

Investigating the miRNA pathways contribution to intra-tumour heterogeneity in glioblastoma and RNA binding of isoforms of the miRNA effector protein Argonaute

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Doctor of Philosophy

under the supervision of Gyorgy Hutvagner, Daniel Catchpoole, and Jinyan Li

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Certificate of Original Authorship

I, Christopher Smith, declare that this thesis is submitted in fulfilment of the requirements for the award of Doctor of Philosophy, in the School of Biomedical Engineering, Faculty of Engineering and IT, at the University of Technology Sydney.

This thesis is wholly my own work unless otherwise referenced or acknowledged. In addition, I certify that all information sources and literature used are indicated in the thesis.

This document has not been submitted for qualifications at any other academic institution.

This research is supported by the Australian Government Research Training Program.

Signature:

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Impact of the COVID-19 Pandemic on this Thesis

Our original plan to investigate our first hypothesis included an experiment to generate single cell RNA and small RNA data in a heterogeneous cancer model such as glioblastoma or neuroblastoma. This data was to be used to address both aims by enabling a direct comparison between miRNA and RNA expression profiles in potential cancer subpopulations. However, several months into the COVID-19 pandemic we decided to exclude this experiment, which we deemed high risk due to the scale and ambition of the project as well as the challenges in collaborating cross-institution during this period. We had concerns that access to researchers and laboratories would be impaired and make completion of this project unfeasible. In response, we retained our original aims but had to rely exclusively on public data. We chose glioblastoma as a heterogenous cancer model as it is one of the most extensively sequenced cancers with expression data available for RNAs and small RNAs in bulk tumours and single cells.

We also planned and initiated an experiment in 2021, approximately one year prior to my end of candidature. This involved collaboration with the Tumour Bank, part of The Children's Cancer Research Unit (CCRU) at The Children's Hospital at Westmead. This experiment intended to biologically validate observations of heterogeneity with miRNAs from the Dlk1-Dio3 locus in glioblastoma, by using a fluorescent in-situ hybridization assay (miRNAscope) for detection of miRNAs in formalin-fixed paraffin embedded glioblastoma samples. The project at the Tumour Bank was initially planned to begin late March in 2021. However, due to supplier errors and delays in shipping we were not able to receive all the required reagents to proceed with the experiment until June 2021, right before the second lockdown was instated in Sydney. The proximity of the outbreaks to Westmead hospital and the challenges in coordinating work with other lab members for assistance, access to facilities, and equipment installation, meant I was forced to delay this project until September 2021 with only a few months remaining before my final candidature assessment. While our preliminary results indicated that we were able to detect RNA using the control probes, the staining was inconsistent across cells, and we were unable to resolve this issue through optimization in the available timeframe.

We also had planned a second experiment in 2021, at the University of Technology, Sydney, to sequence Argonaute 2 using Nanopore sequencing for full length isoforms. This was intended to support the findings in chapter 4 where we used Capture-seq and short read sequencing to detect multiple isoforms of Argonaute 2 in human tissues and cell lines. To make this project feasible, we developed a modified library preparation protocol which could generate Argonaute 2 cDNA through targeted amplification. However, we were met with significant challenges with generating full length

amplicons of this gene, and because of constant delays throughout 2021 our timeline for both experiments clashed, and we chose to prioritise the miRNAscope project at the Tumour Bank.

Despite the challenges imposed by COVID-19 I was able to publish 3 first author papers – 1 review and 2 research papers.

Publications

Smith, C. M., Catchpoole, D. & Hutvagner, G. Non-Coding RNAs in Pediatric Solid Tumors. *Front. Genet.* **10**, 1–18 (2019).

Smith, C. *et al.* Cataloguing the small RNA content of honey using next generation sequencing. *Food Chem. Mol. Sci.* **2**, 100014 (2021).

Smith, C. M. & Hutvagner, G. A comparative analysis of single cell small RNA sequencing data reveals heterogeneous isomiR expression and regulation. *Sci. Rep.* **12**, 2834 (2022).

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Abstract

miRNAs are highly abundant small non-coding RNAs that are essential for post-transcriptional gene regulation. Many of the mechanisms which regulate miRNA expression and function are poorly understood. Their dysregulation is documented extensively in many diseases including cancer.

Glioblastoma is a highly aggressive and heterogeneous brain cancer that affects patients of all ages. Intra-tumoural heterogeneity describes the existence of genomically distinct subpopulations of tumour cells which can lead to differences in growth rate, metastatic potential, or vulnerability to certain treatments. The first part of my thesis investigates how miRNAs and miRNA variants (isomiRs) may be involved in intra-tumoural heterogeneity by applying bioinformatics analyses to single cell small RNA and RNA sequencing data generated from previous studies. This work identified two miRNA clusters, the Dlk1-Dio3 locus and miR-224/452, as potential contributors to intra-tumour heterogeneity in glioblastoma and may be involved in cell state regulation. Additionally, we found evidence of cell autonomous regulation and function of isomiRs, highlighting another regulatory mechanism that may play a role in heterogenous cancers. These miRNAs may have utility as cancer biomarkers and implicate a novel set of targets for therapeutic research.

The second part of my thesis investigates splice variants (isoforms) of Argonaute, an essential protein in the miRNA pathway that mediates their regulatory effects. Numerous Argonaute isoforms have been previously annotated, with alterations in protein domains critical for miRNA binding and function. However, current studies base their assumptions of miRNA activity through a single variant of Argonaute and the consequence of these alterations and their biological relevance has not been investigated yet. We identified two variants of Argonaute with altered miRNA binding characteristics that are variably expressed in normal and cancerous cells, revealing a novel form of miRNA regulation that could also have implications in cancer research.