Jacob¹, Ryan S. Anderton^{2,3*}, Jodie L. Cochrane Wilkie^{1,4}, Brent Rogalski⁵, Simon M. Laws^{6,7,8}, Anthony Jones⁵, Tania Spiteri¹, Dana Hince² and Nicolas H. Hart^{1,2,4,9,10*} 

Abstract

Introduction: Australian Football is a dynamic team sport that requires many athletic traits to succeed. Due to this combination of traits, as well as technical skill and physicality, there are many types of injuries that could occur. Injuries are not only a hindrance to the individual player, but to the team as a whole. Many strength and conditioning personnel strive to minimise injuries to players to accomplish team success.

Purpose: To investigate whether selected polymorphisms have an association with injury occurrence in elite male Australian Football players.

Methods: Using DNA obtained from 46 elite male players, we investigated the associations of injury-related polymorphisms across multiple genes (*ACTN3*, *CCL2*, *COL1A1*, *COL5A1*, *COL12A1*, *EMILIN1*, *IGF2*, *NOGGIN*, *SMAD6*) with injury incidence, severity, type (contact and non-contact), and tissue (muscle, bone, tendon, ligament) over 7 years in one Australian Football League team.

Results: A significant association was observed between the rs1372857 variant in *NOGGIN* ($p = 0.023$) and the number of total muscle injuries, with carriers of the GG genotype having a higher estimated number of injuries, and moderate, or combined moderate and high severity rated total muscle injuries. The *COL5A1* rs12722TT genotype also had a significant association ($p = 0.028$) with the number of total muscle injuries. The *COL5A1* variant also had a significant association with contact bone injuries ($p = 0.030$), with a significant association being found with moderate rated injuries. The *IGF2* rs3213221-CC variant was significantly associated with a higher estimated number of contact tendon injuries per game ($p = 0.028$), while a higher estimated number of total ligament ($p = 0.019$) and non-contact ligament ($p = 0.002$) injuries per game were significantly associated with carriage of the *COL1A1* rs1800012-TT genotype.

Conclusions: Our preliminary study is the first to examine associations between genetic variants and injury in Australian Football. *NOGGIN* rs1372857-GG, *COL5A1* rs12722-TT, *IGF2* rs3213221-CC, and *COL1A1* rs1800012-TT genotypes

*Correspondence: ryan.anderton@nd.edu.au; n.hart@ecu.edu.au

¹ School of Medical and Health Sciences, Edith Cowan University, Perth, WA, Australia

² Institute for Health Research, University of Notre Dame Australia, Perth, WA, Australia

Full list of author information is available at the end of the article

held various associations with muscle-, bone-, tendon- and ligament-related injuries of differing severities. To further increase our understanding of these, and other, genetic variant associations with injury, competition-wide AFL studies that use more players and a larger array of gene candidates is essential.

Keywords: Injury, Muscle, Tendon, Ligament, Bone, Contact, Non-contact, Genetics, Genes

Key Points

- In this select cohort of elite Australian Football players, those with the rs1372857-*NOGGIN* GG or *COL5A1* rs12722-TT genotype had a higher estimated number of total muscle-related injuries per game. However, those with the *NOGGIN* rs1372857-GG genotype also had a higher estimated number of moderate-to-high severity rated injuries.
- The *IGF2* rs3213221-CC genotype had a higher estimated number of contact tendon injuries per game, and a higher number of low severity rated injuries.
- The rs1800012-TT genotype of *COL1A1* had a higher number of ligament-related injuries per game, with significant associations seen between the genotype and low severity rated injuries.
- The *COL5A1* rs12722-TT genotype had a higher number of contact bone injuries per game, and a higher number of moderate severity rated injuries.

Introduction

Australian Football (AF) is a unique endurance-based team sport interspersed with many high-intensity efforts across a match [1–4]. In the elite competition, the Australian Football League (AFL), players regularly run over 13-kms per match over a 120-min period [5]. Players continuously perform at an intense physical level due to the dynamic nature of the sport which requires players to accelerate and decelerate, change direction, and explosively jump repeatedly, while also performing sport specific skills such as kicking, handballing, marking, and tackling [6–10]. Due to the demands of the sport, injuries occur frequently with the most common injuries including hamstring strains, anterior cruciate ligament (ACL) ruptures, glenohumeral dislocations, leg and foot fractures (i.e. mainly stress fractures), and ankle joint injuries [11]. To be successful, AFL teams not only need talented athletes, but also lower injury rates to optimise player availability for team selection [12], as greater team continuity allows less disruption of athlete personnel during a season, which can lead to better team consistency and success [12]. Despite concerted efforts from strength and conditioning coaches, and medical staff, there is still an unknown combination of extrinsic and intrinsic

mechanisms that may affect an athlete's rate of injury and rate of recovery from injury, including genetic factors [13–15].

The expression of certain genes influences various physical athletic qualities such as body composition, muscle stiffness, elasticity and strength, and response to exercise-induced muscle damage [16–18]. Single nucleotide polymorphisms (SNPs) are naturally occurring variances in the human genome at a specific position, where an individual may have a pair of the same DNA bases (homozygous) or two different DNA bases (heterozygous) [19]. These polymorphisms could have no effect or could be beneficial or deleterious to the individual [19]. SNPs can have an effect on musculoskeletal formation, structure, repair, blood flow and metabolism [20–22], influencing muscles, tendons, and ligaments [23]. The presence of SNPs in different genes has been shown to be related to musculoskeletal injury risk (alpha-actinin-3 [*ACTN3*] [24]; collagen type I alpha 1 [*COL1A1*] [25]), musculoskeletal injury occurrence (collagen type V alpha 1 [*COL5A1*] [14]), muscle injury severity (*COL5A1*; insulin-like growth factor-2 [*IGF2*]; chemokine CC motif ligand-2 [*CCL2*] [26]), muscular strength (*CCL2* [23]), an influence on muscle function (*COL5A1* [27]), increased risk of ACL injury (*COL1A1*; and collagen type XII alpha 1 [*COL12A1*] [28, 29]), ligament injury severity (elastin microfibril interface 1 [*EMILIN1*] [26]), bone mineral density (*NOGGIN*; and SMAD Family Member 6 [*SMAD6*] [30–32]), and injury recovery time (*IGF2*; *CCL2* [26]).

SNPs in the *COL1A1*, *COL5A1*, and *COL12A1* genes have all been reported to affect the production of their associated collagen types. *COL1A1* affects cell adhesion and differentiation [33], *COL5A1* is a part of the extracellular matrix and can influence the production of type V collagen by altering mRNA stability [34], and *COL12A1* is the link between fibrils and its mutations [35]. *ACTN3* influences skeletal muscle formation as it forms the part of the fibres responsible for rapid and forceful contractions [36]. *IGF2* regulates cell proliferation, growth, migration, differentiation, and survival via its protein hormone [37]. *CCL2* can recruit monocytes, memory T cells, and dendritic cells as part of the CC chemokine superfamily responsible for chemotactic activity and increases in calcium influx [38]. *EMILIN1* regulates systemic blood pressure, as well as being part of the

extracellular matrix [39–41]. *NOGGIN* is an extracellular antagonist of the bone morphogenetic proteins (BMPs) that regulate heart development via complex morphogenetic processes [42]. *SMAD6* affects its protein which is an intracellular mediator of signalling caused by BMPs [43].

Genetic variants in multiple genes have been associated with injuries in clinical [25, 44, 45] and athletic [44, 46–48] populations. Within AF, team success relies on many factors such as talent, skill, and fitness; however, lower injury rates are also important, as injuries can keep players from playing for a period of time or to the best of their ability, and therefore, understanding potential links to injury, such as genetics, is important. Research has focussed on mechanisms of injury, for example, type of technique, loading, muscle strength, timing of muscle activation, previous injury history, age and fatigue [49–51], and external factors, such as ground contact forces and ground–shoe friction [11, 49], causing injury in AF. However, the association between genetic variants and injuries in AF has not been investigated. Accordingly, the purpose of this study is to investigate whether the previously researched candidate polymorphism has an association with injury occurrence in an elite AFL playing squad. We hypothesise that there will be a genetic influence on injury occurrence for at least one of the candidate polymorphisms.

Materials and Methods

Study Design

A prospective longitudinal cohort study was conducted across seven consecutive seasons (2011 to 2017) used to investigate the association of injury-related SNPs, in nine genes (*ACTN3*, *CCL2*, *COL1A1*, *COL5A1*, *COL12A1*, *EMILIN1*, *IGF2*, *NOGGIN*, and *SMAD6*), in a population of professional, elite AF players to explore possible relationships between candidate SNPs and injury outcomes (incidence [total, contact, non-contact] and severity [low, moderate, high]) relating to muscle, tendon, ligament, and bone. Estimated numbers of injuries per genotype for each genetic variant are reported to provide easy-to-interpret presentation of data for use by strength and conditioning coaches.

Participants

Forty-six ($n=46$) elite male AF players were recruited from one AFL team to participate in the study, as previously described [52, 53]. For each in-season round, 22 to 23 players were selected to play in the AFL competition, with the remaining 24 to 25 playing in the state competition for that given round (WAFL; Western Australian Football League). Anonymity was ensured by assigning players with a randomised, non-identifiable code. All

participants provided written informed consent after being provided with information letters outlining the purpose of the study and potential benefits and risks. All data collection and management procedures conformed to the Declaration of Helsinki (World Medical Association) with ethics approved by the Edith Cowan University Human Research and Ethics Committee (ID: 2019-00181-JACOB).

Injury Data Collection

Data were collected from medical and injury reports provided by the AFL club, recorded by their medical professionals, including doctors, physiotherapists, and strength and conditioning personnel. Injuries were diagnosed with a description of location and categorised as non-contact (i.e. intrinsic injuries that are acute, chronic, or overuse injuries stemming from the athlete themselves) or contact (i.e. extrinsic injuries that are acute collision or contact injuries stemming from external forces). Club medical staff provided each injury with a severity rating that is based on the number of training sessions and games that elapsed from the date of injury to the date of the player's return to full participation in team training with availability for match selection. Specifically, severity was graded as low (i.e. training is modified or less than 1 week of training is missed, with no games missed), moderate (i.e. 1–2 weeks of training missed, or unavailable for 1 to 2 games), or high (i.e. 3 or more weeks of training missed, or unavailable for 3 or more games) as an internal club-determined metric. All injuries were then placed into a sub-category: bursitis; concussion; contusion/bruise/haematoma; dislocation/subluxation; fracture; lesion of meniscus, cartilage or disc; muscle strain/tear/rupture/cramps; other bone injuries; other injuries; sprain/ligament injury; joint instability; tendinopathy; or tendon injury/rupture. In our study, injuries that could be categorised into the broad categories of (1) muscle (including muscle strain/tear/rupture/cramps), (2) tendon (tendinopathy, and tendon injury/rupture), (3) ligament (including sprain/ligament injury, and joint instability), and (4) bone (including fracture, and other bone injuries) were used. Data were collected over 7 consecutive seasons (2011 to 2017). Injuries occurring in training sessions or matches during the pre-season (between 17 and 21 weeks depending on the year and season over the Australian summer) and in-season (between 23 and 27 weeks depending on the year and season over the Australian winter with the inclusion of a final series when applicable) were included. Players who were injury-free in-season were either selected to play in the national AFL competition or played in the state WAFL competition. Due to the professional nature of the

team and club, all players undertook a similar volume of training when not injured.

Sample Collection and DNA Analysis

Buccal saliva samples were collected via mouth swabs with participants instructed to brush the edge of a soft tip swab along the insides of their cheek and gums for 30 s [52–54]. Samples were collected before a pre-season training session and players were asked not to consume coffee, alcohol, or food for two hours prior to saliva collection. Collected samples were labelled with a numeric code for de-identification and were sent to the Australian Genome Research Facility (AGRF; Brisbane, QLD, Australia; NATA 17025) for DNA extraction and genotyping using Agena Bioscience MassARRAY system (AGRF; Brisbane, QLD, Australia). Genetic variants examined were within the following genes: *ACTN3* (rs1815739), *CCL2* (rs2857656), *COL1A1* (rs1800012), *COL5A1* (rs12722), *COL12A1* (rs970547), *EMILIN1* (rs2289360), *IGF2* (rs3213221), *NOGGIN* (rs1372857), *SMAD6* (rs2053423), with all genotypes being within Hardy–Weinberg equilibrium (HWE), as previously reported [52].

Statistical Analysis

Data were statistically analysed using IBM-SPSS V.24 (Armonk, NY, USA) and Stata/BE v17 (StataCorp LLC,

Collage Station, TX, USA). A negative binomial distribution model was used to assess the relationship between the total number of injuries per injury category (muscle, tendon, ligament, and bone) and the genotypes of each of the candidate genes. Each injury category was analysed in total and via non-contact (i.e. intrinsic), and contact (i.e. extrinsic) injury mechanism. The number of games played was included as the offset variable to account for differences in exposure to the risk of injury. Genotype was considered a continuous variable to test for the linear trend in the association. Significant association with a particular gene was followed up with separate models to determine if there was an association between the number of injuries within each injury category and their severity (low, moderate, high, and a combination group of moderate and high). These results are reported as incident rate ratios (IRRs) and estimated number of injuries with 95% confidence intervals (95% CI). A significant nominal *p* value of < 0.05 was employed.

Results

Longitudinal team descriptive results are presented in Table 1, with genotype frequencies presented in Table 2. The average number of injuries for each variant per season is presented in Table 3 for muscle, tendon, ligament, and bone-related injuries. Over the seven seasons, 992 incidents of injury were included, of which 351

Table 1 Descriptive statistics and mean player injuries for each season

AFL season							
Variable	Season 1 (n = 46)	Season 2 (n = 40)	Season 3 (n = 29)	Season 4 (n = 25)	Season 5 (n = 21)	Season 6 (n = 18)	Season 7 (n = 15)
Age (Years)	24.94 (4.26)	24.20 (4.02)	23.90 (3.21)	23.60 (2.87)	23.38 (2.42)	22.50 (2.50)	21.93 (2.38)
Active weeks	35.23 (6.25)	35.73 (6.02)	41.45 (5.42)	36.60 (3.43)	35.19 (5.38)	37.28 (5.85)	41.20 (3.92)
Games played	17.45 (45)	18.78 (2.92)	20.03 (6.17)	18.60 (3.43)	17.57 (4.50)	19.28 (5.85)	20.73 (3.97)
Games lost	4.62 (6.72)	1.60 (2.92)	4.31 (6.93)	2.56 (3.56)	4.62 (4.58)	3.83 (6.29)	3.40 (4.38)
All Injuries	1.85 (1.61)	2.14 (3.35)	1.61 (1.37)	1.78 (1.19)	2.20 (1.56)	1.57 (1.16)	1.50 (1.15)
Muscle	3.40 (1.80)	2.08 (1.00)	2.06 (1.65)	1.91 (0.85)	3.05 (1.60)	1.72 (0.72)	1.82 (1.06)
Tendon	0.67 (0.47)	2.33 (1.25)	1.13 (0.33)	1.14 (0.35)	2.00 (1.41)	1.45 (0.66)	1.08 (0.28)
Ligament	1.26 (1.31)	1.65 (1.56)	1.90 (1.60)	2.08 (1.62)	1.48 (1.43)	1.56 (1.54)	1.27 (1.12)
Bone	0.62 (1.02)	0.40 (0.58)	0.83 (1.05)	0.68 (0.79)	0.48 (0.73)	0.39 (0.49)	0.53 (0.72)
Non-Contact Injuries	2.12 (1.78)	1.50 (0.78)	1.58 (1.34)	1.50 (0.76)	2.29 (1.35)	1.35 (0.58)	1.54 (0.93)
Muscle	2.70 (2.10)	1.67 (0.62)	1.94 (1.65)	1.75 (0.77)	2.70 (1.45)	1.62 (0.72)	1.75 (1.00)
Tendon	2.00 (0.00)	2.00 (1.41)	1.00 (0.00)	1.29 (0.45)	2.20 (1.17)	1.18 (0.39)	1.09 (0.29)
Ligament	1.00 (0.00)	1.00 (0.00)	1.50 (0.50)	1.46 (0.93)	1.50 (0.71)	1.21 (0.41)	1.07 (0.26)
Bone	1.50 (0.50)	1.00 (0.00)	1.00 (0.00)	1.13 (0.33)	2.13 (1.27)	1.13 (0.33)	2.13 (1.27)
Contact Injuries	1.39 (0.59)	1.67 (1.05)	1.55 (0.74)	1.35 (0.62)	1.65 (0.90)	1.59 (0.93)	1.55 (0.76)
Muscle	1.00 (0.00)	1.33 (0.47)	1.00 (0.00)	1.00 (0.00)	1.50 (0.50)	1.33 (0.47)	0.00 (0.00)
Tendon	0.00 (0.00)	0.00 (0.00)	1.00 (0.00)	1.00 (0.00)	1.00 (0.00)	1.00 (0.00)	1.00 (0.00)
Ligament	1.67 (0.67)	1.92 (1.19)	1.77 (0.80)	1.41 (0.60)	1.95 (0.97)	1.82 (1.03)	1.60 (0.80)
Bone	1.25 (0.43)	1.00 (0.00)	1.25 (0.43)	1.40 (0.80)	1.00 (0.00)	1.00 (0.00)	1.40 (0.49)

Table 2 Player genotype and allele distribution of candidate variants

	n (%)
<i>COL5A1</i> (rs1800012)	
CC	8 (17.0)
CT	23 (48.9)
TT	16 (34.0)
C	39 (41.5)
T	55 (58.5)
<i>NOGGIN</i> (rs1372857)	
GG	9 (19.1)
AG	22 (46.8)
AA	16 (34.0)
G	40 (42.6)
A	54 (57.4)
<i>COL1A1</i> (rs1800012)	
TT	32 (69.6)
GT	14 (30.4)
GG	0 (0.0)
T	78 (84.8)
G	54 (15.2)
<i>ACTN3</i> (rs1815739)	
CC	21 (44.7)
CT	24 (51.1)
TT	2 (4.3)
C	66 (70.2)
T	28 (29.8)
<i>SMAD6</i> (rs2053423)	
CC	8 (17.0)
CT	15 (31.9)
TT	24 (51.1)
C	31 (33.0)
T	63 (67.0)
<i>EMILIN1</i> (rs2289360)	
GG	20 (42.6)
AG	15 (31.9)
AA	12 (25.5)
G	55 (58.5)
A	39 (41.5)
<i>CCL2</i> (rs2857656)	
CC	2 (4.3)
CG	24 (51.1)
GG	21 (44.7)
C	28 (29.8)
G	66 (70.2)
<i>IGF2</i> (rs3213221)	
CC	7 (14.9)
GC	24 (51.1)
GG	16 (34.0)
C	38 (40.4)
G	56 (59.6)

Table 2 (continued)

	n (%)
<i>COL12A1</i> (rs970547)	
GG	1 (2.1)
AG	13 (27.7)
AA	33 (70.2)
G	15 (16.0)
A	79 (84.0)

incidents were contact injuries, and 553 were non-contact injuries, with 88 injuries unclassified by the football club personnel at the point of collection. Within injury categories, (1) muscle injuries had 299 incidents [21 contact, 261 non-contact, 17 unclassified]; (2) tendon injuries had 73 incidents [8 contact, 60 non-contact, 5 unclassified]; (3) ligament injuries had 304 incidents [203 contact, 73 non-contact, 28 unclassified]; and (4) bone injuries had 111 incidents [39 contact, 66 non-contact, 6 unclassified].

Genetic associations with the total number of injuries, with muscle, tendon, ligament, and bone are presented in Table 4. The rs1372857 variant within *NOGGIN* ($p=0.050$) and the rs12722 variant within the *COL5A1* ($p=0.028$) genes were the variants associated with all muscle-related injuries. Trends were seen for the rs2857656 *CCL2* ($p=0.082$) and the rs3213221 *IGF2* ($p=0.097$) variants. Trends were also seen with *COL1A1* rs1800012 for non-contact muscle injuries ($p=0.100$) and *NOGGIN* rs1372857 for contact muscle injuries ($p=0.054$). Associations were seen for the *NOGGIN* rs1372857 variant for moderate ($p=0.044$) and moderate and high combined ($p=0.016$) severity total muscle injuries. Trends were seen between *NOGGIN* rs1372857 and high severity total muscle injuries ($p=0.084$), as well as for the *COL5A1* rs12722 variant for low ($p=0.073$) and moderate ($p=0.073$) severity total muscle injuries. There is an association for the rs3213221 *IGF2* variant for contact tendon injuries ($p=0.028$). Trends were seen for the rs970547 *COL12A1* variant for all tendon ($p=0.084$) and non-contact tendon injuries ($p=0.079$); however, an association for the rs970547 *COL12A1* variant was seen for low severity contact tendon injuries ($p=0.026$). Significant associations were found for ligament injuries, specifically between the rs1800012 *COL1A1* variant and all ligament injuries ($p=0.019$), with further associations observed with low severity ($p=0.002$), and non-contact ligament injuries ($p=0.002$), with further associations with low severity rated non-contact injuries ($p=0.004$). The rs12722 *COL5A1* variant was associated with contact bone injuries ($p=0.030$), with a further association

Table 3 Mean number of total muscle and tendon, ligament, and bone injuries when categorised by candidate variant genotype

Gene	Genotype	Muscle		Tendon		Ligament		Bone	
		Mean	95% CI	Mean	95% CI	Mean	95% CI	Mean	95% CI
<i>All injuries</i>									
<i>COL5A1</i>	CC	4.57	[− 0.08 to 9.22]	2.14	[− 0.70 to 4.99]	6.57	[0.90 to 12.24]	1.57	[− 0.27 to 3.41]
(rs12722)	CT	6.96	[5.15 to 8.76]	1.48	[0.77 to 2.19]	8.09	[4.25 to 1.92]	2.78	[1.56 to 4.01]
	TT	7.40	[3.76 to 11.13]	1.40	[0.54 to 2.26]	6.53	[2.37 to 10.69]	2.47	[1.36 to 3.57]
<i>NOGGIN</i>	GG	7.29	[2.67 to 11.90]	2.71	[− 0.10 to 5.53]	7.71	[1.03 to 14.40]	3.00	[1.00 to 5.00]
(rs1372857)	AG	7.73	[5.25 to 10.20]	1.27	[0.78 to 1.77]	6.73	[4.03 to 9.43]	2.82	[1.64 to 3.99]
	AA	5.13	[2.59 to 7.66]	1.44	[0.37 to 2.50]	8.00	[2.39 to 13.61]	1.81	[0.61 to 3.02]
<i>COL1A1</i>	TT	7.00	[5.27 to 8.73]	1.59	[0.92 to 2.27]	8.72	[5.51 to 11.93]	2.69	[1.77 to 3.60]
(rs1800012)	GT	6.08	[2.24 to 9.92]	1.46	[0.29 to 2.63]	3.92	[2.12 to 5.73]	2.00	[0.65 to 3.35]
	GG	–	–	–	–	–	–	–	–
<i>ACTN3</i>	CC	8.15	[5.68 to 10.62]	1.95	[0.82 to 3.08]	8.45	[5.33 to 11.57]	2.85	[1.86 to 3.84]
(rs1815739)	CT	6.00	[3.82 to 8.18]	1.30	[0.76 to 1.85]	6.96	[3.04 to 10.87]	2.26	[1.05 to 3.47]
	TT	1.00	[1.00 to 1.00]	0.50	[− 5.85 to 6.85]	0.50	[− 5.85 to 6.85]	1.50	[− 4.85 to 7.85]
<i>SMAD6</i>	CC	7.57	[0.63 to 14.51]	1.57	[0.39 to 2.75]	4.14	[1.30 to 6.99]	3.57	[0.70 to 6.44]
(rs2053423)	CT	6.13	[3.68 to 8.59]	2.20	[0.73 to 3.67]	11.00	[5.31 to 16.69]	3.13	[1.99 to 4.28]
	TT	6.87	[4.64 to 9.10]	1.13	[0.59 to 1.67]	5.91	[3.12 to 8.71]	1.74	[0.74 to 2.74]
<i>EMILIN1</i>	GG	6.70	[4.19 to 9.21]	1.80	[0.91 to 2.69]	8.60	[5.09 to 12.11]	2.55	[1.28 to 3.82]
(rs2289360)	AG	6.60	[4.19 to 9.01]	1.40	[0.15 to 2.65]	6.53	[1.03 to 12.04]	2.47	[1.16 to 3.77]
	AA	7.00	[2.25 to 11.75]	1.30	[0.54 to 2.06]	6.00	[2.14 to 9.86]	2.40	[0.78 to 4.02]
<i>CCL2</i>	CC	0.00	[0.00 to 0.00]	0.00	[0.00 to 0.00]	1.00	[− 11.71 to 13.71]	3.00	[3.00 to 3.00]
(rs2857656)	CG	7.48	[5.22 to 9.73]	1.96	[0.96 to 2.95]	9.17	[4.95 to 13.40]	3.00	[1.81 to 4.19]
	GG	6.55	[4.13 to 8.97]	1.25	[0.71 to 1.79]	5.85	[3.53 to 8.17]	1.85	[0.87 to 2.83]
<i>IGF2</i>	CC	5.00	[0.96 to 9.04]	0.83	[− 0.39 to 2.06]	5.33	[1.15 to 9.51]	1.17	[− 0.06 to 2.39]
(rs3213221)	GC	8.88	[6.54 to 11.21]	2.04	[1.07 to 3.02]	9.58	[5.54 to 13.62]	3.04	[2.18 to 3.90]
	GG	4.00	[1.96 to 6.04]	1.07	[0.58 to 1.56]	4.53	[1.86 to 7.20]	2.13	[0.37 to 3.90]
<i>COL12A1</i>	GG	–	–	–	–	–	–	–	–
(rs970547)	AG	7.31	[3.33 to 11.28]	1.15	[0.42 to 1.89]	9.92	[2.91 to 16.93]	3.38	[2.24 to 4.53]
	AA	6.23	[4.57 to 7.88]	1.74	[0.97 to 2.51]	6.19	[4.05 to 8.34]	2.10	[1.14 to 3.05]
<i>Non-Contact Injuries</i>									
<i>COL5A1</i>	CC	4.14	[− 0.14 to 8.43]	1.86	[− 0.73 to 4.44]	1.00	[− 0.41 to 2.41]	1.00	[− 0.41 to 2.41]
(rs12722)	CT	6.04	[4.58 to 7.50]	1.30	[0.71 to 1.89]	1.65	[0.86 to 2.44]	1.87	[0.72 to 3.02]
	TT	6.00	[2.76 to 9.24]	1.13	[0.35 to 1.91]	1.80	[0.67 to 2.93]	0.93	[0.36 to 1.51]
<i>NOGGIN</i>	GG	6.43	[2.62 to 10.24]	2.43	[− 0.01 to 4.87]	1.86	[− 0.50 to 4.21]	1.86	[− 0.44 to 4.15]
(rs1372857)	AG	6.50	[4.28 to 8.72]	1.00	[0.61 to 1.39]	1.50	[0.83 to 2.17]	1.55	[0.46 to 2.63]
	AA	4.38	[2.41 to 6.34]	1.31	[0.35 to 2.28]	1.63	[0.54 to 2.71]	1.06	[0.35 to 1.78]
<i>COL1A1</i>	TT	5.91	[4.43 to 7.38]	1.38	[0.78 to 1.97]	2.06	[1.35 to 2.77]	1.59	[0.84 to 2.35]
(rs1800012)	GT	5.31	[2.01 to 8.60]	1.23	[0.21 to 2.25]	0.46	[0.06 to 0.86]	1.00	[− 0.28 to 2.28]
	GG	–	–	–	–	–	–	–	–
<i>ACTN3</i>	CC	7.20	[4.96 to 9.44]	1.55	[0.60 to 2.50]	2.10	[1.19 to 3.01]	1.90	[0.95 to 2.85]
(rs1815739)	CT	4.91	[3.24 to 6.59]	1.22	[0.66 to 1.77]	1.30	[0.56 to 2.05]	1.13	[0.19 to 2.07]
	TT	0.50	[− 5.85 to 6.85]	0.50	[− 5.85 to 6.85]	0.00	[0.00 to 0.00]	0.00	[0.00 to 0.00]
<i>SMAD6</i>	CC	7.00	[0.73 to 13.27]	1.43	[0.38 to 2.48]	0.86	[0.22 to 1.50]	2.29	[− 1.04 to 5.61]
(rs2053423)	CT	5.20	[3.13 to 7.27]	1.87	[0.58 to 3.15]	2.20	[1.03 to 3.37]	1.67	[0.84 to 2.50]
	TT	5.70	[3.87 to 7.52]	0.96	[0.48 to 1.44]	1.43	[0.64 to 2.23]	1.00	[0.21 to 1.79]
<i>EMILIN1</i>	GG	5.60	[3.43 to 7.77]	1.35	[0.57 to 2.13]	2.00	[1.03 to 2.97]	1.50	[0.37 to 2.63]
(rs2289360)	AG	5.60	[3.75 to 7.45]	1.40	[0.30 to 2.50]	1.07	[0.17 to 1.97]	1.67	[0.49 to 2.84]
	AA	6.20	[1.99 to 10.41]	1.20	[0.46 to 1.94]	1.60	[0.47 to 2.73]	0.90	[− 0.02 to 1.82]
<i>CCL2</i>	CC	0.00	[0.00 to 0.00]	0.00	[0.00 – 0.00]	0.00	[0.00 to 0.00]	0.50	[− 5.85 to 6.85]

Table 3 (continued)

Gene	Genotype	Muscle		Tendon		Ligament		Bone	
		Mean	95% CI	Mean	95% CI	Mean	95% CI	Mean	95% CI
(rs2857656)	CG	6.48	[4.54 to 8.41]	1.74	[0.86 to 2.62]	1.87	[0.88 to 2.86]	2.04	[0.96 to 3.13]
	GG	5.45	[3.42 to 7.48]	1.00	[0.54 to 1.46]	1.45	[0.89 to 2.01]	0.80	[0.16 to 1.44]
IGF2 (rs3213221)	CC	4.17	[1.02 to 7.31]	0.50	[- 0.38 to 1.38]	1.50	[0.40 to 2.60]	0.67	[- 0.60 to 1.94]
	GC	7.50	[5.46 to 9.54]	1.67	[0.80 to 2.54]	2.08	[1.16 to 3.01]	1.54	[0.81 to 2.28]
	GG	3.53	[1.81 to 5.26]	1.13	[0.67 to 1.60]	0.87	[0.21 to 1.52]	1.53	[- 0.03 to 3.10]
COL12A1 (rs970547)	GG	-	-	-	-	-	-	-	-
	AG	6.31	[2.78 to 9.83]	0.92	[0.46 to 1.38]	1.85	[0.41 to 3.28]	2.15	[0.92 to 3.38]
	AA	5.23	[3.90 to 6.55]	1.52	[0.82 to 2.21]	1.52	[0.92 to 2.11]	1.13	[0.36 to 1.90]
<i>Contact injuries</i>									
COL5A1 (rs12722)	CC	0.29	[- 0.17 to 0.74]	0.29	[- 0.17 to 0.74]	5.14	[1.28 to 9.01]	0.57	[- 0.33 to 1.47]
	CT	0.61	[0.20 to 1.02]	0.13	[- 0.02 to 0.28]	4.61	[3.00 to 6.22]	0.65	[0.23 to 1.08]
	TT	0.33	[- 0.01 to 0.68]	0.20	[- 0.03 to 0.43]	4.07	[1.29 to 6.85]	1.27	[0.47 to 2.06]
NOGGIN (rs1372857)	GG	0.43	[- 0.07 to 0.92]	0.29	[- 0.17 to 0.74]	5.29	[1.48 to 9.09]	1.14	[0.02 to 2.27]
	AG	0.55	[0.25 to 0.84]	0.23	[0.04 to 0.42]	4.32	[2.49 to 6.15]	0.91	[0.46 to 1.36]
	AA	0.38	[- 0.17 to 0.92]	0.06	[- 0.07 to 0.20]	4.44	[2.04 to 6.84]	0.65	[- 0.10 to 1.35]
COL1A1 (rs1800012)	TT	0.59	[0.28 to 0.91]	0.22	[0.07 to 0.37]	5.09	[3.42 to 6.77]	0.94	[0.49 to 1.39]
	GT	0.15	[- 0.07 to 0.38]	0.08	[- 0.09 to 0.24]	3.08	[1.67 to 4.48]	0.62	[0.03 to 1.20]
	GG	-	-	-	-	-	-	-	-
ACTN3 (rs1815739)	CC	0.45	[0.13 to 0.77]	0.25	[0.04 to 0.46]	5.45	[3.43 to 7.47]	0.80	[0.35 to 1.25]
	CT	0.52	[0.13 to 0.91]	0.13	[- 0.02 to 0.28]	4.09	[2.35 to 5.82]	0.87	[0.27 to 1.47]
	TT	0.00	[0.00 to 0.00]	0.00	[0.00 to 0.00]	0.00	[0.00 to 0.00]	1.00	[1.00 to 1.00]
SMAD6 (rs2053423)	CC	0.29	[- 0.17 to 0.74]	0.00	[0.00 to 0.00]	2.86	[0.69 to 5.02]	0.86	[- 0.27 to 1.98]
	CT	0.53	[0.18 to 0.89]	0.27	[0.01 to 0.52]	6.20	[3.77 to 8.63]	1.13	[0.51 to 1.76]
	TT	0.48	[0.07 to 0.89]	0.17	[0.01 to 0.34]	3.91	[2.08 to 5.75]	0.65	[0.14 to 1.17]
EMILIN1 (rs2289360)	GG	0.50	[0.18 to 0.82]	0.30	[0.08 to 0.52]	5.40	[3.15 to 7.65]	0.85	[0.26 to 1.44]
	AG	0.33	[0.06 to 0.60]	0.07	[- 0.08 to 0.21]	3.80	[1.88 to 5.72]	0.60	[0.14 to 1.06]
	AA	0.60	[- 0.30 to 1.50]	0.10	[- 0.13 to 0.33]	3.80	[1.04 to 6.56]	1.20	[0.20 to 2.20]
CCL2 (rs2857656)	CC	0.00	[0.00 to 0.00]	0.00	[0.00 - 0.00]	0.50	[- 5.85 to 6.85]	1.00	[- 11.71 to 13.71]
	CG	0.48	[0.09 to 0.87]	0.17	[0.01 to 0.34]	5.52	[3.51 to 7.53]	0.78	[0.26 to 1.30]
	GG	0.50	[0.18 to 0.82]	0.20	[0.01 to 0.39]	3.75	[2.14 to 5.36]	0.90	[0.35 to 1.45]
IGF2 (rs3213221)	CC	0.50	[- 0.38 to 1.38]	0.33	[- 0.21 to 0.88]	3.50	[0.55 to 6.45]	0.50	[- 0.38 to 1.38]
	GC	0.63	[0.23 to 1.02]	0.25	[0.06 to 0.44]	5.67	[3.71 to 7.62]	1.29	[0.76 to 1.83]
	GG	0.20	[- 0.03 to 0.43]	0.00	[0.00 to 0.00]	3.07	[1.15 to 4.98]	0.27	[- 0.18 to 0.71]
COL12A1 (rs970547)	GG	-	-	-	-	-	-	-	-
	AG	0.46	[0.06 to 0.86]	0.08	[- 0.09 to 0.24]	5.54	[2.53 to 8.55]	0.85	[0.25 to 1.44]
	AA	0.45	[0.14 to 0.76]	0.23	[0.07 to 0.38]	4.00	[2.59 to 5.41]	0.81	[0.35 to 1.26]

seen with the number of moderate severity injuries ($p=0.049$). A trend was also seen with respect to the number of moderate and high combined severity injuries ($p=0.065$).

The *NOGGIN* rs1372857 variant had an IRR of 0.813 [0.66 to 1.00] for total muscle injuries, with analysis finding those with the *NOGGIN* GG genotype experiencing an estimated number of injuries per game of 8.44 [6.16 to 1.71], compared to 6.86 [7.27 to 9.40] for the

heterozygous genotype and 5.58 [4.28 to 6.87] for the AA genotype. When examining moderate severity injuries, the IRR was 0.691 [0.48 to 0.99], with the estimated number of injuries being 3.45 (GG) [1.88 to 5.01], 2.38 (AG) [1.80 to 2.95], and 1.64 (AA) [0.96 to 2.32]. When analysing moderate and high severity injuries, the IRR was 0.639 [0.44 to 0.92], with the estimated number of injuries being 4.71 (GG) [2.53 to 6.89], 3.01 (AG) [2.27 to 3.74], and 1.92 (AA) [1.13 to 2.72].

Table 4 Negative binominal model of genetic variants and total number of muscle, tendon, ligament, and bone-related injuries

Variant	Muscle			Tendon			Ligament			Bone			
	IRR	p value	95% CI	IRR	p value	95% CI	IRR	p value	95% CI	IRR	p value	95% CI	
<i>All injuries</i>													
COL5A1	Intercept	0.026	0.000	[0.02 to 0.04]	0.012	0.000	[0.00 to 0.03]	0.037	0.000	[0.02 to 0.08]	0.10	0.000	[0.00 to 0.02]
(rs12722)	COL5A1	1.253	0.028	[1.03 to 1.53]	0.920	0.705	[0.60 to 1.42]	1.114	0.504	[0.81 to 1.53]	1.206	0.244	[0.88 to 1.65]
NOGGIN	Intercept	0.066	0.000	[0.04 to 0.11]	0.016	0.000	[0.01 to 0.04]	0.043	<0.001	[0.02 to 0.10]	0.024	<0.001	[0.01 to 0.04]
(rs1372857)	NOGGIN	0.813	0.050	[0.66 to 1.00]	0.802	0.285	[0.54 to 1.20]	1.038	0.833	[0.74 to 1.46]	0.820	0.159	[0.60 to 1.09]
COL1A1	Intercept	0.044	0.000	[0.03 to 0.07]	0.009	0.000	[0.00 to 0.02]	0.090	<0.001	[0.05 to 0.17]	0.021	0.000	[0.01 to 0.04]
(rs1800012)	COL1A1	0.967	0.842	[0.69 to 1.34]	1.087	0.808	[0.55 to 2.13]	0.567	0.019	[0.35 to 0.91]	0.763	0.360	[0.43 to 1.36]
ACTN3	Intercept	0.055	0.000	[0.04 to 0.09]	0.012	0.000	[0.01 to 0.03]	0.045	0.000	[0.02 to 0.08]	0.016	0.000	[0.01 to 0.03]
(rs1815739)	ACTN3	0.845	0.226	[0.64 to 1.11]	0.861	0.594	[0.50 to 1.49]	1.031	0.884	[0.69 to 1.54]	0.972	0.896	[0.63 to 1.50]
SMAD6	Intercept	0.034	0.000	[0.02 to 0.05]	0.011	0.000	[0.00 to 0.03]	0.040	0.000	[0.02 to 0.08]	0.022	0.000	[0.01 to 0.04]
(rs2053423)	SMAD6	1.103	0.317	[0.91 to 1.34]	0.938	0.758	[0.62 to 1.41]	1.074	0.590	[0.83 to 1.40]	0.844	0.207	[0.65 to 1.10]
EMILIN1	Intercept	0.040	0.000	[0.03 to 0.06]	0.014	0.000	[0.01 to 0.03]	0.063	0.000	[0.04 to 0.10]	0.018	0.000	[0.01 to 0.03]
(rs2289360)	EMILIN1	1.040	0.672	[0.88 to 1.25]	0.824	0.320	[0.56 to 1.21]	0.826	0.141	[0.64 to 1.07]	0.905	0.506	[0.67 to 1.21]
CCL2	Intercept	0.023	0.000	[0.01 to 0.05]	0.013	<0.001	[0.00 to 0.05]	0.091	<0.001	[0.03 to 0.25]	0.042	<0.001	[0.01 to 0.13]
(rs2857656)	CCL2	1.292	0.082	[1.00 to 1.23]	0.887	0.679	[0.50 to 1.56]	0.756	0.208	[0.49 to 1.17]	0.646	0.093	[0.39 to 1.08]
IGF2	Intercept	0.066	0.000	[0.04 to 0.11]	0.010	<0.001	[0.00 to 0.03]	0.076	<0.001	[0.03 to 0.18]	0.007	0.000	[0.00 to 0.02]
(rs3213221)	IGF2	0.814	0.097	[0.64 to 1.04]	0.996	0.988	[0.59 to 1.69]	0.799	0.277	[0.53 to 1.20]	1.393	0.093	[0.95 to 2.05]
COL12A1	Intercept	0.051	0.000	[0.03 to 0.10]	0.002	<0.001	[0.00 to 0.01]	0.080	<0.001	[0.04 to 0.17]	0.019	0.000	[0.00 to 0.05]
(rs970547)	COL12A1	0.937	0.623	[0.72 to 1.22]	1.698	0.084	[0.93 to 3.10]	0.812	0.181	[0.60 to 1.10]	0.920	0.639	[0.65 to 1.30]
<i>Non-contact injuries</i>													
COL5A1	Intercept	0.025	0.000	[0.02 to 0.04]	0.011	0.000	[0.00 to 0.03]	0.005	0.000	[0.00 to 0.01]	0.11	<0.001	[0.00 to 0.05]
(rs12722)	COL5A1	1.188	0.120	[0.96 to 1.48]	0.882	0.582	[0.57 to 1.38]	1.384	0.127	[0.91 to 2.10]	0.919	0.802	[0.48 to 1.78]
NOGGIN	Intercept	0.057	0.000	[0.04 to 0.09]	0.013	0.000	[0.00 to 0.03]	0.008	0.000	[0.00 to 0.02]	0.013	<0.001	[0.00 to 0.05]
(rs1372857)	NOGGIN	0.810	0.054	[0.66 to 1.00]	0.811	0.324	[0.54 to 1.23]	1.081	0.712	[0.71 to 1.64]	0.837	0.549	[0.47 to 1.50]
COL1A1	Intercept	0.036	0.000	[0.02 to 0.06]	0.008	0.000	[0.00 to 0.02]	0.049	<0.001	[0.02 to 0.13]	0.014	<0.001	[0.00 to 0.05]
(rs1800012)	COL1A1	1.009	0.961	[0.72 to 1.42]	1.042	0.906	[0.52 to 2.08]	0.255	0.002	[0.11 to 0.60]	0.711	0.476	[0.28 to 1.82]
ACTN3	Intercept	0.051	0.000	[0.03 to 0.08]	0.009	0.000	[0.00 to 0.02]	0.015	0.000	[0.01 to 0.04]	0.014	<0.001	[0.00 to 0.05]
(rs1815739)	ACTN3	0.795	0.110	[0.60 to 1.05]	0.991	0.974	[0.57 to 1.73]	0.745	0.275	[0.44 to 1.26]	0.746	0.448	[0.35 to 1.59]
SMAD6	Intercept	0.032	0.000	[0.02 to 0.05]	0.010	0.000	[0.00 to 0.03]	0.007	0.000	[0.00 to 0.02]	0.020	<0.001	[0.01 to 0.07]
(rs2053423)	SMAD6	1.057	0.588	[0.87 to 1.29]	0.907	0.644	[0.60 to 1.37]	1.191	0.388	[0.80 to 1.77]	0.704	0.204	[0.41 to 1.21]
EMILIN1	Intercept	0.032	0.000	[0.02 to 0.05]	0.009	0.000	[0.00 to 0.02]	0.014	0.000	[0.01 to 0.03]	0.011	0.000	[0.00 to 0.03]
(rs2289360)	EMILIN1	1.069	0.483	[0.89 to 1.29]	0.950	0.798	[0.64 to 1.40]	0.834	0.313	[0.59 to 1.19]	0.897	0.693	[0.53 to 1.54]
CCL2	Intercept	0.022	0.000	[0.01 to 0.05]	0.014	<0.001	[0.00 to 0.05]	0.007	<0.001	[0.00 to 0.03]	0.031	<0.001	[0.01 to 0.19]
(rs2857656)	CCL2	1.236	0.170	[0.91 to 1.67]	0.812	0.480	[0.46 to 1.45]	1.128	0.669	[0.65 to 1.96]	0.586	0.160	[0.28 to 1.24]

Table 4 (continued)

Variant	Muscle			Tendon			Ligament			Bone		
	IRR	p value	95% CI	IRR	p value	95% CI	IRR	p value	95% CI	IRR	p value	95% CI
<i>IGF2</i>	0.053	0.000	[0.03 to 0.09]	0.005	<0.001	[0.00 to 0.02]	0.025	<0.001	[0.01 to 0.07]	0.008	<0.001	[0.00 to 0.04]
(rs32132221)	0.838	0.175	[0.65 to 1.08]	1.272	0.389	[0.74 to 2.20]	0.648	0.082	[0.40 to 1.06]	1.071	0.846	[0.54 to 2.14]
<i>COL12A1</i>	0.049	0.000	[0.02 to 0.10]	0.002	<0.001	[0.00 to 0.01]	0.008	<0.001	[0.00 to 0.03]	0.041	0.005	[0.00 to 0.37]
(rs970547)	0.894	0.405	[0.69 to 1.16]	1.750	0.079	[0.94 to 2.37]	1.085	0.765	[0.64 to 1.86]	0.561	0.170	[0.25 to 1.28]
<i>Contact injuries</i>												
<i>COL5A1</i>	0.003	<0.001	[0.00 to 0.01]	0.001	<0.001	[0.00 to 0.01]	0.035	0.000	[0.02 to 0.07]	0.001	0.000	[0.00 to 0.01]
(rs12722)	1.021	0.950	[0.53 to 1.97]	0.979	0.967	[0.35 to 2.76]	0.906	0.518	[0.67 to 1.22]	1.824	0.030	[1.06 to 3.13]
<i>NOGGIN</i>	0.003	<0.001	[0.00 to 0.01]	0.004	<0.001	[0.01 to 0.03]	0.028	0.000	[0.02 to 0.05]	0.009	0.000	[0.00 to 0.03]
(rs1372857)	0.930	0.824	[0.49 to 1.76]	0.528	0.216	[0.19 to 1.45]	0.998	0.989	[0.75 to 1.33]	0.768	0.288	[0.47 to 1.25]
<i>COL1A1</i>	0.012	<0.001	[0.00 to 0.06]	0.003	<0.001	[0.00 to 0.04]	0.042	0.000	[0.02 to 0.08]	0.008	<0.001	[0.00 to 0.03]
(rs1800012)	0.294	0.100	[0.07 to 1.26]	0.399	0.390	[0.05 to 3.24]	0.727	0.184	[0.46 to 1.16]	0.736	0.502	[0.30 to 1.80]
<i>ACTN3</i>	0.002	0.000	[0.00 to 0.01]	0.002	<0.001	[0.00 to 0.02]	0.037	0.000	[0.02 to 0.07]	0.003	0.000	[0.00 to 0.01]
(rs1815739)	1.273	0.545	[0.58 to 2.79]	0.638	0.521	[0.16 to 2.52]	0.840	0.364	[0.58 to 1.22]	1.552	0.151	[0.85 to 2.83]
<i>SMAD6</i>	0.001	<0.001	[0.00 to 0.01]	<0.001	<0.001	[0.00 to 0.01]	0.021	0.000	[0.01 to 0.04]	-	-	-
(rs2053423)	1.341	0.353	[0.72 to 2.49]	1.567	0.400	[0.55 to 4.46]	1.130	0.399	[0.85 to 1.50]	-	-	-
<i>EMILIN1</i>	0.003	0.000	[0.00 to 0.01]	0.004	<0.001	[0.00 to 0.02]	0.039	0.000	[0.02 to 0.06]	0.004	0.000	[0.00 to 0.01]
(rs2289360)	1.040	0.889	[0.60 to 1.81]	0.447	0.163	[0.14 to 1.39]	0.830	0.143	[0.65 to 1.07]	1.175	0.491	[0.74 to 1.86]
<i>CCL2</i>	0.001	<0.001	[0.00 to 0.01]	<0.001	<0.001	[0.00 to 0.01]	0.031	<0.001	[0.01 to 0.08]	0.005	<0.001	[0.00 to 0.03]
(rs2857656)	1.349	0.477	[0.59 to 3.08]	1.469	0.572	[0.39 to 5.58]	0.963	0.845	[0.66 to 1.41]	1.066	0.858	[0.53 to 2.13]
<i>IGF2</i>	0.010	<0.001	[0.00 to 0.05]	0.017	<0.001	[0.00 to 0.17]	0.039	0.000	[0.02 to 0.08]	0.015	<0.001	[0.00 to 0.06]
(rs3213221)	0.559	0.146	[0.26 to 1.23]	0.249	0.028	[0.07 to 0.86]	0.865	0.392	[0.62 to 1.21]	0.603	0.108	[0.33 to 1.12]
<i>COL12A1</i>	0.003	<0.001	[0.00 to 0.02]	<0.001	<0.001	[0.00 to 0.01]	0.038	<0.001	[0.01 to 0.10]	0.006	<0.001	[0.00 to 0.03]
(rs970547)	0.987	0.974	[0.46 to 2.14]	3.506	0.225	[0.46 to 26.62]	0.896	0.562	[0.62 to 1.30]	0.946	0.856	[0.52 to 1.72]

Significant effects are **bolded**

The *COL5A1* rs12722 variant had an IRR of 1.253 [1.03 to 1.53] for total muscle injuries, with the TT genotype having a higher estimated number of injuries per game (8.00 [6.37 to 9.61]) than the CT (6.38 [5.50 to 7.25]) and CC genotypes (5.09 [3.67 to 6.51]).

In terms of contact tendon injuries, the *IGF2* rs3213221 variant had an IRR of 0.249 [0.07 to 0.86], with the CC genotype (0.66 [− 0.11 to 1.44]) having a higher estimated number of injuries per games than the CG (0.17 [0.04 to 0.29]) and GG (0.04 [− 0.03 to 0.11]) genotypes. When investigating low severity contact tendon injuries, the IRR was 0.222 [0.06 to 0.84], with the CC genotype (0.63 [− 0.13 to 1.40]) having a higher estimated number of injuries per games than the CG (0.14 [0.02 to 0.26]) and GG (0.03 [− 0.03 to 0.09]) genotypes.

For the *COL1A1* rs1800012 variant, with an observed IRR of 0.567 [0.35 to 0.91] for total ligament injuries, carriers of the TT genotype experienced an estimated number of injuries per game of 8.12 [6.31 to 9.92], compared to 4.61 [2.69 to 6.52] for the heterozygous genotype. The GG genotype was not represented in the current population. The IRR for low severity injuries was 0.602 [0.38 to 0.94], with the estimated number of injuries of the TT and GT genotypes being 7.00 [5.61 to 8.40] and 4.21 [2.52 to 5.91], respectively. For non-contact ligament injuries, the rs1800012 variant had an IRR of 0.255 [0.11 to 0.60], with the TT genotype having a higher estimated number of injuries per game (1.99 [1.48 to 2.50]) compared to the GT genotype (0.51 [0.10 to 0.92]). When analysing low severity non-contact ligament injuries, the IRR was 0.243, with the TT genotype (1.75 [1.22 to 2.27]) having a higher number of estimated injuries per game than the GT genotype (0.42 [0.04 to 0.81]).

For contact bone injuries, the *COL5A1* rs12722 variant had an IRR of 1.824 [1.06 to 3.13] with the TT genotype (1.28 [0.66 to 1.90]) having a higher number of estimated injuries per game compared to the CT (0.70 [0.42 to 0.98]) and CC genotypes (0.38 [0.07 to 0.70]). There were significant associations for moderate contact bone injuries with an IRR of 2.82 [1.01 to 1.89] with TT genotype (0.47 [0.12 to 0.82]) having a higher number of estimated injuries per game compared to the CT (0.17 [0.03 to 0.30]) and CC genotypes (0.06 [− 0.04 to 0.16]).

Discussion

Our preliminary investigation into the association of candidate genetic variants with injury number and severity in elite AFL players resulted in novel findings, and the identification of novel genetic markers for injury classification within AF. We discovered an association between the *NOGGIN* polymorphism (rs1372857) and all muscle injuries, with significantly higher muscle injury incidence for those with the GG genotype, which also trended

towards moderate-to-high combined severity injuries. The rs1372857 variant within the *NOGGIN* gene has been linked to bone fractures in a clinical study investigating motor vehicle accident, fall, and direct blow patients [31], with the homozygous GG genotype associated with non-union fractures [31]. While we did not identify a relationship between rs1372857 and bone injuries in our AFL cohort, we did observe a similar link between the GG genotype and a larger number of total injuries per season, together with more moderate-to-high severity of injuries. Given the inextricable link between muscle, tendon, and bone, and their co-adaptive processes [9], it seems reasonable to observe some crossover with *NOGGIN* expression and injuries to these tissues. Further research needs to be done to confirm whether this association between genetic variants within *NOGGIN* and subsequent muscle and tendon injuries in AFL players exists.

The current study found an association between the *COL5A1* rs12722 polymorphism and all muscle injuries, with the TT genotype having a higher estimated number of injuries per game. This follows previously reported associations between the CC genotype of rs12722 and less severe non-contact muscle injuries in soccer players [26, 55]. However, the variant was also found to show no significant difference between muscle injury and no muscle injury groups in a Japanese group of varied athletes [44]. The *COL5A1* gene encodes for the collagen type V $\alpha 1$ chain of protein and forms part of the extracellular matrix of skeletal muscles and can affect passive muscle stiffness as well as joint flexibility [56, 57]. The rs12722 polymorphism has also been associated with a higher susceptibility to ligament injuries [58]. We also found significant associations between the rs12722 TT genotype, contact bone injuries, and moderate severity contact bone injuries. Due to the findings being with contact, the polymorphism may have more of an effect on response to acute trauma.

We also found an association with the *IGF2* rs3213221 polymorphism and contact tendon injuries. This variant has previously been linked with tendon and muscle injuries in elite Caucasian soccer players, with a higher number of tendon injuries related to the presence of the C allele [59]. Our study found that those with the CC genotype had a higher estimated number of injuries per game compared to its counterparts, which coincides with previous findings. *IGF2* plays a role in modulating satellite cell activation and differentiation, thereby affecting soft tissue growth, as well as response to cell degeneration and regeneration following injury [26, 60].

Our study also discovered a significant association between the total number of ligament injuries with the rs1800012 variant of the *COL1A1* gene, with a significantly higher likelihood of low severity ligament-related

injuries in those with the *COL1A1* TT genotype, and a significant association between non-contact ligament injuries. These results are contradictory to previous research that has found that those with the TT genotype had less prevalence of ACL injuries [61] and cruciate ligament or shoulder dislocation injuries [62]. The *COL1A1* gene encodes the protein chain in type 1 collagen, a structural component in ligaments [61]. The T allele of the *COL1A1* gene is associated with an increased production of the protein chain in type 1 collagen [62]. Previous studies have investigated the effect of collagen peptides in muscle damage post-eccentric training [63] and in post-traumatic osteoarthritis in mice [64] and suggest that the supplementation of collagen may reduce inflammation. It has also been suggested that the increased production of the protein chain in type 1 collagen associated with the T allele increases the tensile strength of tendons and ligaments; however, the precise mechanism is unknown [62]. The contradictory results of the current study could be due to the GG genotype not being represented in the current population. Similarly, ligament injuries are multifactorial, ligaments are passive anatomical structures, and injuries cannot be solely predicted by genetic variations alone; thus, it could be the significant association between the *COL1A1* TT genotype and low-grade ligament injuries which infers a possible protective effect resulting in lower-grade rather than higher-grade injuries when the injurious events occur. This is speculative and would require thorough exploration of these associations and implications between *COL1A1* and the TT genotype with ligament injuries in AFL more broadly.

Stress fractures are prevalent in AFL, particularly for first- and second-year players who have less training and game experience at the elite level, and immature or developing physical structures yet to wholly tolerate these elevated physical demands [7, 11, 49, 65]. Although our study found no significant result between bone injuries and the genetic variants we explored in our AFL cohort, this may be due to the prospective observation of the same forty-six of various physical maturity overtime. Instead, for bone stress injuries (as disparate from other injury types), it may be better for future studies to prospectively observe early career AFL players in their first two seasons [66], who are typical candidates for stress fractures [65, 67], and use these early career cohorts to delineate genetic variant differences between those who sustain or avoid stress fractures. Despite previous associations with bone mineral density, the variant within the *SMAD6* gene had no significant association within the current study.

This study has several strengths and limitations that warrant acknowledgement. This is the first study to explore the associations between a select panel of genetic

variants to different types of injuries in elite AFL players. Another strength was the collection of injury incidence and severity data over 7 consecutive years using the AFL's highly standardised and reliable injury recording and reporting methods. However, our study's limitations include its small sample size for a genetic study, and the single elite AFL squad investigated, ensuring the results must be delimited to this one cohort. Age represents one important contributor to injury risk though it was unable to be evaluated in our statistical model. In addition, the panel of genes included in this study will not be reflective of all genetic variants that may be important, and thus, our results are also delimited to the gene candidates evaluated. Replication of the current study using as many elite AFL teams as possible (i.e. preferably competition-wide studies), and broader range of genetic variants of interest, would be recommended to maximise this line of investigation and seek to confirm our findings into elite-level male footballers and the genetic underpinnings of injury incidence and severity. Lastly, predisposition to, or associations with, injuries is likely to be highly complex and polygenic in nature; thus, future research could focus on the cumulative impact of genetic variation assessed through polygenic risk scores, such as the total genotype score (TGS) method [68]. Regardless, our preliminary study raises some intriguing insights into potential relationships among genotypes and its link to injury incidence and severity that warrants further investigation.

Conclusion

Several novel and significant associations were found during the current study. The rs12722 SNP within the *COL5A1* gene was significantly associated with all muscle injuries, as was *NOGGIN* rs1372857, while *COL5A1* was also significantly associated with contact bone injuries. The *IGF2* rs3213221 polymorphism was significantly associated with contact tendon injuries, while the *COL1A1* rs1800012 gene was significantly associated with all ligament injuries, with further associations with low severity injuries and non-contact ligament injuries. Several trends towards significance were observed between *CCL2* rs2857656 and *IGF2* rs3213221 for all muscle injuries, between *COL1A1* rs1800012 and contact muscle injuries, between *NOGGIN* rs1372857 and non-contact muscle injuries, and *COL12A1* rs970547 and all tendon and non-contact tendon injuries. Future research should expand the population pool by undertaking a competition-wide study and may investigate how such genetic variants could influence a person's injury rate when compared to playing position, physiological abilities, and training loads or exposure time (hours of training and competition) across a season. Future research

should replicate this work in elite female Australian Football (AFLW) population.

These findings present potential applications for developing training regimes around genetic predisposition. Players who are known to have a higher risk of injury due to underlying genetic susceptibilities could have more specific training based around their individual needs, including targeted and specific strength programs, and more scrutinised loading practises to reduce injury onset. For example, elite male Australian Football players with genotypes associated with potential susceptibility to bone-related injuries should be prioritised for (1) closer screening and ongoing monitoring of lower-body musculoskeletal morphology status and training adaptation through dual-energy X-ray absorptiometry (DXA) and peripheral quantitative computed tomography (pQCT) if available; (2) individualised load management practices with lower running volumes (relative to others) in favour of targeted lower-body mechanical loading programs honouring osteogenic principles of mechano-adaptation (i.e. high-magnitude, low-volume strength training, or multi-directional plyometric exercises) to optimise musculoskeletal cross-sectional area, promote skeletal robustness, and improve skeletal fatigue resistance [69–72]; and (3) nutritional review by a sports dietitian to evaluate energy availability, calcium, and vitamin D intake for potential supplementation in accordance with the Australian Football Anti-Doping Code (signatory to the World Anti-Doping Code, World Anti-Doping Agency (WADA)) [73–75].

Abbreviations

ACL: Anterior Cruciate Ligament; ACTN3: Alpha-Actinin-3; AF: Australian Football; AFL: Australian Football League; AFLW: Australian Football League Women; AGRF: Australian Genome Research Facility; BMP: Bone Morphogenetic Proteins; CCL2: Chemokine CC Motif Ligand-2; COL1A1: Collagen Type I Alpha 1; COL12A1: Collagen Type XII Alpha 1; COL5A1: Collagen Type V Alpha 1; DNA: Deoxyribonucleic Acid; DXA: Dual-energy X-ray Absorptiometry; EMILIN1: Elastin Microfibril Interface 1; HWE: Hardy–Weinberg Equilibrium; IGF2: Insulin-like Growth Factor-2; IRRs: Incident Rate Ratios; pQCT: Peripheral Quantitative Computed Tomography; SMAD6: SMAD Family Member 6; SNP: Single Nucleotide Polymorphism; TGS: Total Genotype Score; WADA: World Anti-Doping Agency; WAFL: Western Australian Football League.

Acknowledgements

Results of this study have been presented clearly, honestly, and without fabrication, falsification, or inappropriate data manipulation. The authors declare no conflicts of interest. The authors would also like to acknowledge the West Coast Eagle Football Club and their players for their participation in this research. The authors would like to also thank the Australian Football League for their approval and support for this research to occur.

Author Contributions

YJ, RSA, TS, and NHH conceptualised the research. YJ, TS, NHH, BR, and AJ collected the data. YJ, RSA, JLCW, DH, SML, and NHH conducted data analysis. DH provided biostatistical support to YJ and the study. All authors were involved in manuscript drafting, data interpretation, manuscript completion, and revisions during the review process.

Funding

No funding was received to complete this research. YJ is supported by an Australian Government Research Training Scholarship.

Availability of Data and Materials

Raw datasets generated and analysed during this study are not publicly available due to agreement with the football club, and to protect the confidentiality and individual privacy of the athlete participants within the elite football club.

Declarations

Ethics Approval and Consent to Participate

Data collection and management procedures conformed to the Declaration of Helsinki (World Medical Association). Human Research Ethics Approval was provided by the Edith Cowan University Human Research and Ethics Committee (ID: 2019-00181-JACOB). All participants provided written informed consent to participate in this study.

Consent for Publication

Not applicable.

Competing interests

BR is an employee of the West Coast Eagles Football Club, and AJ was an employee of the West Coast Eagles Football Club at the time of data collection and analysis. No other financial or non-financial competing interests are declared by any authors.

Author details

¹School of Medical and Health Sciences, Edith Cowan University, Perth, WA, Australia. ²Institute for Health Research, University of Notre Dame Australia, Perth, WA, Australia. ³School of Health Science, University of Notre Dame Australia, Perth, WA, Australia. ⁴Exercise Medicine Research Institute, Edith Cowan University, WA, Perth, Australia. ⁵West Coast Eagles Football Club, Perth, WA, Australia. ⁶Centre for Precision Health, Edith Cowan University, Perth, WA, Australia. ⁷Collaborative Genomics and Translation Group, School of Medical and Health Sciences, Edith Cowan University, Perth, WA, Australia. ⁸School of Pharmacy and Biomedical Sciences, Faculty of Health Sciences, Curtin Health Innovation Research Institute, Curtin University, Perth, WA, Australia. ⁹Caring Futures Institute, College of Nursing and Health Sciences, Flinders University, Adelaide, SA, Australia. ¹⁰Faculty of Health, School of Nursing, Queensland University of Technology, Brisbane, QLD, Australia.

Received: 21 March 2022 Accepted: 29 September 2022

Published online: 11 October 2022

References

- Boyd LJ, Ball K, Aughey RJ. Quantifying external load in Australian football matches and training using accelerometers. *Int J Sports Physiol Perform.* 2013;8(1):44–51.
- Coutts AJ, Quinn J, Hocking J, Castagna C, Rampinini E. Match running performance in elite Australian Rules Football. *J Sci Med Sport.* 2010;13(5):543–8.
- Gray AJ, Jenkins DG. Match analysis and the physiological demands of Australian football. *Sports Med.* 2010;40(4):347–60.
- Wisbey B, Montgomery PG, Pyne DB, Rattray B. Quantifying movement demands of AFL football using GPS tracking. *J Sci Med Sport.* 2010;13(5):531–6.
- Colby MJ, Dawson B, Heasman J, Rogalski B, Rosenberg M, Lester L, et al. Preseason workload volume and high-risk periods for noncontact injury across multiple Australian football league seasons. *J Strength Cond Res.* 2017;31(7):1821–9.
- Hart NH, Nimphius S, Cochrane JL, Newton RU. Leg mass characteristics of accurate and inaccurate kickers: an Australian football perspective. *J Sports Sci.* 2013;31(15):1647–55.

7. Hart NH, Nimphius S, Spiteri T, Cochrane JL, Newton RU. Relationship between Leg Mass, Leg Composition and Foot Velocity on Kicking Accuracy in Australian Football. *J Sports Sci Med*. 2016;15(2):344–51.
8. Hart NH, Nimphius S, Spiteri T, Newton RU. Leg strength and lean mass symmetry influences kicking performance in Australian football. *J Sports Sci Med*. 2014;13(1):157–65.
9. Hart NH, Nimphius S, Weber J, Spiteri T, Rantalainen T, Dobbin M, et al. Musculoskeletal asymmetry in football athletes: a product of limb function over time. *Med Sci Sports Exerc*. 2016;48(7):1379–87.
10. Hart NH, Spiteri T, Lockie RG, Nimphius S, Newton RU. Detecting deficits in change of direction performance using the preplanned multidirectional Australian football league agility test. *J Strength Cond Res*. 2014;28(12):3552–6.
11. Saw R, Finch CF, Samra D, Baquie P, Cardoso T, Hope D, et al. Injuries in Australian rules football: an overview of injury rates, patterns, and mechanisms across all levels of play. *Sports Health*. 2018;10(3):208–16.
12. Eirale C, Tol JL, Farooq A, Smiley F, Chalabi H. Low injury rate strongly correlates with team success in Qatari professional football. *Br J Sports Med*. 2013;47(12):807–8.
13. Collins M, Raleigh SM. Genetic risk factors for musculoskeletal soft tissue injuries. *Med Sport Sci*. 2009;54:136–49.
14. McCabe K, Collins C. Can genetics predict sports injury? The Association of the genes GDF5, AMPD1, COL5A1 and IGF2 on soccer player injury occurrence. *Sports*. 2018;6(1):21.
15. McCall A, Carling C, Davison M, Nedelec M, Le Gall F, Berthoin S, et al. Injury risk factors, screening tests and preventative strategies: a systematic review of the evidence that underpins the perceptions and practices of 44 football (soccer) teams from various premier leagues. *Br J Sports Med*. 2015;49(9):583–9.
16. Baumert P, Lake MJ, Stewart CE, Drust B, Erskine RM. Genetic variation and exercise-induced muscle damage: implications for athletic performance, injury and ageing. *Eur J Appl Physiol*. 2016;116(9):1595–625.
17. Kim JH, Jung ES, Kim CH, Youn H, Kim HR. Genetic associations of body composition, flexibility and injury risk with ACE, ACTN3 and COL5A1 polymorphisms in Korean ballerinas. *J Exerc Nutr Biochem*. 2014;18(2):205–14.
18. Maciejewska-Skrendo A, Leznicka K, Leonska-Duniec A, Wilk M, Filip A, Cieszczyk P, et al. Genetics of muscle stiffness, muscle elasticity and explosive strength. *J Hum Kinet*. 2020;74:143–59.
19. Karki R, Pandya D, Elston RC, Ferlini C. Defining, “mutation” and “polymorphism” in the era of personal genomics. *BMC Med Genom*. 2015;8:37.
20. Kostek M, Hubal MJ, Pescatello LS. The role of genetic variation in muscle strength. *Am J Lifestyle Med*. 2011;5(2):156–70.
21. Pickering C, Kiely J. ACTN3: more than just a gene for speed. *Front Physiol*. 2017;8:1080.
22. Yang N, MacArthur DG, Gulbin JP, Hahn AG, Beggs AH, Eastale S, et al. ACTN3 genotype is associated with human elite athletic performance. *Am J Hum Genet*. 2003;73(3):627–31.
23. Harmon BT, Orkunoglu-Suer EF, Adham K, Larkin JS, Gordish-Dressman H, Clarkson PM, et al. CCL2 and CCR2 variants are associated with skeletal muscle strength and change in strength with resistance training. *J Appl Physiol*. 2010;109(6):1779–85.
24. Iwao-Koizumi K, Ota T, Hayashida M, Yonetani Y, Nakata K, Kinoshita K, et al. The ACTN3 gene is a potential biomarker for the risk of non-contact sports injury in female athletes. *J Mol Biomark Diagn*. 2014;56(002):1–7.
25. Gibbon A, Raleigh SM, Ribbans WJ, Posthumus M, Collins M, September AV. Functional COL1A1 variants are associated with the risk of acute musculoskeletal soft tissue injuries. *J Orthop Res*. 2020;38(10):2290–8.
26. Pruna R, Artells R, Ribas J, Montoro B, Cos F, Munoz C, et al. Single nucleotide polymorphisms associated with non-contact soft tissue injuries in elite professional soccer players: influence on degree of injury and recovery time. *BMC Musculoskelet Disord*. 2013;14:221.
27. Stastny P, Lehnert M, De Ste CM, Petr M, Svoboda Z, Maixnerova E, et al. Effect of COL5A1, GDF5, and PPARA genes on a movement screen and neuromuscular performance in adolescent team sport athletes. *J Strength Cond Res*. 2019;33(8):2057–65.
28. John R, Prabhakar S, Dhillon MS, Anand A, Minhas G. Association of ACL tears and single nucleotide polymorphisms in the collagen 12 A1 gene in the Indian population: a preliminary case–control study. *Muscles Ligaments Tendons J*. 2016;6(2):253–7.
29. O’Connell K, Knight H, Ficek K, Leonska-Duniec A, Maciejewska-Karlowska A, Sawczuk M, et al. Interactions between collagen gene variants and risk of anterior cruciate ligament rupture. *Eur J Sport Sci*. 2015;15(4):341–50.
30. Urano T, Shiraki M, Usui T, Sasaki N, Ouchi Y, Inoue S. Bone mass effects of a Smad6 gene polymorphism in Japanese postmenopausal women. *J Bone Miner Metab*. 2009;27(5):562–6.
31. Dimitriou R, Carr IM, West RM, Markham AF, Giannoudis PV. Genetic predisposition to fracture non-union: a case control study of a preliminary single nucleotide polymorphisms analysis of the BMP pathway. *BMC Musculoskelet Disord*. 2011;12:44.
32. Devlin RD, Du Z, Pereira RC, Kimble RB, Economides AN, Jorgetti V, et al. Skeletal overexpression of noggin results in osteopenia and reduced bone formation. *Endocrinology*. 2003;144(5):1972–8.
33. Sun S, Wang Y, Wu Y, Gao Y, Li Q, Abdulrahman AA, et al. Identification of COL1A1 as an invasion-related gene in malignant astrocytoma. *Int J Oncol*. 2018;53(6):2542–54.
34. Laguette MJ, Abrahams Y, Prince S, Collins M. Sequence variants within the 3’-UTR of the COL5A1 gene alters mRNA stability: implications for musculoskeletal soft tissue injuries. *Matrix Biol*. 2011;30(5–6):338–45.
35. Hicks D, Farsani GT, Laval S, Collins J, Sarkozy A, Martoni E, et al. Mutations in the collagen XII gene define a new form of extracellular matrix-related myopathy. *Hum Mol Genet*. 2014;23(9):2353–63.
36. MacArthur DG, North KN. ACTN3: a genetic influence on muscle function and athletic performance. *Exerc Sport Sci Rev*. 2007;35(1):30–4.
37. Bergman D, Halje M, Nordin M, Engstrom W. Insulin-like growth factor 2 in development and disease: a mini-review. *Gerontology*. 2013;59(3):240–9.
38. Zhu S, Liu M, Bennett S, Wang Z, Pflieger KDG, Xu J. The molecular structure and role of CCL2 (MCP-1) and C-C chemokine receptor CCR2 in skeletal biology and diseases. *J Cell Physiol*. 2021;236(10):7211–22.
39. Litteri G, Carnevale D, D’Urso A, Cifelli G, Braghetta P, Damato A, et al. Vascular smooth muscle Emilin-1 is a regulator of arteriolar myogenic response and blood pressure. *Arterioscler Thromb Vasc Biol*. 2012;32(9):2178–84.
40. Zacchigna L, Vecchione C, Notte A, Cordenonsi M, Dupont S, Maretto S, et al. Emilin1 links TGF-beta maturation to blood pressure homeostasis. *Cell*. 2006;124(5):929–42.
41. Lim T, Santiago C, Pareja-Galeano H, Iturriaga T, Sosa-Pedreschi A, Fuku N, et al. Genetic variations associated with non-contact muscle injuries in sport: a systematic review. *Scand J Med Sci Sports*. 2021;31(11):2014–32.
42. Choi M, Stottmann RW, Yang YP, Meyers EN, Klingensmith J. The bone morphogenetic protein antagonist noggin regulates mammalian cardiac morphogenesis. *Circ Res*. 2007;100(2):220–8.
43. Galvin KM, Donovan MJ, Lynch CA, Meyer RI, Paul RJ, Lorenz JN, et al. A role for smad6 in development and homeostasis of the cardiovascular system. *Nat Genet*. 2000;24(2):171–4.
44. Miyamoto-Mikami E, Miyamoto N, Kumagai H, Hirata K, Kichuni N, Zempo H, et al. COL5A1 rs12722 polymorphism is not associated with passive muscle stiffness and sports-related muscle injury in Japanese athletes. *BMC Med Genet*. 2019;20(1):192.
45. Zhao D, Zhang Q, Lu Q, Hong C, Luo T, Duan Q, et al. Correlations between the genetic variations in the COL1A1, COL5A1, COL12A1, and beta-fibrinogen genes and anterior cruciate ligament injury in Chinese patients. *J Athl Train*. 2020;55(5):515–21.
46. Ficek K, Cieszczyk P, Kaczmarczyk M, Maciejewska-Karlowska A, Sawczuk M, Cholewinski J, et al. Gene variants within the COL1A1 gene are associated with reduced anterior cruciate ligament injury in professional soccer players. *J Sci Med Sport*. 2013;16(5):396–400.
47. Massidda M, Voisin S, Culigioni C, Piras F, Cugia P, Yan X, et al. ACTN3 R577X polymorphism is associated with the incidence and severity of injuries in professional football players. *Clin J Sport Med*. 2019;29(1):57–61.
48. Stepien-Slodkowska M, Ficek K, Eider J, Leonska-Duniec A, Maciejewska-Karlowska A, Sawczuk M, et al. The +1245g/t polymorphisms in the collagen type I alpha 1 (col1a1) gene in polish skiers with anterior cruciate ligament injury. *Biol Sport*. 2013;30(1):57–60.
49. McCaskie CJ, Sim M, Newton RU, Hart NH. Lower-limb injury in elite Australian football: a narrative review of kinanthropometric and physical risk factors. *Phys Ther Sport*. 2021;52:69–80.

50. Finch CF, Twomey DM, Fortington LV, Doyle TL, Elliott BC, Akram M, et al. Preventing Australian football injuries with a targeted neuromuscular control exercise programme: comparative injury rates from a training intervention delivered in a clustered randomised controlled trial. *Inj Prev*. 2016;22(2):123–8.
51. Stares J, Dawson B, Peeling P, Heasman J, Rogalski B, Drew M, et al. Identifying high risk loading conditions for in-season injury in elite Australian football players. *J Sci Med Sport Sports Med Aust*. 2018;21(1):46–51.
52. Jacob Y, Hart NH, Cochrane Wilke J, Spiteri T, Laws SM, Jones A, et al. ACTN3 (R577X) genotype is associated with Australian football league players. *J Strength Cond Res*. 2022;36(2):573–6.
53. Jacob Y, Anderton RS, Cochrane Wilkie JL, Rogalski B, Laws SM, Jones A, et al. Association of genetic variances in ADRB1 and PPARGC1a with two-kilometre running time-trial performance in Australian football league players: a preliminary study. *Sports*. 2021;9(2):22.
54. Jacob Y, Chivers P, Anderton RS. Genetic predictors of match performance in sub-elite Australian football players: a pilot study. *J Exerc Sci Fit*. 2019;17(2):41–6.
55. Massidda M, Bachis V, Corrias L, Piras F, Scorcu M, Calò CM. Influence of the COL5A1 rs12722 on musculoskeletal injuries in professional soccer players. *J Sports Med Phys Fitness*. 2015;55(11):1348–53.
56. Collins M, Posthumus M. Type V collagen genotype and exercise-related phenotype relationships: a novel hypothesis. *Exerc Sport Sci Rev*. 2011;39(4):191–8.
57. Kadler KE, Baldock C, Bella J, Boot-Handford RP. Collagens at a glance. *J Cell Sci*. 2007;120(Pt 12):1955–8.
58. Guo R, Ji Z, Gao S, Aizezi A, Fan Y, Wang Z, et al. Association of COL5A1 gene polymorphisms and musculoskeletal soft tissue injuries: a meta-analysis based on 21 observational studies. *J Orthop Surg Res*. 2022;17(1):129.
59. Pruna R, Ribas J, Montoro JB, Artells R. The impact of single nucleotide polymorphisms on patterns of non-contact musculoskeletal soft tissue injuries in a football player population according to ethnicity. *Med Clin*. 2015;144(3):105–10.
60. Del Coso J, Valero M, Salinero JJ, Lara B, Gallo-Salazar C, Areces F. Optimum polygenic profile to resist exertional rhabdomyolysis during a marathon. *PLoS ONE*. 2017;12(3): e0172965.
61. Posthumus M, September AV, Keegan M, O’Cuiineagain D, Van der Merwe W, Schweltnus MP, et al. Genetic risk factors for anterior cruciate ligament ruptures: COL1A1 gene variant. *Br J Sports Med*. 2009;43(5):352–6.
62. Khoschnau S, Melhus H, Jacobson A, Rahme H, Bengtsson H, Ribom E, et al. Type I collagen alpha1 Sp1 polymorphism and the risk of cruciate ligament ruptures or shoulder dislocations. *Am J Sports Med*. 2008;36(12):2432–6.
63. Clifford T, Ventress M, Allerton DM, Stansfield S, Tang JCY, Fraser WD, et al. The effects of collagen peptides on muscle damage, inflammation and bone turnover following exercise: a randomized, controlled trial. *Amino Acids*. 2019;51(4):691–704.
64. Dar QA, Schott EM, Catheline SE, Maynard RD, Liu Z, Kamal F, et al. Daily oral consumption of hydrolyzed type 1 collagen is chondroprotective and anti-inflammatory in murine posttraumatic osteoarthritis. *PLoS ONE*. 2017;12(4): e0174705.
65. Ekstrand J, Torstveit MK. Stress fractures in elite male football players. *Scand J Med Sci Sports*. 2012;22(3):341–6.
66. Fortington LV, Berry J, Buttifant D, Ullah S, Diamantopoulou K, Finch CF. Shorter time to first injury in first year professional football players: a cross-club comparison in the Australian Football League. *J Sci Med Sport Sports Med Aust*. 2016;19(1):18–23.
67. Nose-Ogura S, Yoshino O, Dohi M, Kigawa M, Harada M, Hiraike O, et al. Risk factors of stress fractures due to the female athlete triad: differences in teens and twenties. *Scand J Med Sci Sports*. 2019;29(10):1501–10.
68. Del Coso J, Salinero JJ, Lara B, Gallo-Salazar C, Areces F, Herrero D, Puente C. Polygenic profile and exercise-induced muscle damage by a competitive half-ironman. *J Strength Cond Res*. 2020;34(5):1400–8.
69. Hart NH, Nimphius S, Rantalainen T, Ireland A, Sifarakas A, Newton RU. Mechanical basis of bone strength: influence of bone material, bone structure and muscle action. *J Musculoskelet Neuronal Interact*. 2017;17(3):114–39.
70. Hart NH, Newton RU, Tan J, Rantalainen T, Chivers P, Sifarakas A, Nimphius S. Biological basis of bone strength: anatomy, physiology and measurement. *J Musculoskelet Neuronal Interact*. 2020;20(3):347–71.
71. Fortington LV, Hart NH. Models for understanding and preventing fractures in sport. In: Robertson GAJ, Maffuli N, editors. *Fractures in sport*. Champaign: Springer; 2020. p. 75–84.
72. Hart NH, Newton RU, Weber J, Spiteri T, Rantalainen T, Dobbin M, Chivers P, Nimphius S. Functional basis of asymmetrical lower-body skeletal morphology in professional Australian rules footballers. *J Strength Cond Res*. 2020;34(3):791–9.
73. Maughan RJ, Burke LM, Dvorak J, Larson-Meyer DE, Peeling P, Phillips SM, Rawson ES, Walsh NP, Garthe I, Geyer H, Meeusen R. IOC consensus statement: dietary supplements and the high-performance athlete. *Int J Sport Nutr Exerc Metab*. 2018;28(2):104–25.
74. Fredericson M, Kussman A, Misra M, Barrack MT, De Souza MJ, Kraus E, Koltun KJ, Williams NI, Joy E, Nattiv A. The male athlete triad: a consensus statement from the female and male athlete triad coalition. Part II: diagnosis, treatment, and return-to-play. *Clin J Sport Med*. 2021;31(4):349–66.
75. Australian Football League: Australian Football Anti-Doping Code, Version 2021.1. <https://www.afl.com.au/clubhelp/policies/member-protection-and-integrity/anti-doping-code> (2021). Accessed 15 Aug 2022.

Publisher’s Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Submit your manuscript to a SpringerOpen[®] journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► [springeropen.com](https://www.springeropen.com)