Emerging diseases in Australian oysters and the challenges of climate change and uncertain futures

- Running head: Climate change and disease in Australian oysters
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Abstract

- Oysters are a valuable and iconic seafood, deeply rooted in Australian culture. However,
- oysters have always been vulnerable to disease, with disease outbreaks leading to mass
- mortality events that regularly cost the oyster aquaculture industry millions of dollars and
- affect livelihoods. Notably, there is evidence that climate change is rapidly causing the
- 20 emergence of new disease alongside the amplification of impacts of existing diseases. This is
- 21 because warming, acidification and freshening of coastal and estuarine habitats is affecting
- 22 the three axes of disease; the host, the external environment and the pathogens. Here we

explore how climate change is likely to impact all three axes and disease in Australian oyster aquaculture. Climate change is affecting oyster physiology, leading to weaker immune defences that allow for increased susceptibility to viral and bacterial infections. For example, there is evidence that recent heavy rain events precede oyster disease in estuaries. In addition, climate change is increasing the abundance and virulence of bacterial and viral pathogens, potentially resulting in the introduction of novel disease into new habitats. In order to remain viable, we suggest that the Australian oyster industry needs to enhance selective breeding programs currently underway with a diversification of products and research on emerging diseases to ensure resilience in the sector.

Keywords: Oysters; Disease; Climate change; aquaculture; Vibrio; Bacteria

Background

Oysters are a valuable and iconic seafood, deeply rooted in Australian culture. Historically, oysters were a valued food source for Indigenous Australians and the early colonies following European settlement of Australia (Nell 2001). In recent years, oyster aquaculture has employed thousands of Australians in regional areas, with approximately 1,500 people employed in New South Wales (NSW) alone, where the Sydney rock oyster *Saccostrea glomerata* is the primary product (Barclay *et al.*, 2016). Importantly, oyster aquaculture particularly creates employment opportunities for young people in many Australian coastal towns (Pierce and Robinson 2013; Barclay *et al.*, 2016; ABARES 2021). Australia-wide, oyster products are valued at \$145 million (ABARES 2012), with the Pacific oyster *Crassostrea gigas* forming the basis of oyster production in Tasmania and South Australia. Beyond the product value, oyster production provides valuable secondary benefits such as employment for regional communities (Pierce and Robinson 2013). For example, Oyster aquaculture

products in NSW, mainly S. glomerata, are valued at over \$50 million annually but with a 46 total value of ~\$300 million when the social and economic benefits are factored into 47 calculations (Barclay et al., 2016). 48 49 Oysters are farmed in estuaries which are among the most urbanised and developed 50 ecosystems in Australia. Over 85% of Australians live within 50 kms of the ocean, and nine 51 of the ten largest cities built on estuaries. Concerningly, climate change, is also impacting these ecosystems, causing estuaries to warm and acidify faster than the oceans (Scanes et 52 al. 2020) with a range of consequences for oysters. Oysters and oyster aquaculture are 53 54 commonly located in estuaries which also provide habitat for commercial fish species, and function to store blue carbon among other ecological services (Fodrie et al. 2017). 55 South eastern Australia (NSW, Victoria and Tasmania) is currently the breadbasket of 56 57 Australia's oyster production. It is also this region that will experience the greatest effects of climate change as the ocean warming "hotspot" in the Tasman sea warms at a rate faster 58 than the global average (Pörtner et al., 2022). Australia's oyster production is at the global 59 60 forefront of climate change and provides an opportunity to learn and apply new knowledge regarding disease and climate change to other regions in Australia and around the globe. 61 Disease has been a major factor causing the decline in oyster aquaculture from the 1970's. 62 NSW oyster production peaked in 1975 at 14 million dozen year⁻¹ (Raftos et al. 2014), but 63 this number has since decreased to 5.8 million dozen year-1 in 2019-2020 (NSW DPI 64 Production Reports 2020). Recent declines have also occurred in Tasmania and South 65 Australia, where between 2008 and 2020 production fell by 750,000 dozen, and 3,350,000 66 dozen in these states respectively (ABARES 2021). While these declines can be attributed to 67

different causes, a range of diseases have afflicted both S. glomerata and C. gigas since the

1970s with at least one of these diseases arriving on Australian shores in the last 50 years (Nell 2001). Climate change is predicted to intensify the pressure on oyster farming by warming and acidifying coastal habitats (Scanes *et al.*, 2020), leading to increasing disease events (King *et al.* 2019). Disease occurs as a result of the interactions among host, pathogen and environment, and this disease "triangle" has been used as conceptual model to understand the triggers of disease outbreaks (Scholthof 2007). Here we apply the disease triangle conceptual approach to explore how climate change will impact on the three axes of disease in Australian oyster aquaculture; host, environment and pathogen.

Current and emerging disease in Australian oysters

Crassostrea gigas

The Pacific oyster *Crassostrea* (alternatively named *Magallana*) *gigas* is the most intensely farmed oyster species in the world. *C. gigas* has recently been afflicted by the Pacific Oyster Mortality Syndrome (POMS), which is a disease triggered by the osterid herpes virus OsHV-1, and has devastated the global aquaculture industry, causing up to 35 % mortality on oyster farms in France each year (Fleury et al., 2020). This virus however, is not solely responsible for oyster death, with POMS likely to result from a 'polymicrobial infection'. Research has shown that the OsHV-1 virus supresses the *C. gigas* immune system, allowing bacterial infections, by members of the *Vibrio* genus, to follow. These bacterial infections are then responsible for oyster death, rather than the OsHV-1 virus itself (De Lorgeril *et al.* 2018).

Bacterial infections are common in *C. gigas* and are suspected to be responsible for other diseases. Among these, the disease called "summer mortality" causes significant *C. gigas* mortality in aquaculture during the summer months (King *et al.* 2019). However, until

recently the aetiological agent has remained elusive. It is now suspected that summer mortality is caused by a weakening of the immune system caused by heat stress, followed by bacterial infection and oyster death (Siboni et al. Submitted, King et al. 2019). In-line with this hypothesis, oyster death following marine heatwaves has been shown to be due to bacterial infections, rather than thermal stress to the oyster (Green et al. 2019). In many cases where bacterial infections cause oyster death, the bacteria responsible are suspected to be members of the Vibrio genus. Vibrio species are commonly implicated in C. gigas disease dynamics (Destoumieux-Garzón et al. 2020). Vibrio pathogens have been identified across C. gigas life stages including: V. alginolyticus, V. tubiashii, V. anguillarum, V. parahaemolyticus and V. harveyi (Go et al. 2017; Kirs et al. 2011; Lemire et al. 2014; Tubiash et al. 1970). In a recent study (Siboni et al. Submitted) total Vibrio abundance as well as the relative abundance of V. owensii, V. brasiliensis, V. jasicida and V. harveyi was significantly elevated when oyster mortality was greatest. V. owensii represented the principal driver of Vibrio communities and was also the most dominant species in dead oysters. Progressive increases of relative abundance were also observed for V. harveyi and V. campbellii when oyster mortality occurred. All three Vibrio species are known pathogens of marine organisms (Siboni et al. Submitted). In most cases where Vibrio have been implicated in the death of oysters, habitat warming has also occurred (Green et al. 2019; Petton et al. 2013). Analysis of C. gigas gene expression patterns have shown a strong interplay among gene expression, the microbiome and temperature (Scanes et al. Submitted) and Vibrio are known to proliferate in warming conditions (Baker-Austin et al. 2013; Vezzulli et al. 2015). Vibrio infection may be further

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aided when oysters experience a weakened immune system at elevated temperatures (Green et al. 2019).

Saccostrea glomerata

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Disease and climate change will also impact on the Sydney rock oyster, S. glomerata. This species is cultured largely on the east coast of Australia, and is responsible for 81% of oyster aquaculture in the state of NSW, where it forms the basis of a culturally iconic industry. S. glomerata is also afflicted by diverse diseases, with evidence that their impact may also be exacerbated by climate change (Scanes et al. 2021b). The main diseases currently impacting S. glomerata are Winter Mortality and QX disease. Winter mortality is a disease mostly recorded within southern estuaries in NSW and strikes during periods of cold weather (Nell 2001). For these reasons winter mortality may become less prevalent as climate change warms NSW estuaries. The impacts of QX disease, however, may be exacerbated. This disease called "Queensland unknown" or QX was first reported in the 1970's and precipitated a significant contraction of the S. qlomerata industry in New South Wales and Queensland (Nell 2001). QX disease is caused by the haplosporidian protozoan Marteilia sydneyi, which invades the oyster's digestive tract and causes starvation or secondary infection (Nell 1993). QX infection functions by supressing the oyster immune system allowing for infection by opportunistic microbes (Peters and Raftos 2003). Wet weather events during 2021 and 2022 led to record outbreaks of QX disease amongst the Sydney rock oyster, pushing the oyster industry to the brink of collapse in some regions (NSW DPI 2022). Notably, QX disease is now present in regions that had previously not experienced disease outbreaks including Port Stephens, NSW. Research has shown that periods of low salinity can compromise the immune defence of S. glomerata allowing for M.

sydneyi infection (Butt and Raftos 2007; Butt et al. 2006). There is evidence that this compromised immunity may be triggered during periods of low salinity by the suppression of the phenoloxidase system, which can then compromise the immune process (Peters and Raftos 2003).

Selective breeding of *S. glomerata* has significantly improved survivability to QX infection;

wild genotype survival rates are often reported as 10% following QX infection, while selected oysters typically can have 70% survival following infection (Dove *et al.* 2013). While this is a significant improvement, selected oysters must be produced in a hatchery setting, thereby limiting the number of oysters available and increasing the price of juvenile oysters. For these reasons the uptake of the selected oysters has been limited in areas where QX has previously not been an issue, leaving farmers vulnerable.

Winter mortality and QX are but two of a range of diseases recorded in *S. glomerata*, with disseminating hemocytic neoplasia, Steinhausia-like infections, Rickettsia-like infections and digenean trematodes having also been observed (Anderson 1996; Green *et al.* 2008). To date these pathogens and parasites have not been responsible for wide-spread commercial losses, but, this may change with altered environmental conditions. Disconcertingly, these diseases were all observed at the northern, warmer extent of the range of *S. glomerata*, which raises the possibility of their southward spread as warming occurs.

Ostrea angasi

The "Native", or "Flat" oyster, *Ostrea angasi*, is found around southern Australia and was once the basis of a significant wild fishery (Nell 2001). Today it is cultivated in small numbers, mostly in NSW, but attempts to culture the species have occurred in Tasmania, Victoria, and southern Western Australia. *Ostrea angasi* displays fast growth rates and can

tolerate warming (Pereira *et al.,* 2020). The greatest impediment to its production has been sporadic outbreaks of the haplosporidian parasite *Bonamia ostreae* that causes the disease Bonamiosis. However, this pathogen is largely found at the northern extent of its range in NSW estuaries (Heasman *et al.* 2004), and few mortalities have been associated with its presence.

Effects of Climate change on the host (oyster)

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Research has shown that warming and acidification of estuaries caused by climate change (Scanes et al. 2020) can significantly affect both larval and adult Sydney rock and Pacific oysters (Parker et al. 2013; Ross et al. 2011). Climate change has increased mortality and abnormality of larvae and decreased growth (Cole et al. 2016; Kurihara et al. 2007; Parker et al. 2010). Climate change can lead to decreased shell growth of adult oysters and acidified internal body spaces and altered metabolic rates (Lannig et al. 2010; Parker et al. 2011; Scanes et al. 2017; Wright et al. 2014). These impacts on the physical and physiological characteristics may translate into increased disease for oysters. Disease in oysters is suspected to be the result of change in the environment, which shift the total community of microbes in oysters, known as the microbiome. Microbiomes while vital to the well-being of all organisms, are comprised of both harmful and beneficial bacteria (Wendling et al. 2017). Changes to the oyster microbiome alter the oyster immune response (King et al. 2019), subsequently triggering opportunistic pathogens to infect oysters (Lokmer and Wegner 2015). Only recently has research begun to investigate how warming and acidification can interact with the microbiome of oysters. Warming and ocean acidification can alter the bacterial

communities of oyster species including the Sydney rock oyster, S. glomerata, however,

responses of the microbiome are heterogenous among S. glomerata genotypes. For example, it has been shown that the impacts of warming and ocean acidification can interact to significantly alter the bacterial community associated with S. glomerata when sourced from wild oysters (i.e. not selectively bred for aquaculture) (Scanes et al. 2021b). Ocean acidification increased bacterial diversity, but these increases were offset by a loss of diversity under warming (Scanes et al. 2021b). Oysters sourced from aquacultural genetic lineages, however, did not experience such significant changes. Rather, the microbiome of only two of the nine tested genetic lineages was significantly affected by either warming or acidification (Scanes et al. 2021c). Genotypic based responses to warming and acidification have previously been found in oysters. For example, ocean acidification had a heterogenous effect on the growth of S. glomerata across aquacultural genetic lineages (Parker et al. 2011). Similarly, the Standard Metabolic Rate (SMR) of Pacific oysters C. gigas was affected by ocean acidification, and the effect was dependant on genetic lineages. Some genetic lineages of C. gigas experienced an increase in SMR, and others a decrease (Wright et al. 2014). Recent evidence suggests that these genotype-based physiological responses may be driving the response of the microbiome. For example, bacterial communities were shown to strongly correlate with physiological traits, especially SMR (Scanes et al. 2021a). When warming and acidification caused a shift in oyster metabolic rates, this triggered a concurrent shift in their microbiome (Scanes et al. 2021a). These findings indicate that oyster physiological responses to climate change will likely have flow-on effects to their bacterial communities and subsequently to oyster survival.

Effects of climate change on the environment

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Oyster disease is often triggered by changes in their external environment. As estuarine organisms, oysters are faced with frequent environmental changes such as lowered salinity following rainfall. Climate change, however, is increasing the magnitude of these environmental changes. Back-to-back La Nina events in Eastern Australia have delivered record amounts of rainfall with Sydney recording the wettest year-to-date on record and caused much of Australia's east coast to be inundated by floods (BOM 2022). Oyster farming is an intensive anthropogenic practice that is incorporated within a functioning marine ecosystem. The oysters are therefore interacting and reacting with organisms in their environment. These interactions can have significant consequences for health and disease of oysters. For example, seagrass and macroalgae are often found in the estuarine and coastal ecosystems where oysters are cultured. Seagrasses can act as a microbial filter removing pathogens from the water column, significantly reducing the bacterial load faced by marine organisms (Lamb et al. 2017). Microbial filtering by aquatic vegetation can also reduce disease in oysters. The presence of red (Solieria) and brown algae (Fucus) significantly increased the survival of C. gigas when challenged with the OSHV-1 virus (Dugeny et al. 2022). This increased survival was suggested to be facilitated by stabilisation of the microbiome performed by the algae (Dugeny et al. 2022). Similar interactions between oyster and aquatic vegetation have also been observed in the Sydney Rock oyster S. glomerata. The microbiome of S. glomerata was significantly altered by an increased bacterial diversity in the presence of the seagrass Zostera muelleri (Garner et al. 2021). This relationship was, however, lost when oysters and seagrass were kept together at elevated pCO₂ conditions designed to reflect future (year 2100) atmospheric conditions (Garner et al. 2021).

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Climate change is predicted to trigger changes in marine ecosystem interactions (Cole *et al.* 2021), including the complete breakdown of ecosystem function in some extreme examples (Harley *et al.* 2017; Harley *et al.* 2006). An alteration or breakdown in current ecosystem interactions could have serious consequences for oyster disease. Already, we know that seagrasses and other aquatic vegetation are important in reducing disease, yet seagrasses are among the most threatened ecosystems on earth, with an annual rate of decline estimated at 7 % year⁻¹ (Waycott *et al.* 2009). The future loss of seagrasses and other key species from ecosystems will likely impact oyster health and exacerbate disease.

Effects of climate change on the pathogens

Most of the diseases afflicting *C. gigas* and *S. glomerata* in Australia have emerged in the last century. Some of these arrivals have been more recent than others, with OSHV-1 appearing within the last decade (Jenkins *et al.* 2013). As international shipping and trade into Australia continues to grow, the risk of marine bio-invasions increases. International shipping is the primary means by which goods are transported around the globe, and inbetween 2014 and 2018 ships made 34,117 port calls, an increase of 18.8% (BITRE 2020). Shipping is a major vector for marine bio-invasions with ballast water potentially containing a broad range of parasitic taxa, including many protistan and metazoan parasites (Pagenkopp Lohan *et al.* 2022). This ever-increasing ship traffic also poses the threat of oysters hitching a ride amongst fouling communities, and thereby introducing disease into new areas (Minchin and Gollasch 2003).

These risks are also likely to be intensified by climate change. The rapid warming of Australia's east coast is re-drawing the map of thermal niches resulting in range shifts across all levels of ecosystems (Vergés *et al.* 2019). For example, QX disease historically afflicted

QLD oyster growing areas before becoming apparent in northern NSW in the late 1970s, then occurring in the Georges and Hawkesbury rivers near Sydney in 1994 and 2004, respectively. Similarly, in the USA, a parasite *Perkinsus marinus* responsible for the oyster disease called Dermo was limited in its distribution by water temperatures, but, recent warming has permitted its spread throughout the Gulf of Mexico (Burge et al. 2014). Conceivably, the recent spread of QX disease may have been facilitated by the extreme warming of NSW estuaries experienced in the summers of 2018 and 2019 (Scanes et al. 2020), and may facilitate its range shift further south. Not all pathogens will benefit from climate change. Bonamia ostreae is a protozoan parasite responsible for Bonamiosis in flat oysters of the Ostrea genus, including the native flat oyster Ostrea angasi. This parasite appears to cause less disease in northern NSW estuaries, and research from Europe has shown that the protozoan has significantly lower survival at 25 °C compared to 15 and 4 °C (Arzul et al. 2009). It is, therefore, conceivable that warming oceans will not favour this parasite. Many oyster diseases are not fatal without the final blow of pathogenic and opportunistic bacteria (De Lorgeril et al. 2018; Green et al. 2019, Scanes et al., Submitted). Bacteria, including Vibrio, like most oceanic organisms are not homogenously distributed over the globe, but rather have distributions governed by temperature and other abiotic and biotic factors (Seymour 2022; Williams et al. 2022). Global climate change, such as the strengthening of the East Australian Current is likely to alter these distributions and trigger microbial range shifts (Messer et al. 2020), potentially exposing oysters to new and deadly bacterial pathogens. Climate change is predicted to further intensify heavy rainfall events in eastern Australia as the future climate will bring "more intense heavy rainfall events" (BOM

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2020). Short, but intense rainfall events trigger flooding and pulses of freshwater, leading to localised low salinity in estuaries, which can enhance the incidence of diseases such as QX (Raftos *et al.* 2014).

Solutions

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Increasing efforts are being made by the aquaculture industry and government to sustain oyster production. Selective breeding remains a popular tool that has significantly reduced the incidence of disease. For example, selective breeding of S. glomerata has significantly improved survivability to QX infection (Dove et al. 2013). Furthermore, selection of C. gigas to overcome OSHV-1 has also shown promising results, with resistant genotypes shown to be capable of maintaining a functional immune system in the face of infection (De Lorgeril et al. 2018). Oysters selected for disease resistance have also consistently shown resilience to warming and acidification (Parker et al. 2011), with their microbiome remaining stable (Scanes et al. 2021c). Transgenerational-plasticity (TGP) has also been shown to reduce the physiological effects of climate change, however, it remains unknown how this may translate to reduce disease (Ross et al. 2016). Despite the performance of selectively bred oysters, they are produced in a hatchery, which increases costs to farmers, and in some cases may be prohibitively expensive. Management actions will be required to curb the spread and potential risk of oyster diseases, particularly in the face of global climate change. Current protocols for the movement of oysters between estuaries will need to be frequently updated to keep pace with disease outbreaks. Diversification will also be key; too often the oyster industry has relied too heavily upon a single oyster species that has then suffered a significant disease event such as occurred in the Hawkesbury River in 2013 (ABC News 2013 "Shell Shocked").

Flat oysters *Ostrea angasi* and Akoya pearl oysters *Pinctada imbricata* both provide edible alternatives to *S. glomerata* and *C. gigas* and could help the industry to diversify when coupled with consumer awareness campaigns. Culturing oysters alongside seagrass or macroalgae in what is termed "multi-trophic aquaculture" has been shown to buffer some effects of climate change (Falkenberg *et al.* 2021) and may also help reduce the incidence of disease (Garner *et al.* 2021).

Conclusions

Climate change will present unique challenges for the culture of oysters in Australia and across the globe (Figure 1). This review has focused on disease, however, disease is only one of many stressors that will be exacerbated by climate change. Interactions among biological processes and the environment are complex, and must be considered if we are to gain a holistic understanding. Warming and acidification will combine with extreme events, including heavy rainfall and heatwaves to push oysters to their physiological limits, alter the microbiome and divert energy away from immune processes. The impacts of lowered immunity, which will often occur in synergy with the emergence of novel pathogens, will likely result in disease outbreaks and reduced oyster production, with a higher cost placed on the product for the consumer. Emerging diseases such as OSHV-1 and existing diseases like QX may also be exacerbated by secondary, opportunistic bacterial infections. Oyster farmers will likely need to be proactive if they are to successfully manage disease and maintain production under a future climate, however, this will potentially increase the cost of oysters for the consumer.

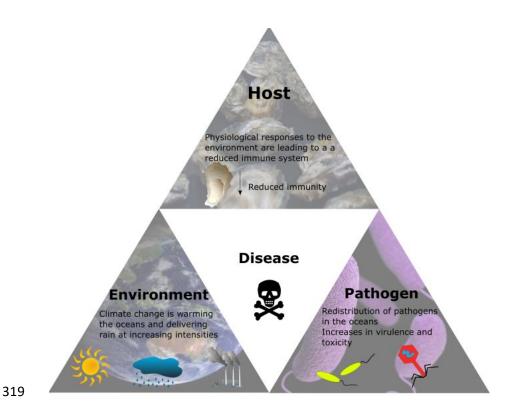


Figure 1. The effects of climate change on oyster production considered within the disease triangle conceptual approach.

References

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- ABARES. 2012. Australian Bureau of Agricultural and Resource Economics and Sciences.
- Fisheries and aquaculture statistics 2020. Accessed August 2022:
- 326 https://www.agriculture.gov.au/abares/research-topics/fisheries/fisheries-and-aquaculture-
- 327 <u>statistics</u>

329 **ABC News 2013.** Shell Shocked. Accessed August 2022:

330 https://www.abc.net.au/local/archives/landline/content/2013/s3712209.htm

331

332	Anderson, D.1. 1996. Atlas of invertebrate anatomy. Onsw Press, NSW, Australia
333	
334	Arzul, I., Gagnaire, B., Bond, C., Chollet, B., Morga, B., Ferrand, S., Robert, M. and Renault,
335	T. 2009 . Effects of temperature and salinity on the survival of Bonamia ostreae, a parasite
336	infecting flat oysters Ostrea edulis. <i>Diseases of aquatic organisms</i> 85 : 67-75.
337	
338	Baker-Austin, C., Trinanes, J.A., Taylor, N.G., Hartnell, R., Siitonen, A. and Martinez-Urtaza
339	J. 2013. Emerging Vibrio risk at high latitudes in response to ocean warming. Nature Climate
340	Change 3 : 73-77.
341	Barcley, K., McIlgorm, A., Mazur, N., Voyer, M., Schnierer, S., Payne, M. 2016. Social and
342	Economic Evaluation of NSW Coastal Aquaculture FRDC, Canberra, ACT. Acessed August
343	2022: https://www.uts.edu.au/sites/default/files/fass-report-social-economic-evaluation-
344	nsw-coastal-aquaculture.pdf
345	
346	BITRE (Bureau of Infrastructure and Transport Research Economics) 2020. Yearbook 2020:
347	Australian infrastructure statistics, BITRE, Canberra
348	
349	Burge, C.A., Mark Eakin, C., Friedman, C.S., Froelich, B., Hershberger, P.K., Hofmann, E.E.,
350	Petes, L.E., Prager, K.C., Weil, E. and Willis, B.L. 2014. Climate change influences on marine
351	infectious diseases: implications for management and society. Annual Review of Marine
352	Science 6 : 249-277.

Butt, D. and Raftos, D. 2007. Immunosuppression in Sydney rock oysters (Saccostrea 354 glomerata) and QX disease in the Hawkesbury River, Sydney. Marine and Freshwater 355 356 Research **58**: 213-221. 357 Butt, D., Shaddick, K. and Raftos, D. 2006. The effect of low salinity on phenoloxidase 358 activity in the Sydney rock oyster, Saccostrea glomerata. Aquaculture 251: 159-166. 359 360 Cole, V.J., Parker, L.M., O'Connor, S.J., O'Connor, W.A., Scanes, E., Byrne, M. and Ross, 361 P.M. 2016. Effects of multiple climate change stressors: ocean acidification interacts with 362 363 warming, hyposalinity, and low food supply on the larvae of the brooding flat oyster Ostrea angasi. Marine Biology 163: 1-17. 364 365 Cole, V.J., Parker, L.M., Scanes, E., Wright, J., Barnett, L. andRoss, P.M. 2021. Climate 366 change alters shellfish reef communities: A temperate mesocosm experiment. Marine 367 368 *Pollution Bulletin* **173**: 113113. 369 370 De Lorgeril, J., Lucasson, A., Petton, B., Toulza, E., Montagnani, C., Clerissi, C., Vidal-Dupiol, J., Chaparro, C., Galinier, R. and Escoubas, J.-M. 2018. Immune-suppression by 371 OsHV-1 viral infection causes fatal bacteraemia in Pacific oysters. Nature communications 9: 372 373 1-14.

375	Destoumieux-Garzón, D., Canesi, L., Oyanedel, D., Travers, M.A., Charrière, G.M., Pruzzo,
376	C. and Vezzulli, L. 2020. Vibrio-bivalve interactions in health and disease. Environmental
377	Microbiology 22 : 4323-4341.
378	
379	Dove, M.C., Nell, J.A. andO'Connor, W.A. 2013. Evaluation of the progeny of the fourth-
380	generation Sydney rock oyster Saccostrea glomerata (Gould, 1850) breeding lines for
381	resistance to QX disease (Marteilia sydneyi) and winter mortality (Bonamia roughleyi).
382	Aquaculture Research 44: 1791-1800.
383	
384	Dugeny, E., de Lorgeril, J., Petton, B., Toulza, E., Gueguen, Y. and Pernet, F. 2022.
385	Seaweeds influence oyster microbiota and disease susceptibility. Journal of Animal Ecology
386	91 : 805-818.
387	
388	Falkenberg, L.J., Scanes, E., Ducker, J. and Ross, P.M. 2021. Biotic habitats as refugia under
389	ocean acidification. Conservation Physiology 9 : coab077.
390	
391	Fodrie, F.J., Rodriguez, A.B., Gittman, R.K., Grabowski, J.H., Lindquist, N.L., Peterson, C.H.,
392	Piehler, M.F. andRidge, J.T. 2017 . Oyster reefs as carbon sources and sinks. <i>Proceedings of</i>
393	the Royal Society B: Biological Sciences 284 : 20170891.
394	

Garner, N., Ross, P.M., Falkenberg, L.J., Seymour, J.R., Siboni, N. and Scanes, E. 2021. Can seagrass modify the effects of ocean acidification on oysters? Marine Pollution Bulletin 177: Go, J., Deutscher, A.T., Spiers, Z.B., Dahle, K., Kirkland, P.D. and Jenkins, C. 2017. Mass mortalities of unknown aetiology in pacific oysters Crassostrea gigas in port stephens, New south wales, Australia. Diseases of aquatic organisms 125: 227-242. Green, T.J., Jones, B.J., Adlard, R.D. and Barnes, A.C. 2008. Parasites, pathological conditions and mortality in QX-resistant and wild-caught Sydney rock oysters, Saccostrea glomerata. Aquaculture 280: 35-38. Green, T.J., Siboni, N., King, W.L., Labbate, M., Seymour, J.R. and Raftos, D. 2019. Simulated marine heat wave alters abundance and structure of Vibrio populations associated with the Pacific Oyster resulting in a mass mortality event. *Microbial ecology* 77: 736-747. Harley, C.D., Connell, S.D., Doubleday, Z.A., Kelaher, B., Russell, B.D., Sarà, G. and Helmuth, B. 2017. Conceptualizing ecosystem tipping points within a physiological framework. *Ecology and evolution* **7**: 6035-6045.

416 Harley, C.D., Randall Hughes, A., Hultgren, K.M., Miner, B.G., Sorte, C.J., Thornber, C.S., Rodriguez, L.F., Tomanek, L. and Williams, S.L. 2006. The impacts of climate change in 417 418 coastal marine systems. Ecology Letters 9: 228-241. 419 420 Heasman, M., Diggles, B.K., Hurwood, D., Mather, P., Pirozzi, I. and Dworjanyn, S. 2004. 421 Paving the way for continued rapid development of the flat (angasi) oyster (Ostrea angasi) farming industry in New South Wales. Final Report to the Department of Transport & 422 423 Regional Services Project No. NT002/0195, NSW, Australia. 424 425 Jenkins, C., Hick, P., Gabor, M., Spiers, Z., Fell, S.A., Gu, X., Read, A., Go, J., Dove, M. and O'Connor, W. 2013. Identification and characterisation of an ostreid herpesvirus-1 426 microvariant (OsHV-1 μ-var) in *Crassostrea gigas* (Pacific oysters) in Australia. *Diseases of* 427 *aquatic organisms* **105**: 109-126. 428 429 King, W.L., Jenkins, C., Seymour, J.R. and Labbate, M. 2019. Oyster disease in a changing 430 431 environment: decrypting the link between pathogen, microbiome and environment. Marine 432 environmental research **143**: 124-140. 433 Kirs, M., Depaola, A., Fyfe, R., Jones, J.L., Krantz, J., Van Laanen, A., Cotton, D. and Castle, 434 M. 2011. A survey of oysters (Crassostrea gigas) in New Zealand for Vibrio parahaemolyticus 435 and Vibrio vulnificus. Int J Food Microbiol 147: 149-53. doi: 436 10.1016/j.ijfoodmicro.2011.03.012. 437

438	
439	Kurihara, H., Kato, S. and Ishimatsu, A. 2007. Effects of increased seawater pCO ₂ on early
440	development of the oyster <i>Crassostrea gigas</i> . Aquatic Biology 1 : 91-98.
441	
442	Lamb, J.B., Van De Water, J.A., Bourne, D.G., Altier, C., Hein, M.Y., Fiorenza, E.A., Abu, N.,
443	Jompa, J. and Harvell, C.D. 2017. Seagrass ecosystems reduce exposure to bacterial
444	pathogens of humans, fishes, and invertebrates. Science 355 : 731-733.
445	
446	Lannig, G., Eilers, S., Pörtner, H.O., Sokolova, I.M. and Bock, C. 2010. Impact of ocean
447	acidification on energy metabolism of oyster, Crassostrea gigas—changes in metabolic
448	pathways and thermal response. <i>Marine drugs</i> 8 : 2318-2339.
449	
450	Lemire, A., Goudenège, D., Versigny, T., Petton, B., Calteau, A., Labreuche, Y. and Le Roux
451	F. 2014. Populations, not clones, are the unit of vibrio pathogenesis in naturally infected
452	oysters. The ISME Journal 9. doi: 10.1038/ismej.2014.233.
453	
454	Lokmer, A. and Wegner, K.M. 2015 . Hemolymph microbiome of Pacific oysters in response
455	to temperature, temperature stress and infection. <i>The ISME journal</i> 9 : 670-682.
45.6	
456	
457	Minchin, D. and Gollasch, S. 2003. Fouling and Ships' Hulls: How Changing Circumstances
458	and Spawning Events may Result in the Spread of Exotic Species. <i>Biofouling</i> 19: 111-122.
459	doi: 10.1080/0892701021000057891.

460	
461	Nell, J.A. 1993. Farming the Sydney rock oyster (Saccostrea commercialis) in Australia.
462	Reviews in Fisheries Science 1: 97-120.
463	
464	Nell, J.A. 2001. The history of oyster farming in Australia. <i>Marine Fisheries Review</i> 63: 14-
465	25.
466	
	Nov. Courth Wales Deposition and of Drives are Industries 2020. As we call the production account
467	New South Wales Department of Primary Industries 2020. Aquaculture production report.
468	Accessed August 2022:
469	https://www.dpi.nsw.gov.au/ data/assets/pdf_file/0004/1389640/Aquaculture-
470	Production-Report-2020-2021.pdf
471	Pagenkopp Lohan, K.M., Darling, J.A. and Ruiz, G.M. 2022. International shipping as a
472	potent vector for spreading marine parasites. <i>Diversity and Distributions</i> 00 .
473	
4/3	
474	Parker, L.M., Ross, P.M., O'Connor, W.A., Pörtner, H.O., Scanes, E. and Wright, J.M. 2013.
475	Predicting the response of molluscs to the impact of ocean acidification. <i>Biology</i> 2 : 651-692.
476	
477	Parker, L.M., Ross, P.M. and O'Connor, W.A. 2010. Comparing the effect of elevated pCO ₂
478	and temperature on the fertilization and early development of two species of oysters.
479	Marine biology 157 : 2435-2452.
480	

481 Parker, L.M., Ross, P.M. and O'Connor, W.A. 2011. Populations of the Sydney rock oyster, 482 Saccostrea glomerata, vary in response to ocean acidification. Marine biology 158: 689-697. 483 484 Pereira, R.R., Scanes, E., Gibbs, M., Byrne, M. and Ross, P.M., 2020. Can prior exposure to stress enhance resilience to ocean warming in two oyster species?. PloS one, 15(4), 485 486 p.e0228527. 487 Peters, R. and Raftos, D.A. 2003. The role of phenoloxidase suppression in QX disease 488 489 outbreaks among Sydney rock oysters (Saccostrea glomerata). Aquaculture 223: 29-39. 490 491 Petton, B., Pernet, F., Robert, R. and Boudry, P. 2013. Temperature influence on pathogen transmission and subsequent mortalities in juvenile Pacific oysters Crassostrea gigas. 492 Aquaculture environment interactions **3**: 257-273. 493 494 Pierce, J. and Robinson, G., 2013. Oysters thrive in the right environment: the social 495 496 sustainability of oyster farming in the Eyre Peninsula, South Australia. Marine Policy, 37, 497 pp.77-85. 498 499 Pörtner, H.O., Roberts, D.C., Adams, H., Adler, C., Aldunce, P., Ali, E., Begum, R.A., Betts, R., Kerr, R.B., Biesbroek, R. and Birkman, J., 2022. IPCC, 2022: Climate Change 2022: 500 501 Impacts, Adaptation, and Vulnerability. Contribution of Working Group II to the Sixth

502	Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge, UK and
503	New York, NY, USA: Cambridge University Press.
504	
505	
506	Raftos, D.A., Kuchel, R., Aladaileh, S. and Butt, D. 2014. Infectious microbial diseases and
507	host defense responses in Sydney rock oysters. Frontiers in microbiology 5: 135.
508	
509	Ross, P.M., Parker, L. and Byrne, M. 2016. Transgenerational responses of molluscs and
510	echinoderms to changing ocean conditions. ICES Journal of Marine Science: Journal du
511	Conseil: fsv254.
512	
513	Ross, P.M., Parker, L., O'Connor, W.A. and Bailey, E.A. 2011. The impact of ocean
514	acidification on reproduction, early development and settlement of marine organisms.
515	Water 3 : 1005-1030.
516	
517	Scanes, E., Parker, L.M., O'Connor, W.A., Stapp, L.S. and Ross, P.M. 2017. Intertidal oysters
518	reach their physiological limit in a future high-CO2 world. Journal of Experimental Biology
519	220 : 765-774.
520	
521	Scanes, E., Parker, L.M., Seymour, J.R., Siboni, N., Dove, M.C., O'Connor, W.A. and Ross,
522	P.M. 2021a. Microbiomes of an oyster are shaped by metabolism and environment.
523	Scientific reports 11: 1-7.

524	
525	Scanes, E., Parker, L.M., Seymour, J.R., Siboni, N., King, W.L., Danckert, N.P., Wegner,
526	K.M., Dove, M.C., O'Connor, W.A. and Ross, P.M. 2021b. Climate change alters the
527	haemolymph microbiome of oysters. Marine Pollution Bulletin 164: 111991.
528	
529	Scanes, E., Parker, L.M., Seymour, J.R., Siboni, N., King, W.L., Wegner, K.M., Dove, M.C.,
530	O'Connor, W.A. and Ross, P.M. 2021c. Microbiome response differs among selected lines of
531	Sydney rock oysters to ocean warming and acidification FEMS Microbiology Ecology 97(8),
532	p.fiab099.
533	
534	Scanes, E., Scanes, P.R. and Ross, P.M. 2020. Climate change rapidly warms and acidifies
535	Australian estuaries. <i>Nature communications</i> 11 : 1803. doi: 10.1038/s41467-020-15550-z.
536	
537	Scanes, E., Siboni, N., Rees, B., and Seymour, J. Heatwave survival in intertidal species is
538	achieved by avoiding bacterial infection. Manuscript Submitted. Global Change Biology
539	
540	Scanes, E., Seymour, J., Giardina, M., Ugalde S., Wright, J., Crawford, C., Siboni, N.
541	Temporal patterns in the Pacific oyster <i>Crassostrea gigas</i> microbiome and gene regulation
542	Manuscript Submitted. Environmental Microbiology
543	
544	Scholthof, KB.G. 2007. The disease triangle: pathogens, the environment and society.
545	Nature Reviews Microbiology 5 : 152-156.

Seymour, J.R. 2022. Forecasting ocean microbiome shifts. *Nature Microbiology* **7**: 747-748. 547 548 Siboni, N., King, W. L., Williams, N. L.R., Scanes, E., Giardina, M., Green, T. J., Ostrowski, 549 M., O'Connor, W., Dove, M., Labbate, M., and Seymour J. Manuscript Submitted. ISME 550 551 **Communications** 552 553 Tubiash, H.S., Colwell, R.R. and Sakazaki, R. 1970. Marine Vibrios Associated with Bacillary 554 Necrosis, a Disease of Larval and Juvenile Bivalve Mollusks. *The Journal of Bacteriology* **103**: 555 271. 556 557 558 Vergés, A., McCosker, E., Mayer-Pinto, M., Coleman, M.A., Wernberg, T., Ainsworth, T. 559 and Steinberg, P.D. 2019. Tropicalisation of temperate reefs: Implications for ecosystem functions and management actions. Functional Ecology. 33 (6): 1000-1013 560 561 Vezzulli, L., Pezzati, E., Brettar, I., Höfle, M. and Pruzzo, C. 2015. Effects of global warming 562 563 on Vibrio ecology. *Microbiology spectrum* **3**: 3.3. 18. 564 Waycott, M., Duarte, C.M., Carruthers, T.J., Orth, R.J., Dennison, W.C., Olyarnik, S., 565 566 Calladine, A., Fourqurean, J.W., Heck, K.L. and Hughes, A.R. 2009. Accelerating loss of seagrasses across the globe threatens coastal ecosystems. Proceedings of the National 567 Academy of Sciences 106: 12377-12381. 568

570 Wendling, C.C., Fabritzek, A.G. and Wegner, K.M. 2017. Population-specific genotype x 571 genotype x environment interactions in bacterial disease of early life stages of Pacific oyster larvae. Evolutionary applications 10: 338-347. 572 573 574 Williams, N.L., Siboni, N., King, W.L., Balaraju, V., Bramucci, A. and Seymour, J.R. 2022. Latitudinal Dynamics of Vibrio along the Eastern Coastline of Australia. Water 14: 2510. 575 576 577 Wright, J.M., Parker, L.M., O'Connor, W.A., Williams, M., Kube, P. and Ross, P.M. 2014. 578 Populations of Pacific oysters Crassostrea gigas respond variably to elevated CO2 and 579 predation by Morula marginalba. The Biological Bulletin 226: 269-281.