

Review

Protective role of acetylcholine and the cholinergic system in the injured heart

Clara Liu Chung Ming,^{1,2} Xiaowei Wang,^{3,4,5,*} and Carmine Gentile^{1,2,*}

SUMMARY

This review explores the roles of the cholinergic system in the heart, comprising the neuronal and non-neuronal cholinergic systems. Both systems are essential for maintaining cardiac homeostasis by regulating the release of acetylcholine (ACh). A reduction in ACh release is associated with the early onset of cardiovascular diseases (CVDs), and increasing evidence supports the protective roles of ACh against CVD. We address the challenges and limitations of current strategies to elevate ACh levels, including vagus nerve stimulation and pharmacological interventions such as cholinesterase inhibitors. Additionally, we introduce alternative strategies to increase ACh in the heart, such as stem cell therapy, gene therapy, microRNAs, and nanoparticle drug delivery methods. These findings offer new insights into advanced treatments for regenerating the injured human heart.

INTRODUCTION

Cardiovascular diseases (CVDs) are a leading contributor to global mortality and morbidity, responsible for an estimated 17.9 million deaths each year, as reported by the World Health Organization.¹ In the early phases of CVD, an imbalance occurs within the autonomic system, marked by an increase in sympathetic activity and a decrease in the parasympathetic system. This disequilibrium is associated with an increase in norepinephrine and a reduction in the release of acetylcholine (ACh), contributing to higher rates of cardiac mortality and hindering myocardial regeneration.^{2–4} Early studies reported that an increase in sympathetic activation and parasympathetic inhibition in dogs leads to tachycardia-induced heart failure (HF).⁵ Additionally, Mahmoud et al.² demonstrated that inhibition of the cholinergic nerve during cardiac injury in neonatal mice and zebrafish leads to incomplete heart regeneration, causing a significant decrease in neonatal cardiac cell proliferation. In clinical trials, such as the Autonomic Tone and Reflexes After Myocardial Infarction study and Cardiac Insufficiency Bisoprolol Study II, reduced cardiac vagal activity is associated with increased heart rate and higher mortality rates in HF patients.^{6,7} In pre-clinical studies, Guimarães⁸ demonstrated that mice with a long-term cholinergic deficit exhibit increased norepinephrine levels and heightened sympathetic activity, resulting in cardiotoxicity. However, the correlation between CVD and autonomic dysfunction is still not well understood.

Several studies have reported that a decrease in ACh release, stemming from reduced cholinergic activity, is linked with various CVDs, including arrhythmias,⁹ atherosclerosis,^{10,11} myocardial infarction (MI),^{12,13} ischemic-reperfusion injury (I/R),^{14,15} doxorubicin (DOX)-induced cardiotoxicity,¹⁶ and HF.¹⁷ Additionally, increasing ACh secretion restores the imbalance of the autonomic system, improves heart rate variability, regulates mitochondrial function, suppresses reactive oxygen species (ROS) production, and alleviates inflammatory responses.^{8,18} Several studies have shown that the cholinergic system in the heart plays a pivotal role in guiding cardiac regeneration in the injured heart in mice, rats, rabbits, swine, and canine.^{19–23} Nonetheless, these pre-clinical findings have not been successfully translated to clinical studies, potentially due to the variability in ACh delivery methods across studies and, more broadly, to the multitarget effects of ACh in the human body. Therefore, a better understanding of the potential protective roles of ACh against myocardial injury, as well as the mechanisms regulating this process, will facilitate the development of new therapeutic targets against cardiac damage.

In this review, we focus on both the neuronal and cardiomyocyte cholinergic systems in the heart. Both systems are interconnected and essential for regulating the release of ACh, which binds to muscarinic and/or nicotinic ACh receptors to activate various signaling pathways, thereby maintaining cardiac homeostasis. We discuss the role of ACh derived from both systems in the heart and its therapeutic potential through muscarinic and/or nicotinic ACh receptors against CVD. The review also examines the limitations and constraints of current methods used to increase ACh levels in pre-clinical and clinical trials, such as vagus nerve stimulation (VNS) and cholinesterase inhibitors. Finally, we highlight alternative approaches to target the cholinergic system for more targeted and less invasive therapeutic strategies.

¹School of Biomedical Engineering, Faculty of Engineering and Information Technology, University of Technology Sydney, Sydney, NSW, Australia

²Cardiovascular Regeneration Group, Heart Research Institute, Newtown, NSW 2042, Australia

³Department of Medicine, Monash University, Melbourne, VIC 3800, Australia

⁴Department of Cardiometabolic Health, University of Melbourne, Melbourne, VIC 3010, Australia

⁵Molecular Imaging and Theranostics Laboratory, Baker Heart and Diabetes Institute, Melbourne, VIC 3004, Australia.

*Correspondence: xiaowei.wang@baker.edu.au (X.W.), carmine.gentile@uts.edu.au (C.G.)

<https://doi.org/10.1016/j.isci.2024.110726>



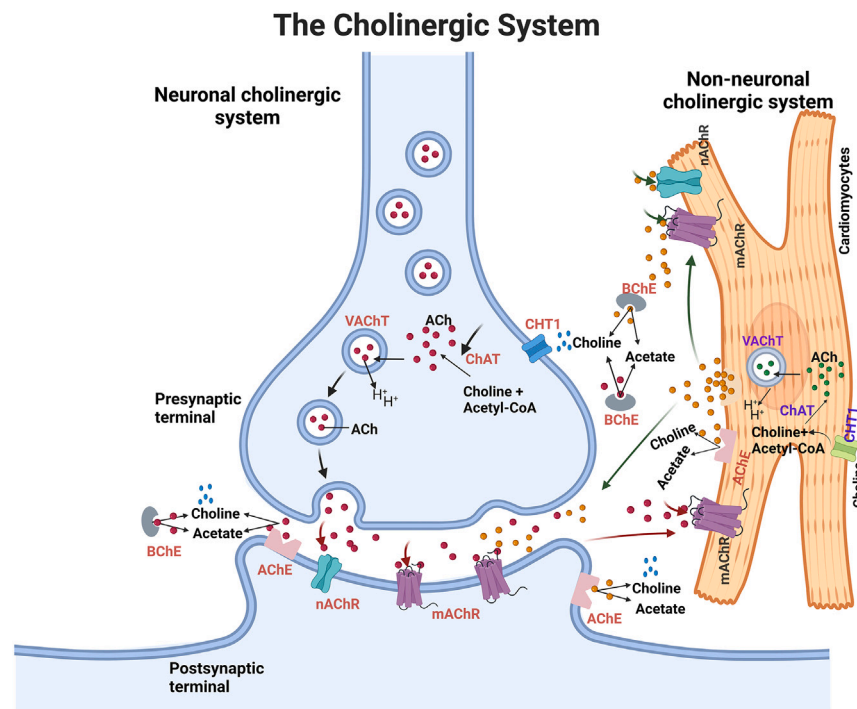


Figure 1. Schematic illustration of the cardiac neuronal cholinergic system in both cardiomyocytes and cholinergic nerves

Acetylcholine (ACh) (red) derived from the presynaptic terminal and ACh (orange) derived from cardiomyocytes are synthesized by choline acetyltransferase (ChAT) from the reaction between acetyl-CoA and choline. ACh from the neuronal and non-neuronal cholinergic system is stored by the vesicular acetylcholine transporter (VAcHT) and released upon stimulation. ACh released from both systems binds to ligand-gated ion channels nicotinic or G protein-coupled muscarinic ACh receptors (nAChRs and mAChRs, respectively). Acetylcholinesterase (AChE) present on postsynaptic terminal and cardiomyocytes and butyrylcholinesterase (BChE) present in the extracellular space degrade ACh to choline and acetate. High-affinity choline transporters (CHT1), present in cholinergic neurons and cardiomyocytes, are responsible for the reuptake of free choline for ACh synthesis.

THE CHOLINERGIC SYSTEM OF THE HEART

The autonomic system is composed of the sympathetic and parasympathetic nervous systems, which work in opposition to each other to balance heart activity. The parasympathetic nervous system, also known as the neuronal cholinergic system, encompasses molecules responsible for the synthesis, storage, release, signaling, and degradation of ACh, collectively regulating extracellular ACh concentrations within the presynaptic terminal.¹⁸ As a neurotransmitter, ACh is employed to modulate cardiac activity through muscarinic ACh receptors (mAChRs) to regulate heart dynamics⁹ and through nicotinic ACh receptors (nAChRs) to modulate inflammatory pathways and cardiac hemodynamic.²⁴

As outlined in Figure 1, ACh synthesis is catalyzed by choline acetyltransferase (ChAT), leading to its release and subsequent action in the cardiac extracellular space. The latter combines choline, supplied by a high-affinity choline transporter (CHT1), with acetyl-coenzyme A (CoA).^{18,25} ACh is stored within synaptic vesicles, which are acidified via an energy-dependent pump (H⁺-ATPase) and mediated via vesicular ACh transporter (VAcHT).²⁶ Upon depolarization, calcium influx triggers exocytosis, wherein ACh-filled synaptic vesicles fuse with the cellular membrane, releasing ACh in the synaptic cleft.⁴ Once released into the extracellular space, neuronal-derived ACh binds to mAChRs and nAChRs to activate various signaling pathways in the cardiovascular system.^{27–29} The degradation of ACh occurs rapidly by cholinesterases, including acetylcholinesterase (AChE) and butyrylcholinesterase (BChE). Both AChE and BChE hydrolyze ACh into choline and acetate.³⁰ CHT1 then transports choline to the presynaptic terminal.^{4,31}

Over the past two decades, a few studies have supported an additional intrinsic cholinergic machinery in heart cells, also known as the non-neuronal cholinergic system (NNCS) (which includes cardiomyocytes,¹⁴ endothelial cells,³² and leukocytes^{32,33}). Recently, Tarnawski et al.³² showed that ACh-derived T cells regulate vascular endothelial function and blood pressure via promoting endothelial nitric oxide synthase activity, vasorelaxation, and reducing vascular endothelial activation. Although leukocytes and endothelial cells have been shown to have the components of cholinergic machinery, their contribution to the total pool of ACh in the heart is low and remains unexplored.^{4,34}

In this review, we also focus on cardiomyocyte cholinergic machinery and the role played by cardiomyocyte-derived ACh in the heart. Cardiomyocytes can synthesize, transport, and release ACh, as they contain enzymes and transporters for ACh synthesis (ChAT), storage (VAcHT), degradation (AChE and BChE), and reuptake of choline for synthesis (CHT1) (Figure 1).^{35,36} Roy et al.³⁷ suggested that cardiomyocyte-derived ACh functions through similar second messenger systems and binds to mAChRs or nAChRs similar to neuronal-derived ACh. Additionally, studies have found that cardiomyocyte-derived ACh was detected intracellularly and extracellularly using cholinesterase inhibitors such as

donepezil, pyridostigmine, and physostigmine.^{35,36} Hence, the NNCS in cardiomyocytes comprises those components that regulate ACh homeostasis and act in an auto-/paracrine manner to mediate signaling.¹⁸ This concludes that both the neuronal cholinergic system and NNCS crosstalk with each other to mediate and regulate homeostasis heart activity.

THE ROLE OF CARDIOMYOCYTE-DERIVED ACh IN THE HEART

While both the neuronal cholinergic system and NNCS operate concurrently to regulate cardiac homeostasis, cardiomyocyte-derived ACh plays a crucial role in the heart and offers protection against CVD. ACh derived from cardiac NNCS sustains or enhances neuronal cholinergic effects, regulates heart rate, counteracts hypertrophic signals, maintains action potential propagation, and regulates cardiac energy metabolism.^{26,35,37–40} For instance, Roy et al.⁴⁰ found that lowering the levels of cardiomyocyte-derived ACh using ChAT and VAcHT knockout mice models resulted in increased heart rate and cardiovascular dysfunction. This suggests that a reduction in cardiomyocyte-derived ACh levels could potentially cause long-term changes in heart function, including ventricular cardiomyocyte hypertrophy, cardiac remodeling, and an increase in ROS.⁴⁰ Similar to neuronal-derived ACh, cardiomyocyte-derived ACh is crucial for maintaining cardiac homeostasis and regulating critical signaling pathways to maintain normal heart activity. Furthermore, Kakinuma et al.⁴¹ demonstrated that ChAT deletion in murine atrial myocardial cells increased oxygen consumption, mitochondrial activity, and reduction of ATP levels. As a result, cardiac NNCS plays a protective role in cardiomyocytes and the entire heart by maintaining physiological ATP levels and inhibiting oxygen consumption.⁴²

NNCS is also crucial for maintaining the balance between parasympathetic and sympathetic heart innervation and could amplify the protective effects of the parasympathetic nervous system.^{4,43} Few studies have shown that cardiomyocyte-derived ACh could modulate the central nervous system via the afferent vagal nerve, initiating crosstalk with other organs such as the brain, liver, and others.⁴⁴ Increasing cardiac NNCS signaling through a genetic approach and/or pyridostigmine, a cholinesterase inhibitor, leads to protective immunomodulatory effects, such as a reduction in CCL2/7 chemokines expression and a decrease of pro-inflammatory CCR2+ monocytes in the heart following cardiac injury.⁴⁵ This supports the cardioprotective role of cardiomyocyte-derived ACh via the modulation of the innate immune system.

Despite the fact that cardiac NNCS could be a potential target for therapeutic intervention, the detailed mechanisms that trigger the release of cardiomyocyte-derived ACh are still unclear. While Rocha-Resende et al.²⁶ proposed that adrenergic stimulation can induce cholinergic gene expression in cardiomyocytes, Roy et al.³⁷ suggested that cardiomyocyte-derived ACh may be regulated by sympathetic activity and the intrinsic cholinergic machinery, to regulate heart rate after stress and exercise. However, further studies are needed to identify the factors that activate NNCS in cardiomyocytes.

The role of cardiomyocyte-derived ACh against CVD

In this sub-section, we explore the potential therapeutic role of targeting cardiac NNCS and the protective role of cardiomyocyte-derived ACh against CVD. The current methods to increase cardiac NNCS activity are cholinesterase inhibitors, including pyridostigmine^{26,45} and donepezil,³⁵ small interfering RNA targeting AChE to increase ACh level,²⁶ and overexpression of ChAT gene^{42,46} or VAcHT gene.⁴⁵ Numerous studies have demonstrated that cardiac NNCS reduces oxygen demand and improves oxygen supply during ischemic injury, thereby preventing ischemia-induced cardiac dysfunction^{42,45} as well as preventing type 1 diabetes-induced heart diseases.⁴⁷

To elucidate the cardioprotective function of the NNCS in ischemic heart disease, Kakinuma et al.⁴² generated a heart-specific ChAT transgenic (ChAT-tg) mouse model that overexpresses ChAT in the heart to increase ACh synthesis. The findings revealed that cardiomyocyte-derived ACh plays an evident role in regulating myocardial energy metabolism through the activation of myocardial glucose utilization and angiogenesis in the infarcted area. Additionally, after 14 days, ChAT-tg mice exhibited increased resistance to MI and a higher survival rate compared to wild-type mice. This could be linked to the fact that cardiomyocyte-derived ACh activates hypoxia-inducible factor (HIF)-1 α , a non-hypoxic mechanism, and efficiently prevents energy depletion in the heart.⁴⁸

Moreover, cardiomyocyte-derived ACh increases cellular ATP levels. By monitoring ATP levels in real time, Oikawa et al.⁴⁹ demonstrated that insulin growth factor (IGF)-1R and Glut-1 protein expressions were upregulated together with an increase of ACh-derived cardiomyocytes, leading to a rise in glucose uptake and utilization. This mechanism preserves cellular ATP levels during oxidative stress and suppresses ROS production.¹⁴ Therefore, overexpression of cardiomyocyte ChAT activates cardiac ACh-HIF-1 α cascade and improves cell survival after myocardial I/R. Moreover, Kakinuma et al.⁴² found that upregulation of cardiomyocyte ChAT activates the cardiac ACh-HIF-1 α cascade to improve cells in ChAT-tg myocardial I/R mice. Hence, loss of cardiac ACh-HIF-1 α transcriptional pathway leads to cardiac dysfunction, which could be due to the synergistic effect of hypovascularity, calcium mishandling, and decreased myocardial energy.¹⁴

While the increase of sympathetic activity and norepinephrine is associated with arrhythmogenesis, cardiotoxicity, and impaired parasympathetic function, no link has been established between cardiomyocyte-derived ACh and cardiac norepinephrine levels.⁸ Consequently, the specific molecular mechanisms by which cardiomyocyte-derived ACh influences the cardiac cholinergic system remain unknown, underscoring the need for additional research. For instance, Kakinuma et al.³⁵ suggested that cardiomyocytes increase the transcriptional activity of the ChAT gene through mAChRs and ChAT protein expression to increase ACh levels in the cholinergic system. Furthermore, studies have shown that the NNCS enhances vagus nerve activity by increasing the release of nitric oxide from cardiomyocytes, contributing to beneficial cardiac effects^{42,44} as well as extracardiac effect.⁵⁰ Heart-specific ChAT-tg mice play a protective role in the central nervous system via the activation of the vagus nerve.⁴⁴ This pathway is important in the regulation of the inflammatory response in the blood-brain barrier (BBB), as well as in the response to restraint stress, less depressive-like and anxiety-like behaviors, and anti-convulsive effects.^{44,50} Therefore, it is crucial to conduct further investigations to fully understand the true clinical potential of targeting the cardiac NNCS and its effects on peripheral organs. Future research should also focus on determining the optimal strategies for enhancing NNCS-derived ACh levels in the heart.

CARDIOPROTECTIVE ROLES OF ACh VIA mAChRs AND nAChRs

Impairments in ACh signaling from both neuronal and NNCS sources can result in cellular death, heart dysfunction, and suppression of anti-inflammatory pathways, mediated by mAChRs and nAChRs.^{51,52} Stimulation of mAChRs, which are G protein-coupled receptors, leads to a decrease in heart rate and reduced cardiac conductivity.^{53,54} mAChRs regulate ventricular function both directly by counteracting β -adrenergic stimulation and indirectly by inhibiting L-type calcium channels.^{55,56} nAChRs are cholinergic ligand-gated ion channels permeable to Na^+ , K^+ , and Ca^{2+10} and have a central role in the cholinergic anti-inflammatory pathway by maintaining immune homeostasis through the activation and differentiation of immune T cells and reducing pro-inflammatory cytokines.⁵⁷ Moreover, the alpha 7 nicotinic ACh receptor subunit ($\alpha 7$ nAChR) regulates blood flow and cardiac hemodynamics²⁴ and enhances ACh release.^{58,59} While mAChRs and nAChRs trigger distinct signaling cascades across various cell types, they operate synergistically. Targeting both mAChRs and nAChRs through VNS or cholinesterase inhibitors improves left ventricular systolic function, prevents progressive left ventricular enlargement, and modulates the inflammatory response.^{52,60} The following sub-sections outline the cardioprotective effects of ACh in mitigating CVD through actions on mAChRs and nAChRs.

mAChRs

Type 2 muscarinic ACh receptors (M_2 AChRs) are the most commonly present receptor in the mammalian heart that activates several cardioprotective signaling pathways.⁶¹ Previous studies have shown that elevated ACh secretion in an injured heart promotes cardiomyocyte proliferation, suppresses ROS production, and prevents heart injury exacerbation via M_2 AChR.^{62–65} Activation of M_2 AChR also reduces mitochondrial oxidative damage in a DOX-induced rat model. This effect is mediated by Synapsin I, leading to heightened mitochondrial dynamics, while concurrently mitigating sympathetic activity and diminishing cardiac cell death, and necroptosis.^{16,66} ACh prevents cell apoptosis by inhibiting the action of angiotensin II (Ang II) and inhibits ROS production as well as cardiac hypertrophy by the activation of sirtuin 3/AMP-activated protein kinase (SIRT3-AMPK) signaling.^{67–69} ACh also prevents the progression of HF and cardiac remodeling through its inhibition of Ang II, improving survival rates in HF animal models, including mice, rats, and canines.^{9,39,70,71} Furthermore, ACh has the ability to activate superoxide dismutase, a crucial ROS-detoxifying enzyme in mitochondria and cytoplasm, which suppresses ROS production and protects against oxidative stress in I/R.^{72,73}

Several studies have indicated that an increase in ACh levels activates cell survival mechanisms in the heart against I/R.⁷⁴ ACh protects cardiomyocytes from ischemia through the transcription factor HIF-1 α and downstream gene expression for cell survival. However, further studies need to be performed to fully identify these signaling pathways.^{75,76} Additionally, type 3 muscarinic ACh receptor (M_3 AChR) is found abundantly in cardiac fibroblast and plays a critical role in fibroblast proliferation. The activation of M_3 AChR during cardiac fibrosis leads to the inhibition of the mitogen-activated protein kinase (MAPK) signaling pathway, including p38MAPK and ERK1/2 associated with reduced collagen production and cardiac fibrosis.⁷⁷

nAChRs

In the injured heart, ACh also binds to $\alpha 7$ nAChR on various cell types, including cardiomyocytes, monocytes, macrophages, endothelial cells, and others involved in immune responses,¹⁰ thereby regulating systemic inflammatory responses in CVD patients.⁷⁸ ACh inhibits the production of inflammatory cytokines such as tumor necrosis factor (TNF), interleukin (IL)-6, IL-1 β , and IL-18, which are responsible for activating T cells.^{4,21,66,79,80} Additionally, ACh stimulates macrophages through paracrine signaling, resulting in a reduction in the release of pro-inflammatory cytokines, the uptake of oxidized LDL, and the buildup of cholesterol within macrophages.^{10,81}

Through $\alpha 7$ nAChR, ACh attenuates endothelial dysfunction and promotes vasodilation, improving blood flow to the injured heart.⁶⁶ Li et al.⁸¹ showed that vascular injury in $\alpha 7$ nAChR knockout mice led to vascular remodeling, arterial inflammation, and chemokines induction and increased oxidative vascular stress. Activation of $\alpha 7$ nAChR can enhance tissue repair as well as reduce cardiac fibrosis damage.^{10,82} As cardiac fibrosis originates from endothelial cells through a process known as endothelial-to-mesenchymal transition,⁸³ $\alpha 7$ nAChR has the ability to significantly inhibit IL-1 β -induced endothelial-to-mesenchymal transition, nuclear factor κ B, and the induction of autophagy leading to the attenuation of cardiac fibrosis.^{13,76}

In addition, ACh plays a significant role in cardiac angiogenesis via $\alpha 7$ nAChR during cardiac hypertrophy and MI. Elevating ACh levels in MI mice enhances angiogenesis through vascular endothelial growth factor, leading to increased coronary arterial wall thickness and tube formation.^{68,84} Moreover, $\alpha 7$ nAChR activation also plays a critical role in attenuating ROS by promoting mitochondrial fusion via upregulation of mitofusins (1–2) and thus inhibiting DOX-induced autophagy.¹⁶

The cardioprotective function of ACh is intricate, involving various downstream pathways mediated by mAChRs and nAChRs to prevent cell death and avert cardiac dysfunction in the injured human heart. Thus, further research is required to identify and evaluate therapeutic approaches aimed at increasing ACh levels and enhancing the activation of mAChRs and nAChRs without adverse effects.

CHALLENGES AND CONSTRAINTS TO TRANSLATE CURRENT STRATEGIES TO CLINICAL STUDIES AND ALTERNATIVE APPROACHES

ACh has exhibited protective effects in various *in vivo* animal myocardial damage models. However, these benefits have been poorly translated into clinical trials. The primary challenge in translating ACh's attributes to clinical trials is its rapid hydrolysis in the presence of cholinesterase and the widespread distribution of ACh receptors throughout the body. Moreover, in 1985, Shepherd and Vanhoutte⁸⁵ found that

ACh can detect coronary artery spasms (CADs). The current method for identifying CAD in patients is through intracoronary injection of ACh (ACh-provocation test). This technique is highly sensitive and specific for detecting various types of CAD, including epicardial coronary spasm, microvascular spasm, microvascular dysfunction, and coronary stenosis.^{86,87} Abnormal vascular responses to ACh, such as ischemic electrocardiogram changes, may be a consequence of endothelial dysfunction and hyperconstriction of vascular smooth muscle. The ACh-provocation test holds potential for tailoring treatment for CAD patients, as it is considered safe with irreversible non-fatal complications. However, it is crucial to note that injecting large doses of ACh (>100 µg) into the right coronary artery of CAD patients may lead to bradycardia lasting up to 45 min^{88,89}

Several methods are commonly employed to elevate ACh levels in the neuronal cholinergic system, such as VNS via implanted stimulators or electrical stimulation and vagal efferent/afferent stimulation. Additionally, ACh levels can be increased in both the neuronal cholinergic system and NNCS through pharmacological interventions, including cholinesterase inhibitors. This section discusses current pre-clinical and clinical studies aimed at elevating ACh levels using VNS and cholinesterase inhibitors, examining their limitations and constraints, including the invasiveness of VNS and the adverse drug reactions (ADRs) associated with cholinesterase inhibitors. Furthermore, we explore potential alternative strategies to enhance ACh levels in the heart, including stem cell therapy, gene therapy, microRNA (miRNA) therapy, and the use of nanoparticles, highlighting the need for more targeted and less invasive approaches.

VNS

VNS is frequently used to elevate myocardial ACh levels within the neuronal cholinergic system, suggesting its potential as a therapeutic strategy to restore the autonomic balance and its anti-inflammatory effect.⁹⁰ Numerous studies have demonstrated that VNS reduces infarct size, improves ventricular function, attenuates ROS production, and decreases ventricular fibrillation in I/R animal models,^{74,75,91} in hypotension and HF models,⁹² as well for a DOX-induced rat model^{93,94} through mAChRs and nAChRs. Uitterdijk et al.⁹⁵ demonstrated that stimulating the vagus nerve in an ischemic swine model 5 min before reperfusion and continuing for 15 min post-reperfusion resulted in a significant reduction in infarct size. This intervention also led to a decrease in macrophages and neutrophils within the infarct area and mitigation of the no-reflow phenomenon through nAChRs.^{96,97} VNS exhibits protective effects against cardiomyocyte necrosis and microvascular obstruction, which could prevent reperfusion injury through both muscarinic and nicotinic pathways.^{65,98} Furthermore, Li et al.⁹⁹ demonstrated that 6 weeks of VNS in HF mice prevented long-term remodeling and improved overall survival.

Despite the evident promise of VNS as a therapeutic strategy against I/R, Buchholz et al.²³ showed that continuous VNS for 10 min before ischemia significantly increased the infarct size in a rabbit MI heart model. This opposite effect to what is reported earlier could be due to differences in VNS protocols and model species.¹⁰⁰ Moreover, implanting electrodes to stimulate VNS is invasive, and the optimal electrical dosage remains uncertain.¹⁰¹ For instance, Sun et al.¹⁰² initially employed an electrical voltage of 10 Hz for 0.5 ms, which was subsequently fine-tuned to achieve a 10% reduction in heart rate in rats. This approach significantly constrained infarct size through the mAChRs pathway, mitigating cardiac dysfunction via the nAChRs pathway and extending their survival. While Shao et al.¹⁰³ used a pulse frequency of 4 Hz and an intensity of 6 V to stimulate both right and left vagus nerves, ameliorating the myocardial function in rats by decreasing TNF- α level and arrhythmia score and increasing the expression of $\alpha 7$ nAChR. Xue et al.¹⁰⁴ used electrical voltage from 2 to 4 V to achieve a 10% decrease in the baseline heart rate. This method regulated metabolic homeostasis, modulated mitochondrial function and endoplasmic reticulum stress, and prevented myocardial necrosis and contractile dysfunction during MI in rats. This indicates that the effectiveness of VNS is significantly influenced by factors such as the electrical voltage pulses, stimulation duration, timing, and electrode positioning. These findings underscore the importance of carefully considering these parameters in pre-clinical trials to aim at safeguarding the myocardium against cardiac injury.

Among existing clinical trials, the first pilot study of VNS using CardioFit implantable system showed that VNS may improve quality of life and left ventricle function in chronic HF patients.¹⁰⁵ However, large cohort clinical studies have shown mixed results.¹⁰⁶ For instance, the INOVATE-HF clinical trial evaluated the impact of VNS in patients with HF, involving 390 individuals who received implants. Within 90 days, 46 complications occurred in 37 patients, and, after 16 months, VNS failed to demonstrate efficacy in reducing the mortality rate or HF-related events. Furthermore, it did not induce reverse remodeling or increase the left ventricle ejection fraction in this cohort.¹⁰⁷ The ANTHEM-HF study investigated the impact of continuous cyclic stimulation on both the left and right vagus nerves in HF patients. One patient passed away 3 days after experiencing an embolic stroke during implantation while, in the remaining 59 patients, the devices were successfully implanted. At the six-month mark, the patients showed improvements in left ventricular ejection fraction, left ventricular end-systolic diameter, time-domain of heart rate variability, and high-sensitivity C-reactive protein levels. These findings indicate potential autonomic and anti-inflammatory effects of the treatment.¹⁰⁸ Hence, it is crucial to acknowledge that VNS is an invasive procedure, and uncertainties persist regarding its safety and potential effects on the human heart.

Despite the fact that VNS is Food and Drug Administration approved for pharmaco-resistant depression, epilepsy, and stroke rehabilitation, the device implantation involves perioperative risks. Potential adverse effects include bradyarrhythmias, development of peritracheal hematoma, dyspnea, and respiratory complications.^{109,110} Furthermore, the establishment of an appropriate protocol for VNS in ischemic and HF patients remains unresolved, necessitating optimal techniques, and future studies are needed to discern the impact of long-term VNS on acute and chronic myocardial damage.³ Given the invasive nature of VNS and the perioperative risks associated with device implantation, alternative methods to modulate the cholinergic system must be considered. For instance, the application of transcutaneous VNS (tVNS) could provide the best clinical outcome and overcome surgical VNS limitations.¹¹¹ The tVNS device is placed on either the anterior wall of the outer ear canal (tragus) or the cymba conchae, and it is inexpensive, low risk, and easy to administer. For instance, Choudhary

et al.¹⁰¹ demonstrated that tVNS improves cardiac function and reduces MI area in rats. A 6-months clinical study from Stavarakis et al.¹¹² demonstrated that the tVNS device reduces atrial fibrillation burden and TNF- α level and suppressed inflammation in patients with paroxysmal atrial fibrillation. However, there is a relative paucity of literature surrounding tVNS operation, functionality, and therapeutic effects.¹¹³ Future research is needed to identify the most efficient and safest approaches to enhance ACh levels in the heart and reduce side effects to the patients.

Cholinesterase inhibitors

A non-invasive pharmacological treatment could serve as a potential alternative to chronic VNS therapy, offering a means to target both the neuronal cholinergic system and NNCS. Donepezil, rivastigmine, pyridostigmine, and galantamine are reversible noncompetitive cholinesterase inhibitors. Donepezil is commonly used to prevent progressive neuronal damage,^{114,115} improve cognitive function, and delay the progression of Alzheimer's disease¹¹⁶ and vascular dementia.^{117,118} Clinical trials and meta-analyses suggest that cholinesterase inhibitors, particularly donepezil, may be beneficial in combating CVD due to their anti-inflammatory properties and their ability to increase ACh levels in the heart, as observed in Alzheimer's disease and dementia patients with CVD.^{119–121} Ongnok et al.¹¹⁵ reported that the administration of donepezil significantly mitigates brain pathologies caused by cardiac I/R. This treatment led to increase of BBB junction proteins expression, reduced brain inflammation and oxidative stress, improved mitochondrial function and dynamics, and alleviated amyloid- β accumulation and microglial activation. Furthermore, the cohort study by Nordström et al.¹²² indicated that the use of cholinesterase inhibitors was associated with a 35% reduced risk of MI and death in 7,073 individuals diagnosed with Alzheimer's disease. However, the study was observational, and the patients had no prior history of CVD. Additionally, healthier patients received higher doses of cholinesterase inhibitors, which was more effective than the standard dose.¹²³ Another cohort study found that dementia patients receiving cholinesterase inhibitors have a significantly lower risk of acute coronary syndrome. Consequently, donepezil may protect endothelial cells and improve cardiac vagal activity.¹²⁰

Donepezil has demonstrated favorable cardioprotective effects in animal models against DOX-induced cardiotoxicity,^{16,66} MI,¹²⁴ I/R,¹²⁵ and HF.¹⁷ Additionally, Kakinuma et al. demonstrated that donepezil acts as an amplifier of NNCS activity by enhancing ACh synthesis through upregulating ChAT promoter activity in cardiomyocytes¹²⁶ and cholinergic nerve cells.³⁵ This could indicate that donepezil increases mAChRs and nAChRs activity by preventing ACh degradation and providing cardioprotective effects against CVD. Despite the impact of cholinesterase inhibitors on CVD, the benefit-to-harm ratio remains a crucial issue for clinical trials and adverse effects of cholinesterase inhibitors are significant. An analysis by Kröger et al.¹²⁷ showed that the ADRs are neuropsychiatric (31.4%), gastrointestinal (15.9%), and cardiovascular (11.7%) disorders found in 18,955 reports (out of 43,753 ADRs). Cardiovascular disorders include a significant increase in the risk of bradycardia,¹²⁸ hypotension,^{129,130} cardiac arrhythmia,^{131,132} and syncope.¹³³ While a recent meta-analysis study found no association between donepezil and those cardiovascular disorders,¹³⁴ donepezil still leads to ADRs such as tiredness, panic, sweating, diarrhea, vomiting, muscle tension, speech difficulty, and involuntary tremors.¹³⁵ Hence, further research is needed to clarify the protective effects and ADRs of cholinesterase inhibitors against CVD.

Alternative approaches to target the cholinergic system

Multiple pieces of evidence strongly support the cardioprotective effects of ACh against CVD. Nevertheless, therapeutic approaches to target the cholinergic system remain uncertain and limited, warranting further exploration and investigation. This section proposes alternative approaches for future research, including ACh receptor stimulation, stem cell therapy, gene therapy, miRNA therapy, and nanoparticle drug delivery (Figure 2). Stem cell therapy offers a promising avenue for replenishing cholinergic neurons and fostering the regeneration of new, functional cardiomyocytes in hearts damaged by disease or injury. This approach could enhance self-repair mechanisms and improve the functionality of the damaged heart.¹⁸ While replenishing cholinergic neurons through stem cells has not been extensively studied due to the complexity of the human nervous system, cardiac cell-based therapies, such as bone marrow stem cells, embryonic stem cells (ESCs), and induced pluripotent stem cells (iPSCs), have been explored for their potential to regenerate cardiomyocytes that synthesize ACh.^{136–138} For instance, the transplantation of bone marrow-derived mononuclear cells has been shown to significantly elevate left ventricular ejection fraction in patients with non-ischemic dilated cardiomyopathy and acute MI. However, it has not led to myocardial regeneration.^{136,139} Moreover, in large phase 2 randomized controlled trials, stem cell therapy yielded modest results, which could be due to the poor quality of transplanting stem cells.^{140,141} Additionally, there are concerns with ESCs and iPSCs, such as high survival rates potentially leading to teratoma formation.^{142,143} Despite these challenges, further exploration into stem cell therapy could lead to significant advancements in achieving cardiac regeneration and restoring cardiac function.

Another potential approach could be gene therapy, which aims to deliver the appropriate gene to be expressed and rescue the ischemic heart by increasing ACh synthesis or secretion pathways in cardiomyocytes. However, there are different types of vectors (non-viral and viral vectors) for gene delivery, and the selection of these vectors depends on the pathological condition of CVD patients. Different types of vectors lead to different durations of cardiac expression and transfection efficiency.^{144,145} Cholinergic markers, including ChAT^{42,46} or VACHT,⁴⁵ could be considered new targets to increase ACh and restore the balance between the sympathetic and cholinergic systems in the ischemic heart. As mentioned earlier, ChAT-tg and VACHT-tg models provide cardioprotective effects against myocardial damage and ChAT-tg model also acts as a VNS. Moreover, increasing the overexpression of mAChRs and nAChRs could be another approach. Liu et al.¹⁴⁶ showed that cardiac-specific M3-mAChR-tg mice significantly attenuated the hypertrophic response by reducing the expression of atrial natriuretic peptide and β -myosin heavy chain induced by Ang II. Nevertheless, the selection of the type of vector needs to be thoroughly considered and studied, as the main disadvantage of these vectors is that this technique could induce an inflammatory response.¹⁴⁴ Therefore, additional

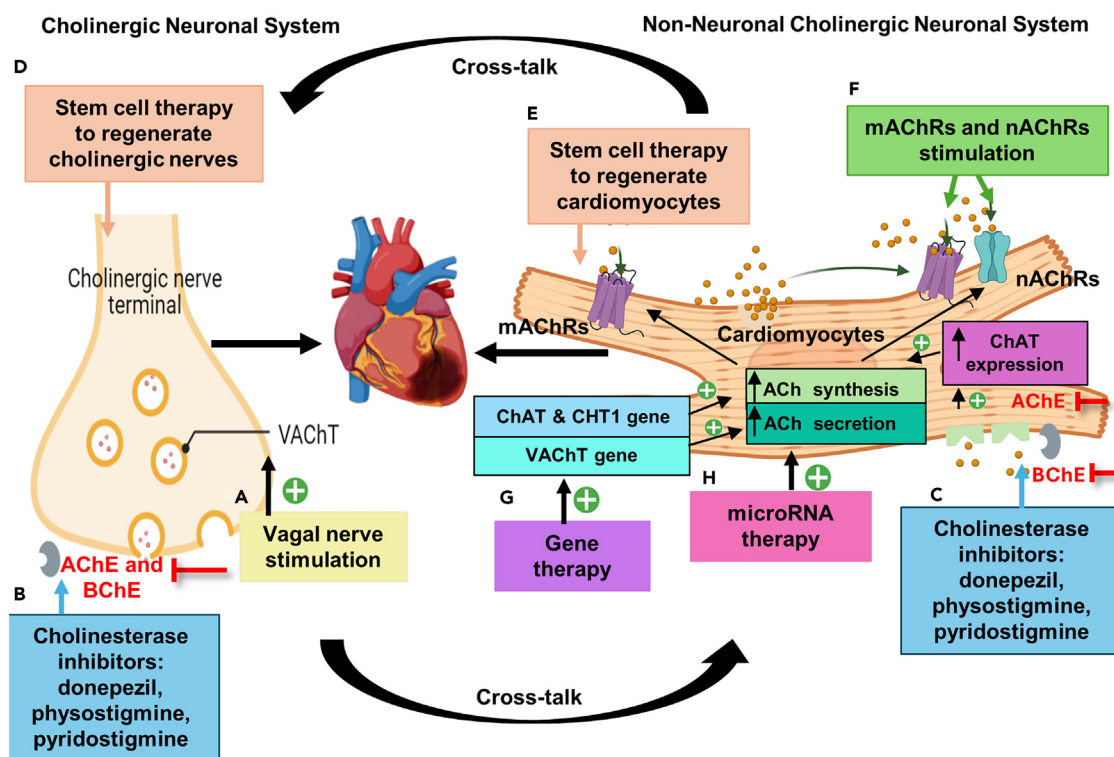


Figure 2. Schematic illustration of therapeutic approaches to target and elevate ACh levels in the infarcted heart

(A) These include vagal/vagus nerve stimulation to produce and secrete ACh.

(B and C) The other approach is through cholinesterase inhibitors to attenuate the degradation of ACh and prolong ACh effect in the extracellular space. Donepezil can simultaneously inhibit cholinesterase enzymes and increase ChAT expression.

(D and E) An alternative approach would be the use of stem cell therapies to either repopulate cholinergic neurons or replace damaged cardiomyocytes respectively.

(F) A fourth approach could include a positive allosteric modulator to stimulate mAChRs and nAChRs activity.

(G) Gene therapy could be a therapeutic strategy to increase the expression of the cholinergic genes such as ChAT, CHT1, and VACHT, which are responsible for ACh synthesis and secretion.

(H) Another possibility is targeting the cardiac miRNAs by diminishing levels of miRNAs that promote ACh degradation. ACh, acetylcholine; AChE, acetylcholinesterase; BChE, butyrylcholinesterase; ChAT, choline acetyltransferase; CHT1, high-affinity choline transporter; mAChRs, muscarinic acetylcholine receptors; nAChRs, nicotinic acetylcholine receptors; VACHT, vesicular acetylcholine transporter.

research is required to find an effective *in vivo* delivery method, the duration of gene expression in the ischemic heart, and any potential toxicity effects in CVD patients.

miRNAs are essential regulators of gene expression. miRNA therapy could potentially target mRNA to inhibit translation or inhibit the degradation of ACh. For example, Oikawa et al.¹⁴⁷ demonstrated that the expression of miR-345 was a regulator of the expression of ChAT mRNA in murine hearts. A synthetic inhibitor for miR-345 could be designed to increase ChAT protein expression and ACh synthesis.¹⁸ While Shaked et al.¹⁴⁸ demonstrated that AChE-targeting miR-132 regulates inflammatory responses, Hanin and Soreq¹⁴⁹ identified 116 and 128 miRNA (miRNA/miRs) that target the 3'-untranslated regions of BChE and AChE (47 for synaptic AChE-S variant and 81 for stress-inducible/readthrough AChE-R variant). However, the cholinesterase-targeting miR sequences show no relevance to most of the cholinesterases' biological functions, suggesting that those sequences have yet to be explored. Hence, further validation is needed to confirm which miRNAs could directly target ChAT, VACHT, and cholinesterases. The effective delivery method and potential toxicity effects of miRNA therapy also remain unclear. More studies are required to find the ideal approach, determine the duration of its effects, and assess its toxicity effects on the heart and other parts of the body. To conclude, targeting the cardiac NNCS through gene or miRNA therapy seems to be the next step.

Furthermore, BChE is abundantly found in the human body,¹⁵⁰ while AChE is present in various conducting tissues such as nerves, muscles, central and peripheral tissues, and motor and sensory fibers.^{22,151} BChE and AChE are present in the body anchored by collagen Q and proline-rich membrane anchors.^{152–154} BChE is also found as a soluble enzyme in the blood, and AChE subunits produce glycosylphosphatidylinositol-anchored dimers and are anchored to red blood cells.^{152,155–157} Under normal conditions, AChE has a higher affinity to hydrolyze ACh than BChE; BChE degrades ACh and carboxylic or phosphoric acid ester-containing compounds and plays important pharmacological and toxicological roles.^{158,159} For instance, increased BChE levels in monkeys¹⁶⁰ and mice¹⁶¹ protected against the toxicity of nerve agents, such as organophosphorus poisoning and cocaine. While AChE inhibition in mice leads to BChE-mediated hydrolysis of

ACh,^{158,162} inhibition of both BChE and AChE could have severe consequences for human health, including lethal intoxication.^{162,163} Recently, Dingová et al.¹⁶⁴ showed that AChE anchored to the membrane of neurons and to the extracellular matrix hydrolyzes neuronal-derived ACh while BChE in the extracellular space hydrolyzes cardiomyocytes-derived ACh in mice hearts. Hence, future work should evaluate the distribution of cholinesterases and their role in the human heart prior to finding an optimal cholinesterase inhibitor to increase the ACh level. Therefore, delivering ACh at small doses and targeting the infarcted area using nanoparticles could be a promising therapeutic approach to increasing ACh levels in the heart. This strategy could potentially enhance the therapeutic effects while minimizing systemic side effects, offering a more precise and effective treatment for conditions requiring cholinergic modulation. Various nanocarriers such as dendrimers, nanometals, nanogels, liposomes, nano-emulsions, polymeric nanoparticles, and nanosuspensions have been studied to deliver drugs at small doses. Nanometal and liposomes nanocarriers have proven to be suitable to deliver ACh, a positively charged and hydrophilic molecule. Studies utilizing silver or gold nanoparticles, as well as chitosan nanoparticles, demonstrated neuroprotective effects against alleviating Alzheimer's disease.^{165–167} However, these approaches are not currently being explored for CVD. Future research should investigate the potential of these nanocarriers for targeted ACh delivery in CVD, which could open new avenues for therapeutic interventions.

CONCLUSIONS AND FUTURE DIRECTIONS

While inhibition of the cholinergic system leads to an increased heart rate, loss of cardiomyocytes,² and increase of production of ROS,¹⁶⁸ there is evidence that the physiological role of both the neuronal cholinergic system and NNCSs protects the heart against ischemic and chronic myocardial damage.^{3,14,19,48} For instance, the cholinergic system prevents cell apoptosis, inhibits ROS production and cardiac hypertrophy,^{61–63} and prevents the progression of HF and cardiac remodeling.^{9,38,64,65} Moreover, the cholinergic system provides beneficial immunomodulatory effects following heart tissue injury, such as stimulating macrophages, inhibiting the release of pro-inflammatory cytokines,^{10,75} attenuating endothelial dysfunction,⁶⁰ and activating cardiac angiogenesis.⁶⁸ However, the relationship between the neuronal cholinergic system and NNCS remains to be fully understood. Understanding the precise mechanisms and interactions between the neuronal cholinergic system and NNCS will also be vital for designing effective treatments.

Additionally, excessive production of ACh and its synthesis can lead to toxicity and bradycardia. Therefore, it is crucial to identify optimal methods for enhancing ACh levels in the heart. VNS and cholinesterase inhibitors are commonly employed to elevate ACh levels in the heart in pre-clinical and clinical studies. However, VNS is invasive, and cholinesterase inhibitors can cause ADRs. Hence, future research should focus on developing targeted and controlled delivery systems to increase ACh in cardiac tissues without causing adverse effects. This includes exploring the use of stem cell therapy, gene therapy, miRNA therapy, and nanoparticles to support and enhance the cholinergic system's protective functions in the heart. In [Figure 2](#), we also mention the potential use of positive allosteric modulators to selectively target specific subtypes of mAChRs and/or nAChRs, potentially increasing receptor affinity for ACh.¹⁶⁹ However, current positive allosteric modulators lack specificity and often interact with multiple subtypes, leading to major ADRs such as increased heart rate and drowsiness due to the varied functions of each subtype and subunit of ACh receptors.^{170,171} Future research should focus on developing more targeted approaches for increasing ACh levels in the heart. This could involve the use of nanoparticle delivery systems or gene therapy to selectively enhance ACh production in cardiac tissue.

More importantly, it is crucial to understand the systemic interactions between the cardiac cholinergic system and other organs, as this will help elucidate the broader physiological impacts of modulating the cholinergic system in the heart. Moreover, long-term studies are necessary to evaluate the safety and efficacy of these alternative approaches and the long-term effects and safety of enhancing ACh levels in the heart, including the chronic impacts on cardiac function and potential off-target effects. Exploring the potential benefits of combining cholinergic modulation with other therapeutic approaches, such as anti-inflammatory treatments or antioxidants, could provide a more comprehensive protective strategy against heart diseases. Addressing these future directions will help harness the full therapeutic potential of the cholinergic system for cardiovascular health and pave the way for innovative treatments for CVD.

Limitations of the study

In this review, we summarized the protective role of the cholinergic system, which produces and releases ACh during myocardial damage. We also highlighted the challenges and limitations of current approaches and suggested alternative approaches to elevate the ACh level in the injured heart. Although our review may not include all relevant studies due to limitations in the literature search, it offers a general overview. Certain findings, particularly ambiguous ones, might not be adequately introduced or discussed. We recommend more in-depth research and discussions to better clarify the roles of ACh in CVDs.

ACKNOWLEDGMENTS

C.G. was supported by a University of Sydney Kick-Start grant, a Cardiothoracic Surgery Research Grant, UTS Seed Funding, Catholic Archdiocese of Sydney Grant for Adult Stem Cell Research, Perpetual IMPACT, Heart Research Australia grant and Heart Research Institute Fellowship. X.W. was supported by a National Heart Foundation Future Leader Fellowship and a Baker Fellowship. C.L.C.M. was supported by a NSW Waratah Scholarship.

AUTHOR CONTRIBUTIONS

C.L.C.M. and C.G. had the idea for the review. C.L.C.M. performed the literature search and drafted the original manuscript. X.W. and C.G. critically revised the work.

DECLARATION OF INTERESTS

The authors declare no competing interests.

REFERENCES

- (2021). Cardiovascular Diseases (CVDs). <https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-cvds>.
- Mahmoud, A.I., O'Meara, C.C., Gemberling, M., Zhao, L., Bryant, D.M., Zheng, R., Gannon, J.B., Cai, L., Choi, W.Y., Egnaczyk, G.F., et al. (2015). Nerves Regulate Cardiomyocyte Proliferation and Heart Regeneration. *Dev. Cell* 34, 387–399.
- Intachai, K., C Chattapakorn, S., Chattapakorn, N., and Shinlapawittayatorn, K. (2018). Revisiting the Cardioprotective Effects of Acetylcholine Receptor Activation against Myocardial Ischemia/Reperfusion Injury. *Int. J. Mol. Sci.* 19, 2466.
- Rocha-Resende, C., da Silva, A.M., Prado, M.A.M., and Guatimosim, S. (2021). Protective and anti-inflammatory effects of acetylcholine in the heart. *Am. J. Physiol. Cell Physiol.* 320, C155–C161.
- Ishise, H., Asanoi, H., Ishizaka, S., Joho, S., Kameyama, T., Umeno, K., and Inoue, H. (1998). Time course of sympathovagal imbalance and left ventricular dysfunction in conscious dogs with heart failure. *J. Appl. Physiol.* 84, 1234–1241.
- Rovere, M.T.L., Bigger, J.T., Jr., Marcus, F.I., Mortara, A., and Schwartz, P.J. (1998). Baroreflex sensitivity and heart-rate variability in prediction of total cardiac mortality after myocardial infarction. *Lancet* 351, 478–484.
- Lechat, P., Hulot, J.S., Escolano, S., Mallet, A., Leizorovicz, A., Werhlen-Grandjean, M., Pochmalicki, G., and Dargie, H. (2001). Heart Rate and Cardiac Rhythm Relationships With Bisoprolol Benefit in Chronic Heart Failure in CIBIS II Trial. *Circulation* 103, 1428–1433.
- Guimarães, D.A., Aquino, N.S.S., Rocha-Resende, C., Jesus, I.C.G., Silva, M.M., Scalzo, S.A., Fonseca, R.C., Durand, M.T., Pereira, V., Tezini, G.C.S.V., et al. (2022). Neuronal cholinergic signaling constrains norepinephrine activity in the heart. *Am. J. Physiol. Cell Physiol.* 322, C794–C801.
- An, X., and Cho, H. (2023). Increased GIRK channel activity prevents arrhythmia in mice with heart failure by enhancing ventricular repolarization. *Sci. Rep.* 13, 22479.
- Vieira-Alves, I., Coimbra-Campos, L.M.C., Sancho, M., da Silva, R.F., Cortes, S.F., and Lemos, V.S. (2020). Role of the $\alpha 7$ Nicotinic Acetylcholine Receptor in the Pathophysiology of Atherosclerosis. *Front. Physiol.* 11, 621769.
- Werner, G.S., Wiegand, V., and Kreuzer, H. (1990). Effect of acetylcholine on arterial and venous grafts and coronary arteries in patients with coronary artery disease. *Eur. Heart J.* 11, 127–137.
- Bezerra, O.C., França, C.M., Rocha, J.A., Neves, G.A., Souza, P.R.M., Teixeira Gomes, M., Malfitano, C., Loleiro, T.C.A., Dourado, P.M., Llesuy, S., et al. (2017). Cholinergic Stimulation Improves Oxidative Stress and Inflammation in Experimental Myocardial Infarction. *Sci. Rep.* 7, 13687.
- Li, X., Zhu, X., Li, B., Xia, B., Tang, H., Hu, J., and Ying, R. (2022). Loss of $\alpha 7$ nAChR enhances endothelial-to-mesenchymal transition after myocardial infarction via NF- κ B activation. *Exp. Cell Res.* 419, 113300.
- Braczko, F., Reinders, J., Lieder, H.R., Heusch, G., and Kleinbongard, P. (2023). Non-neuronal acetylcholine is causal for cardioprotection by hypoxic preconditioning in adult rat cardiomyocytes. *Eur. Heart J.* 44, ehad655-309.
- Calvillo, L., Vanoli, E., Andreoli, E., Besana, A., Omodeo, E., Gneccchi, M., Zerbi, P., Vago, G., Busca, G., and Schwartz, P.J. (2011). Vagal stimulation, through its nicotinic action, limits infarct size and the inflammatory response to myocardial ischemia and reperfusion. *J. Cardiovasc. Pharmacol.* 58, 500–507.
- Prathumsap, N., Ongnok, B., Khuanjing, T., Arinno, A., Maneechote, C., Apaijai, N., Chunchai, T., Arunsak, B., Shinlapawittayatorn, K., Chattapakorn, S.C., and Chattapakorn, N. (2022). Acetylcholine receptor agonists provide cardioprotection in doxorubicin-induced cardiotoxicity via modulating muscarinic M₂ and $\alpha 7$ nicotinic receptor expression. *Transl. Res.* 243, 33–51.
- Handa, T., Katare, R.G., Kakinuma, Y., Arikawa, M., Ando, M., Sasaguri, S., Yamasaki, F., and Sato, T. (2009). Anti-Alzheimer's Drug, Donepezil, Markedly Improves Long-Term Survival After Chronic Heart Failure in Mice. *J. Card. Fail.* 15, 805–811.
- Saw, E.L., Kakinuma, Y., Fronius, M., and Katare, R. (2018). The non-neuronal cholinergic system in the heart: A comprehensive review. *J. Mol. Cell. Cardiol.* 125, 129–139.
- Fujii, T., Mashimo, M., Moriwaki, Y., Misawa, H., Ono, S., Horiguchi, K., and Kawashima, K. (2017). Expression and Function of the Cholinergic System in Immune Cells. *Front. Immunol.* 8, 1085.
- Fujii, T., Mashimo, M., Moriwaki, Y., Misawa, H., Ono, S., Horiguchi, K., and Kawashima, K. (2017). Physiological functions of the cholinergic system in immune cells. *J. Pharmacol. Sci.* 134, 1–21.
- Borovikova, L.V., Ivanova, S., Nardi, D., Zhang, M., Yang, H., Ombrellino, M., and Tracey, K.J. (2000). Role of vagus nerve signaling in CNI-1493-mediated suppression of acute inflammation. *Auton. Neurosci.* 85, 141–147.
- Koelle, G.B. (1954). The histochemical localization of cholinesterases in the central nervous system of the rat. *J. Comp. Neurol.* 100, 211–235.
- Buchholz, B., Donato, M., Perez, V., Ivalde, F.C., Höcht, C., Buitrago, E., Rodríguez, M., and Gelpi, R.J. (2012). Preischemic efferent vagal stimulation increases the size of myocardial infarction in rabbits. Role of the sympathetic nervous system. *Int. J. Cardiol.* 155, 490–491.
- Targosova, K., Kucera, M., Fazekas, T., Kilianova, Z., Stankovicova, T., and Hrabovska, A. (2024). $\alpha 7$ nicotinic receptors play a role in regulation of cardiac hemodynamics. *J. Neurochem.* 168, 414–427.
- Ojiakor, O.A., and Rylett, R.J. (2020). Modulation of sodium-coupled choline transporter CHT function in health and disease. *Neurochem. Int.* 140, 104810.
- Rocha-Resende, C., Roy, A., Resende, R., Ladeira, M.S., Lara, A., de Moraes Gomes, E.R., Prado, V.F., Gros, R., Guatimosim, C., Prado, M.A.M., and Guatimosim, S. (2012). Non-neuronal cholinergic machinery present in cardiomyocytes offsets hypertrophic signals. *J. Mol. Cell. Cardiol.* 53, 206–216.
- Cavey, D., Vincent, J.P., and Lázdunski, M. (1977). The muscarinic receptor of heart cell membranes Association with agonists, antagonists and antiarrhythmic agents. *FEBS Lett.* 84, 110–114.
- Corringer, P.-J., Galzi, J.L., Eiselé, J.L., Bertrand, S., Changeux, J.P., and Bertrand, D. (1995). Identification of a new component of the agonist binding site of the nicotinic $\alpha 7$ homooligomeric receptor. *J. Biol. Chem.* 270, 11749–11752.
- Devillers-Thiéry, A., Galzi, J.L., Eiselé, J.L., Bertrand, S., Bertrand, D., and Changeux, J.P. (1993). Functional architecture of the nicotinic acetylcholine receptor: a prototype of ligand-gated ion channels. *J. Membr. Biol.* 136, 97–112.
- Mesulam, M.M., Guillozet, A., Shaw, P., Levey, A., Duysen, E.G., and Lockridge, O. (2002). Acetylcholinesterase knockouts establish central cholinergic pathways and can use butyrylcholinesterase to hydrolyze acetylcholine. *Neuroscience* 110, 627–639.
- Kuhar, M.J., and Murrin, L.C. (1978). Sodium-dependent, high affinity choline uptake. *J. Neurochem.* 30, 15–21.
- Tarnawski, L., Shavva, V.S., Kort, E.J., Zhuge, Z., Nilsson, I., Gallina, A.L., Martínez-Enguita, D., Heller Sahlgren, B., Weiland, M., Caravaca, A.S., et al. (2023). Cholinergic regulation of vascular endothelial function by human ChAT⁺ T cells. *Proc. Natl. Acad. Sci. USA* 120, e2212476120.
- Eduardo, C.-R.C., Alejandra, T.I.G., Guadalupe, D.R.K.J., Herminia, V.R.G., Lenin, P., Enrique, B.V., Evandro, B.M., Oscar, B., and Iván, G.P.M. (2019). Modulation of the extraneuronal cholinergic system on main innate response leukocytes. *J. Neuroimmunol.* 327, 22–35.
- Kakinuma, Y. (2024). Non-neuronal cholinergic system in the heart influences its homeostasis and an extra-cardiac site, the blood-brain barrier. *Front. Cardiovasc. Med.* 11, 1384637.
- Kakinuma, Y., Akiyama, T., and Sato, T. (2009). Cholinoceptive and cholinergic properties of cardiomyocytes involving an amplification mechanism for vagal efferent effects in sparsely innervated ventricular myocardium. *FEBS J.* 276, 5111–5125.
- Rana, O.R., Schauerte, P., Kluttig, R., Schröder, J.W., Koenen, R.R., Weber, C., Nolte, K.W., Weis, J., Hoffmann, R., Marx, N., and Saygili, E. (2010). Acetylcholine as an age-dependent non-neuronal source in the heart. *Auton. Neurosci.* 156, 82–89.
- Roy, A., Fields, W.C., Rocha-Resende, C., Resende, R.R., Guatimosim, S., Prado, V.F., Gros, R., and Prado, M.A.M. (2013).

- Cardiomyocyte-secreted acetylcholine is required for maintenance of homeostasis in the heart. *FASEB J.* 27, 5072–5082.
38. Gavioli, M., Lara, A., Almeida, P.W.M., Lima, A.M., Damasceno, D.D., Rocha-Resende, C., Ladeira, M., Resende, R.R., Martinelli, P.M., Melo, M.B., et al. (2014). Cholinergic signaling exerts protective effects in models of sympathetic hyperactivity-induced cardiac dysfunction. *PLoS One* 9, e100179.
 39. Lara, A., Damasceno, D.D., Pires, R., Gros, R., Gomes, E.R., Gavioli, M., Lima, R.F., Guimarães, D., Lima, P., Bueno, C.R., Jr., et al. (2010). Dysautonomia due to reduced cholinergic neurotransmission causes cardiac remodeling and heart failure. *Mol. Cell Biol.* 30, 1746–1756.
 40. Roy, A., Dakroub, M., Tezini, G.C.S.V., Liu, Y., Guatimosim, S., Feng, Q., Salgado, H.C., Prado, V.F., Prado, M.A.M., and Gros, R. (2016). Cardiac acetylcholine inhibits ventricular remodeling and dysfunction under pathologic conditions. *FASEB J.* 30, 688–701.
 41. Kakinuma, Y., Akiyama, T., Okazaki, K., Arikawa, M., Noguchi, T., and Sato, T. (2012). A non-neuronal cardiac cholinergic system plays a protective role in myocardium salvage during ischemic insults. *PLoS One* 7, e50761.
 42. Kakinuma, Y., Tsuda, M., Okazaki, K., Akiyama, T., Arikawa, M., Noguchi, T., and Sato, T. (2013). Heart-specific overexpression of choline acetyltransferase gene protects murine heart against ischemia through hypoxia-inducible factor-1 α -related defense mechanisms. *J. Am. Heart Assoc.* 2, e004887.
 43. Reardon, C., Duncan, G.S., Brüstle, A., Brenner, D., Tusche, M.W., Olofsson, P.S., Rosas-Ballina, M., Tracey, K.J., and Mak, T.W. (2013). Lymphocyte-derived ACh regulates local innate but not adaptive immunity. *Proc. Natl. Acad. Sci. USA* 110, 1410–1415.
 44. Oikawa, S., Kai, Y., Tsuda, M., Ohata, H., Mano, A., Mizoguchi, N., Sugama, S., Nemoto, T., Suzuki, K., Kurabayashi, A., et al. (2016). Non-neuronal cardiac cholinergic system influences CNS via the vagus nerve to acquire a stress-refractory propensity. *Clin. Sci.* 130, 1913–1928.
 45. Rocha-Resende, C., Weinheimer, C., Bajpai, G., Adamo, L., Matkovich, S.J., Schilling, J., Barger, P.M., Lavine, K.J., and Mann, D.L. (2019). Immunomodulatory role of nonneuronal cholinergic signaling in myocardial injury. *JCI insight* 5, e128961.
 46. Saw, E.L., Pearson, J.T., Schwenke, D.O., Munasinghe, P.E., Tsuchimochi, H., Rawal, S., Coffey, S., Davis, P., Bunton, R., Van Hout, I., et al. (2021). Activation of the cardiac non-neuronal cholinergic system prevents the development of diabetes-associated cardiovascular complications. *Cardiovasc. Diabetol.* 20, 50.
 47. Munasinghe, P.E., Saw, E.L., Reily-Bell, M., Tonkin, D., Kakinuma, Y., Fronius, M., and Katare, R. (2023). Non-neuronal cholinergic system delays cardiac remodeling in type 1 diabetes. *Heliyon* 9, e17434.
 48. Braczko, F., Reinders, J., Lieder, H., Heusch, G., and Kleinbongard, P. (2021). The Non-Neuronal Cholinergic System is Causal for Cardioprotection by Hypoxic Preconditioning in Isolated Adult Rat Cardiomyocytes. *FASEB J.* 327, H70–H79.
 49. Oikawa, S., Iketani, M., and Kakinuma, Y. (2014). A non-neuronal cholinergic system regulates cellular ATP levels to maintain cell viability. *Cell. Physiol. Biochem.* 34, 781–789.
 50. Oikawa, S., Kai, Y., Mano, A., Sugama, S., Mizoguchi, N., Tsuda, M., Muramoto, K., and Kakinuma, Y. (2019). Potentiating a non-neuronal cardiac cholinergic system reinforces the functional integrity of the blood brain barrier associated with systemic anti-inflammatory responses. *Brain Behav. Immun.* 81, 122–137.
 51. McAreavey, D., Neilson, J.M., Ewing, D.J., and Russell, D.C. (1989). Cardiac parasympathetic activity during the early hours of acute myocardial infarction. *Br. Heart J.* 62, 165–170.
 52. Khuanjing, T., Palee, S., Chattipakorn, S.C., and Chattipakorn, N. (2020). The effects of acetylcholinesterase inhibitors on the heart in acute myocardial infarction and heart failure: From cells to patient reports. *Acta Physiol.* 228, e13396.
 53. LaCroix, C., Freeling, J., Giles, A., Wess, J., and Li, Y.F. (2008). Deficiency of M2 muscarinic acetylcholine receptors increases susceptibility of ventricular function to chronic adrenergic stress. *Am. J. Physiol. Heart Circ. Physiol.* 294, H810–H820.
 54. Tettelbaum, H.A., Newton, J.E.O., and Gantt, W.H. (1971). Cardiovascular responses to acetylcholine: Effects of pentobarbital and autonomic blocking agents. *Cond. Reflex Pavlovian J. Res. Therapy* 6, 101–118.
 55. Hare, J.M., Keaney, J.F., Jr., Balligand, J.L., Loscalzo, J., Smith, T.W., and Colucci, W.S. (1995). Role of nitric oxide in parasympathetic modulation of beta-adrenergic myocardial contractility in normal dogs. *J. Clin. Invest.* 95, 360–366.
 56. Henning, R.J., Khalil, I.R., and Levy, M.N. (1990). Vagal stimulation attenuates sympathetic enhancement of left ventricular function. *Am. J. Physiol.* 258, H1470–H1475.
 57. Ren, C., Tong, Y.L., Li, J.C., Lu, Z.Q., and Yao, Y.M. (2017). The Protective Effect of Alpha 7 Nicotinic Acetylcholine Receptor Activation on Critical Illness and Its Mechanism. *Int. J. Biol. Sci.* 13, 46–56.
 58. Petrov, K.A., Proskurina, S.E., and Krejci, E. (2021). Cholinesterases in Tripartite Neuromuscular Synapse. *Front. Mol. Neurosci.* 14, 811220.
 59. Osipov, A.V., Averin, A.S., Shaykhtudinova, E.R., Dyachenko, I.A., Tsetlin, V.I., and Utkin, Y.N. (2023). Muscarinic and nicotinic acetylcholine receptors in the regulation of the cardiovascular system. *Russ. J. Bioorg. Chem.* 49, 3–22.
 60. Capilupi, M.J., Kerath, S.M., and Becker, L.B. (2020). Vagus Nerve Stimulation and the Cardiovascular System. *Cold Spring Harbor Perspect. Med.* 10, a034173.
 61. Fei, Y.-D., Chen, M., Guo, S., Ueoka, A., Chen, Z., Rubart-von der Lohe, M., Everett, T.H., 4th, Qu, Z., Weiss, J.N., and Chen, P.S. (2021). Simultaneous activation of the small conductance calcium-activated potassium current by acetylcholine and inhibition of sodium current by ajmaline cause J-wave syndrome in Langendorff-perfused rabbit ventricles. *Heart Rhythm* 18, 98–108.
 62. Kakinuma, Y., Ando, M., Kuwabara, M., Katare, R.G., Okudela, K., Kobayashi, M., and Sato, T. (2005). Acetylcholine from vagal stimulation protects cardiomyocytes against ischemia and hypoxia involving additive non-hypoxic induction of HIF-1 α . *FEBS Lett.* 579, 2111–2118.
 63. Soeki, T., Niki, T., Uematsu, E., Bando, S., Matsuura, T., Kusunose, K., Ise, T., Ueda, Y., Tomita, N., Yamaguchi, K., et al. (2013). Ghrelin protects the heart against ischemia-induced arrhythmias by preserving connexin-43 protein. *Heart Vess.* 28, 795–801.
 64. Tsutsumi, T., Ide, T., Yamato, M., Kudou, W., Andou, M., Hirooka, Y., Utsumi, H., Tsutsui, H., and Sunagawa, K. (2008). Modulation of the myocardial redox state by vagal nerve stimulation after experimental myocardial infarction. *Cardiovasc. Res.* 77, 713–721.
 65. Nuntaphum, W., Pongkan, W., Wongjaikam, S., Thummasorn, S., Tanajak, P., Khamsaekaw, J., Intachai, K., Chattipakorn, S.C., Chattipakorn, N., and Shinlapawittayatorn, K. (2018). Vagus nerve stimulation exerts cardioprotection against myocardial ischemia/reperfusion injury predominantly through its efferent vagal fibers. *Basic Res. Cardiol.* 113, 22.
 66. Khuanjing, T., Ongnok, B., Maneechote, C., Siri-Angkul, N., Prathumsap, N., Arinno, A., Chunchai, T., Arunsak, B., Chattipakorn, S.C., and Chattipakorn, N. (2021). Acetylcholinesterase inhibitor ameliorates doxorubicin-induced cardiotoxicity through reducing RIP1-mediated necroptosis. *Pharmacol. Res.* 173, 105882.
 67. Liu, J.-J., Li, D.L., Zhou, J., Sun, L., Zhao, M., Kong, S.S., Wang, Y.H., Yu, X.J., Zhou, J., and Zang, W.J. (2011). Acetylcholine prevents angiotensin II-induced oxidative stress and apoptosis in H9c2 cells. *Apoptosis* 16, 94–103.
 68. Xu, M., Xue, R.Q., Lu, Y., Yong, S.Y., Wu, Q., Cui, Y.L., Zuo, X.T., Yu, X.J., Zhao, M., and Zang, W.J. (2019). Choline ameliorates cardiac hypertrophy by regulating metabolic remodelling and UPRmt through SIRT3-AMPK pathway. *Cardiovasc. Res.* 115, 530–545.
 69. Xue, R.-Q., Zhao, M., Wu, Q., Yang, S., Cui, Y.L., Yu, X.J., Liu, J., and Zang, W.J. (2019). Regulation of mitochondrial cristae remodelling by acetylcholine alleviates palmitate-induced cardiomyocyte hypertrophy. *Free Radic. Biol. Med.* 145, 103–117.
 70. Katare, R.G., Ando, M., Kakinuma, Y., Arikawa, M., Handa, T., Yamasaki, F., and Sato, T. (2009). Vagal nerve stimulation prevents reperfusion injury through inhibition of opening of mitochondrial permeability transition pore independent of the bradycardiac effect. *J. Thorac. Cardiovasc. Surg.* 137, 223–231.
 71. Sun, J., Lu, Y., Huang, Y., and Wugeti, N. (2015). Unilateral vagus nerve stimulation improves ventricular autonomic nerve distribution and functional imbalance in a canine heart failure model. *Int. J. Clin. Exp. Med.* 8, 9334–9340.
 72. Wang, Z., Zhao, G., Zibrila, A.I., Li, Y., Liu, J., and Feng, W. (2021). Acetylcholine ameliorated hypoxia-induced oxidative stress and apoptosis in trophoblast cells via p38 MAPK/NF- κ B pathway. *Mol. Hum. Reprod.* 27, gaab045.
 73. Intachai, K., Chattipakorn, S.C., Chattipakorn, N., and Shinlapawittayatorn, K. (2022). Acetylcholine exerts cytoprotection against hypoxia/reoxygenation-induced apoptosis, autophagy and mitochondrial impairment through both muscarinic and nicotinic receptors. *Apoptosis* 27, 233–245.
 74. Shinlapawittayatorn, K., Chinda, K., Palee, S., Surinkaew, S., Kumfu, S., Kumphune, S.,

- Chattipakorn, S., KenKnight, B.H., and Chattipakorn, N. (2014). Vagus nerve stimulation initiated late during ischemia, but not reperfusion, exerts cardioprotection via amelioration of cardiac mitochondrial dysfunction. *Heart Rhythm* 11, 2278–2287.
75. Zhang, J., Yong, Y., Li, X., Hu, Y., Wang, J., Wang, Y.Q., Song, W., Chen, W.T., Xie, J., Chen, X.M., et al. (2015). Vagal modulation of high mobility group box-1 protein mediates electroacupuncture-induced cardioprotection in ischemia-reperfusion injury. *Sci. Rep.* 5, 15503.
76. Li, Z., Li, X., Zhu, Y., Chen, Q., Li, B., and Zhang, F. (2020). Protective effects of acetylcholine on hypoxia-induced endothelial-to-mesenchymal transition in human cardiac microvascular endothelial cells. *Mol. Cell. Biochem.* 473, 101–110.
77. Zhao, L., Chen, T., Hang, P., Li, W., Guo, J., Pan, Y., Du, J., Zheng, Y., and Du, Z. (2019). Choline Attenuates Cardiac Fibrosis by Inhibiting p38MAPK Signaling Possibly by Acting on M3 Muscarinic Acetylcholine Receptor. *Front. Pharmacol.* 10, 1386.
78. Lampert, R., Bremner, J.D., Su, S., Miller, A., Lee, F., Cheema, F., Goldberg, J., and Vaccarino, V. (2008). Decreased heart rate variability is associated with higher levels of inflammation in middle-aged men. *Am. Heart J.* 156, 759.e1–759.e7.
79. Borovikova, L.V., Ivanova, S., Zhang, M., Yang, H., Botchkina, G.I., Watkins, L.R., Wang, H., Abumrad, N., Eaton, J.W., and Tracey, K.J. (2000). Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxin. *Nature* 405, 458–462.
80. Parrish, W.R., Rosas-Ballina, M., Gallowitsch-Puerta, M., Ochani, M., Ochani, K., Yang, L.H., Hudson, L., Lin, X., Patel, N., Johnson, S.M., et al. (2008). Modulation of TNF release by choline requires alpha7 subunit nicotinic acetylcholine receptor-mediated signaling. *Mol. Med.* 14, 567–574.
81. Li, D.-J., Fu, H., Tong, J., Li, Y.H., Qu, L.F., Wang, P., and Shen, F.M. (2018). Cholinergic anti-inflammatory pathway inhibits neointimal hyperplasia by suppressing inflammation and oxidative stress. *Redox Biol.* 15, 22–33.
82. Huston, J.M., and Tracey, K.J. (2011). The pulse of inflammation: heart rate variability, the cholinergic anti-inflammatory pathway and implications for therapy. *J. Intern. Med.* 269, 45–53.
83. Zeisberg, E.M., Tarnavski, O., Zeisberg, M., Dorfman, A.L., McMullen, J.R., Gustafsson, E., Chandraker, A., Yuan, X., Pu, W.T., Roberts, A.B., et al. (2007). Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. *Nat. Med.* 13, 952–961.
84. Hao, C., Huang, Z.H., Song, S.W., Shi, Y.Q., Cheng, X.W., Murohara, T., Lu, W., Su, D.F., and Duan, J.L. (2014). Arterial Baroreflex Dysfunction Impairs Ischemia-Induced Angiogenesis. *J. Am. Heart Assoc.* 3, e000804.
85. Shepherd, J.T., and Vanhoutte, P.M. (1985). Spasm of the Coronary Arteries: Causes and Consequences (the Scientist's Viewpoint). *Mayo Clin. Proc.* 60, 33–46.
86. Suzuki, S., Kaikita, K., Yamamoto, E., Jinnouchi, H., and Tsujita, K. (2021). Role of acetylcholine spasm provocation test as a pathophysiological assessment in nonobstructive coronary artery disease. *Cardiovasc. Interv. Ther.* 36, 39–51.
87. Ishii, M., Kaikita, K., Sato, K., Tanaka, T., Sugamura, K., Sakamoto, K., Izumiya, Y., Yamamoto, E., Tsujita, K., Yamamoto, M., et al. (2015). Acetylcholine-Provoked Coronary Spasm at Site of Significant Organic Stenosis Predicts Poor Prognosis in Patients With Coronary Vasospastic Angina. *J. Am. Coll. Cardiol.* 66, 1105–1115.
88. Isogai, T., Yasunaga, H., Matsui, H., Tanaka, H., Ueda, T., Horiguchi, H., and Fushimi, K. (2015). Serious cardiac complications in coronary spasm provocation tests using acetylcholine or ergonovine: analysis of 21 512 patients for the diagnosis procedure combination database in Japan. *Clin. Cardiol.* 38, 171–177.
89. Erickson, S.R., and Yousef, M.J. (1991). Hypotension and bradycardia possibly associated with intraocular injection of acetylcholine. *DICP* 25, 1178–1180.
90. Hadaya, J., and Ardell, J.L. (2020). Autonomic modulation for cardiovascular disease. *Front. Physiol.* 11, 617459.
91. Buchholz, B., Donato, M., Perez, V., Deutsch, A.C.R., Höcht, C., Del Mauro, J.S., Rodríguez, M., and Gelpi, R.J. (2015). Changes in the loading conditions induced by vagal stimulation modify the myocardial infarct size through sympathetic-parasympathetic interactions. *Pflugers Arch.* 467, 1509–1522.
92. Cavalcante, G.L., Brognara, F., Oliveira, L.V.d.C., Lатарo, R.M., Durand, M.d.T., de Oliveira, A.P., da Nóbrega, A.C.L., Salgado, H.C., and Sabino, J.P.J. (2021). Benefits of pharmacological and electrical cholinergic stimulation in hypertension and heart failure. *Acta Physiol.* 232, e13663.
93. Guo, F., Wang, Y., Wang, J., Liu, Z., Lai, Y., Zhou, Z., Liu, Z., Zhou, Y., Xu, X., Li, Z., et al. (2022). Choline Protects the Heart from Doxorubicin-Induced Cardiotoxicity through Vagal Activation and Nrf2/HO-1 Pathway. *Oxid. Med. Cell. Longev.* 2022, 4740931.
94. Siripakkaphant, C., Ongnok, B., Prathumsap, N., Khuangjing, T., Chunchai, T., Arunsak, B., Pantiya, P., Chattipakorn, N., and Chattipakorn, S.C. (2023). Vagus Nerve Stimulation Provides Neuroprotection Against Doxorubicin-induced Chemobrain Via Activations of Both Muscarinic and Nicotinic Acetylcholine Receptors. *Alzheimer's Dementia* 19, e073548.
95. Uitterdijk, A., Yetgin, T., te Lintel Hekkert, M., Sneep, S., Krabbendam-Peters, I., van Beusekom, H.M.M., Fischer, T.M., Cornelussen, R.N., Manintveld, O.C., Merkus, D., and Duncker, D.J. (2015). Vagal nerve stimulation started just prior to reperfusion limits infarct size and no-reflow. *Basic Res. Cardiol.* 110, 508.
96. Galasso, G., Schiekofer, S., D'Anna, C., Gioia, G.D., Piccolo, R., Niglio, T., Rosa, R.D., Striscigliolo, T., Cirillo, P., Piscione, F., and Trimarco, B. (2014). No-reflow phenomenon: pathophysiology, diagnosis, prevention, and treatment. A review of the current literature and future perspectives. *Angiology* 65, 180–189.
97. Schwartz, B.G., and Kloner, R.A. (2012). Coronary no reflow. *J. Mol. Cell. Cardiol.* 52, 873–882.
98. Zhao, M., He, X., Bi, X.Y., Yu, X.J., Gil Wier, W., and Zang, W.J. (2013). Vagal stimulation triggers peripheral vascular protection through the cholinergic anti-inflammatory pathway in a rat model of myocardial ischemia/reperfusion. *Basic Res. Cardiol.* 108, 345.
99. Li, M., Zheng, C., Sato, T., Kawada, T., Sugimachi, M., and Sunagawa, K. (2004). Vagal nerve stimulation markedly improves long-term survival after chronic heart failure in rats. *Circulation* 109, 120–124.
100. Liu Chung Ming, C., Sesperez, K., Ben-Sefer, E., Arpon, D., McGrath, K., McClements, L., and Gentile, C. (2021). Considerations to Model Heart Disease in Women with Preeclampsia and Cardiovascular Disease. *Cells* 10, 899.
101. Choudhary, R.C., Ahmed, U., Shoaib, M., Alper, E., Rehman, A., Kim, J., Shinozaki, K., Volpe, B.T., Chavan, S., Zanos, S., et al. (2022). Threshold adjusted vagus nerve stimulation after asphyxial cardiac arrest results in neuroprotection and improved survival. *Bioelectron. Med.* 8, 10.
102. Sun, P., Wang, J., Zhao, S., Yang, Z., Tang, Z., Ravindra, N., Bradley, J., Ornato, J.P., Peberdy, M.A., and Tang, W. (2018). Improved Outcomes of Cardiopulmonary Resuscitation in Rats Treated With Vagus Nerve Stimulation and Its Potential Mechanism. *Shock* 49, 698–703.
103. Shao, W.J., Shu, T.T., Xu, S., Liang, L.C., Grange, J.M.L., Zhou, Y.R., Huang, H., Cai, Y., Zhang, Q., and Sun, P. (2021). Left-sided vagus nerve stimulation improves cardiopulmonary resuscitation outcomes in rats as effectively as right-sided vagus nerve stimulation. *World J. Emerg. Med.* 12, 309–316.
104. Xue, R.-Q., Sun, L., Yu, X.J., Li, D.L., and Zang, W.J. (2017). Vagal nerve stimulation improves mitochondrial dynamics via an M₃ receptor/CaMKKβ/AMPK pathway in isoproterenol-induced myocardial ischaemia. *J. Cell Mol. Med.* 21, 58–71.
105. De Ferrari, G.M., Crijns, H.J.G.M., Borggrefe, M., Milasinovic, G., Smid, J., Zabel, M., Gavazzi, A., Sanzo, A., Dennert, R., Kuschyk, J., et al. (2011). Chronic vagus nerve stimulation: a new and promising therapeutic approach for chronic heart failure. *Eur. Heart J.* 32, 847–855.
106. Zafeiropoulos, S., Ahmed, U., Bikou, A., Mughrabi, I.T., Stavakis, S., and Zanos, S. (2024). Vagus nerve stimulation for cardiovascular diseases: Is there light at the end of the tunnel? *Trends Cardiovasc. Med.* 34, 327–337.
107. Gold, M.R., Van Veldhuisen, D.J., Hauptman, P.J., Borggrefe, M., Kubo, S.H., Lieberman, R.A., Milasinovic, G., Berman, B.J., Djordjevic, S., Neelagaru, S., et al. (2016). Vagus Nerve Stimulation for the Treatment of Heart Failure. *J. Am. Coll. Cardiol.* 68, 149–158.
108. Premchand, R.K., Sharma, K., Mittal, S., Monteiro, R., Dixit, S., Libbus, I., DiCarlo, L.A., Ardell, J.L., Rector, T.S., Amurthur, B., et al. (2014). Autonomic Regulation Therapy via Left or Right Cervical Vagus Nerve Stimulation in Patients With Chronic Heart Failure: Results of the ANTHEM-HF Trial. *J. Card. Fail.* 20, 808–816.
109. Dawson, J., Liu, C.Y., Francisco, G.E., Cramer, S.C., Wolf, S.L., Dixit, A., Alexander, J., Ali, R., Brown, B.L., Feng, W., et al. (2021). Vagus nerve stimulation paired with rehabilitation for upper limb motor function after ischaemic stroke (VNS-REHAB): a randomised, blinded, pivotal, device trial. *Lancet* 397, 1545–1553.
110. Yap, J.Y.Y., Keatch, C., Lambert, E., Woods, W., Stoddart, P.R., and Kameneva, T. (2020).

- Critical Review of Transcutaneous Vagus Nerve Stimulation: Challenges for Translation to Clinical Practice. *Front. Neurosci.* 14, 284.
111. Elamin, A.B.A., Forsat, K., Senok, S.S., and Goswami, N. (2023). Vagus Nerve Stimulation and Its Cardioprotective Abilities: A Systematic Review. *J. Clin. Med.* 12, 1717.
 112. Stavrakis, S., Stoner, J.A., Humphrey, M.B., Morris, L., Filiberti, A., Reynolds, J.C., Elkholey, K., Javed, I., Twidale, N., Riha, P., et al. (2020). TREAT AF (Transcutaneous Electrical Vagus Nerve Stimulation to Suppress Atrial Fibrillation). *JACC. Clin. Electrophysiol.* 6, 282–291.
 113. Butt, M.F., Albusoda, A., Farmer, A.D., and Aziz, Q. (2020). The anatomical basis for transcutaneous auricular vagus nerve stimulation. *J. Anat.* 236, 588–611.
 114. Wiendl, H., Elger, C., Förstl, H., Hartung, H.P., Oertel, W., Reichmann, H., and Schwab, S. (2015). Gaps Between Aims and Achievements in Therapeutic Modification of Neuronal Damage ("Neuroprotection"). *Neurotherapeutics* 12, 449–454.
 115. Ongnok, B., Khuanjing, T., Chunchai, T., Kerdphoo, S., Jaiwongkam, T., Chattipakorn, N., and Chattipakorn, S.C. (2021). Donepezil provides neuroprotective effects against brain injury and Alzheimer's pathology under conditions of cardiac ischemia/reperfusion injury. *Biochim. Biophys. Acta, Mol. Basis Dis.* 1867, 165975.
 116. Birks, J.S., and Harvey, R.J. (2018). Donepezil for dementia due to Alzheimer's disease. *Cochrane Database Syst. Rev.* 6, Cd001190.
 117. Battle, C.E., Abdul-Rahim, A.H., Shenkin, S.D., Hewitt, J., and Quinn, T.J. (2021). Cholinesterase inhibitors for vascular dementia and other vascular cognitive impairments: a network meta-analysis. *Cochrane Database Syst. Rev.* 2, CD013306.
 118. Jian, W.-X., Zhang, Z., Zhan, J.H., Chu, S.F., Peng, Y., Zhao, M., Wang, Q., and Chen, N.H. (2020). Donepezil attenuates vascular dementia in rats through increasing BDNF induced by reducing HDAC6 nuclear translocation. *Acta Pharmacol. Sin.* 41, 588–598.
 119. Kazmierski, J., Messini-Zachou, C., Gkioka, M., and Tsolaki, M. (2018). The Impact of a Long-Term Rivastigmine and Donepezil Treatment on All-Cause Mortality in Patients With Alzheimer's Disease. *Am. J. Alzheimers Dis. Other Dement.* 33, 385–393.
 120. Wu, P.-H., Lin, Y.T., Hsu, P.C., Yang, Y.H., Lin, T.H., and Huang, C.T. (2015). Impact of acetylcholinesterase inhibitors on the occurrence of acute coronary syndrome in patients with dementia. *Sci. Rep.* 5, 15451.
 121. Isik, A.T., Soysal, P., Stubbs, B., Solmi, M., Basso, C., Maggi, S., Schofield, P., Veronese, N., and Mueller, C. (2018). Cardiovascular Outcomes of Cholinesterase Inhibitors in Individuals with Dementia: A Meta-Analysis and Systematic Review. *J. Am. Geriatr. Soc.* 66, 1805–1811.
 122. Nordström, P., Religa, D., Wimo, A., Winblad, B., and Eriksdotter, M. (2013). The use of cholinesterase inhibitors and the risk of myocardial infarction and death: a nationwide cohort study in subjects with Alzheimer's disease. *Eur. Heart J.* 34, 2585–2591.
 123. Wang, H., Zong, Y., Han, Y., Zhao, J., Liu, H., and Liu, Y. (2022). Compared of efficacy and safety of high-dose donepezil vs standard-dose donepezil among elderly patients with Alzheimer's disease: a systematic review and meta-analysis. *Expert Opin. Drug Saf.* 21, 407–415.
 124. Li, M., Zheng, C., Kawada, T., Uemura, K., Yokota, S., Matsushita, H., and Saku, K. (2024). Donepezil attenuates progression of cardiovascular remodeling and improves prognosis in spontaneously hypertensive rats with chronic myocardial infarction. *Hypertens. Res.* 47, 1298–1308.
 125. Khuanjing, T., Palee, S., Kerdphoo, S., Jaiwongkam, T., Anomasiri, A., Chattipakorn, S.C., and Chattipakorn, N. (2021). Donepezil attenuated cardiac ischemia/reperfusion injury through balancing mitochondrial dynamics, mitophagy, and autophagy. *Transl. Res.* 230, 82–97.
 126. Kakinuma, Y., Furihata, M., Akiyama, T., Arikawa, M., Handa, T., Katare, R.G., and Sato, T. (2010). Donepezil, an acetylcholinesterase inhibitor against Alzheimer's dementia, promotes angiogenesis in an ischemic hindlimb model. *J. Mol. Cell. Cardiol.* 48, 680–693.
 127. Kröger, E., Mouls, M., Wilchesky, M., Berkers, M., Carmichael, P.H., van Marum, R., Souverein, P., Egberts, T., and Laroche, M.L. (2015). Adverse Drug Reactions Reported With Cholinesterase Inhibitors: An Analysis of 16 Years of Individual Case Safety Reports From VigiBase. *Ann. Pharmacother.* 49, 1197–1206.
 128. Morris, R., Luboff, H., Jose, R.P., Eckhoff, K., Bu, K., Pham, M., Rohlsen-Neal, D., and Cheng, F. (2021). Bradycardia Due to Donepezil in Adults: Systematic Analysis of FDA Adverse Event Reporting System. *J. Alzheimers Dis.* 81, 297–307.
 129. Bordier, P., Garrigue, S., Lanusse, S., Margaine, J., Robert, F., Gencel, L., and Lafitte, A. (2006). Cardiovascular effects and risk of syncope related to donepezil in patients with Alzheimer's disease. *CNS Drugs* 20, 411–417.
 130. Pu, Z., Xu, W., Lin, Y., Shen, J., and Sun, Y. (2019). Donepezil decreases heart rate in elderly patients with Alzheimer's disease. *Int. J. Clin. Pharmacol. Ther.* 57, 94–100.
 131. Suleyman, T., Tefvik, P., Abdulkadir, G., and Ozlem, S. (2006). Complete atrioventricular block and ventricular tachyarrhythmia associated with donepezil. *Emerg. Med. J.* 23, 641–642.
 132. Hadano, Y., Ogawa, H., Wakeyama, T., Iwami, T., Kimura, M., Mochizuki, M., Akashi, S., Miyazaki, Y., Nakashima, T., and Shimizu, A. (2013). Donepezil-induced torsades de pointes without QT prolongation. *J. Cardiol. Cases* 8, e69–e71.
 133. Malik, B.H., Hamid, P., Khan, S., Gupta, D., and Islam, M. (2019). Correlation between donepezil and QTc prolongation and torsades de pointes: a very rare phenomenon. *Cureus* 11, e6451.
 134. Nham, T., Garcia, M.C., Tsang, K.J., Silva, J.M., Schneider, T., Deng, J., Lohit, S., Mbuagbaw, L., and Holbrook, A. (2024). Proarrhythmic major adverse cardiac events with donepezil: A systematic review with meta-analysis. *J. Am. Geriatrics Soc* 71, 1.
 135. Li, H.-C., Luo, K.X., Wang, J.S., and Wang, Q.X. (2020). Extrapyramidal side effect of donepezil hydrochloride in an elderly patient: a case report. *Medicine* 99, e19443.
 136. Wen, Y., Ding, J., Zhang, B., and Gao, Q. (2018). Bone marrow-derived mononuclear cell therapy for nonischemic dilated cardiomyopathy—A meta-analysis. *Eur. J. Clin. Invest.* 48, e12894.
 137. Nasser, B.A., Ebell, W., Dandel, M., Kukucka, M., Gebker, R., Doltra, A., Knosalla, C., Choi, Y.H., Hetzer, R., and Stamm, C. (2014). Autologous CD133+ bone marrow cells and bypass grafting for regeneration of ischaemic myocardium: the Cardio133 trial. *Eur. Heart J.* 35, 1263–1274.
 138. Liu Chung Ming, C., Ben-Sefer, E., and Gentile, C. (2022). Stem Cell-Based 3D Bioprinting for Cardiovascular Tissue Regeneration. In *Advanced Technologies in Cardiovascular Bioengineering* (Springer), pp. 281–312.
 139. Delewi, R., Andriessen, A., Tijssen, J.G., Zijlstra, F., Piek, J.J., and Hirsch, A. (2012). Impact of intracoronary cell therapy on left ventricular function in the setting of acute myocardial infarction: a meta-analysis of randomised controlled clinical trials. *Heart* 99, 225.
 140. Nollet, E., Hoymans, V.Y., Van Craenenbroeck, A.H., Vrints, C.J., and Van Craenenbroeck, E.M. (2016). Improving stem cell therapy in cardiovascular diseases: the potential role of microRNA. *Am. J. Physiol. Heart Circ. Physiol.* 311, H207–H218.
 141. Afzal, M.R., Samanta, A., Shah, Z.I., Jeevanantham, V., Abdel-Latif, A., Zuba-Surma, E.K., and Dawn, B. (2015). Adult bone marrow cell therapy for ischemic heart disease: evidence and insights from randomized controlled trials. *Circ. Res.* 117, 558–575.
 142. Nasser, M.I., Qi, X., Zhu, S., He, Y., Zhao, M., Guo, H., and Zhu, P. (2020). Current situation and future of stem cells in cardiovascular medicine. *Biomed. Pharmacol.* 132, 110813.
 143. An, Y., Sekinaka, T., Tando, Y., Okamura, D., Tanaka, K., Ito-Matsuoka, Y., Takehara, A., Yaegashi, N., and Matsui, Y. (2019). Derivation of pluripotent stem cells from nascent undifferentiated teratoma. *Dev. Biol.* 446, 43–55.
 144. Rincon, M.Y., VandenDriessche, T., and Chuah, M.K. (2015). Gene therapy for cardiovascular disease: advances in vector development, targeting, and delivery for clinical translation. *Cardiovasc. Res.* 108, 4–20.
 145. Xu, M., and Song, J. (2021). Targeted therapy in cardiovascular disease: A precision therapy era. *Front. Pharmacol.* 12, 623674.
 146. Liu, Y., Wang, S., Wang, C., Song, H., Han, H., Hang, P., Jiang, Y., Wei, L., Huo, R., Sun, L., et al. (2013). Upregulation of M3 muscarinic receptor inhibits cardiac hypertrophy induced by angiotensin II. *J. Transl. Med.* 11, 209.
 147. Oikawa, S., Kai, Y., Mano, A., Ohata, H., Nemoto, T., and Kakinuma, Y. (2017). Various regulatory modes for circadian rhythmicity and sexual dimorphism in the non-neuronal cardiac cholinergic system. *J. Cardiovasc. Transl. Res.* 10, 411–422.
 148. Shaked, I., Meerson, A., Wolf, Y., Avni, R., Greenberg, D., Gilboa-Geffen, A., and Soreq, H. (2009). MicroRNA-132 potentiates cholinergic anti-inflammatory signaling by targeting acetylcholinesterase. *Immunity* 31, 965–973.
 149. Hanin, G., and Soreq, H. (2011). Cholinesterase-Targeting microRNAs Identified in silico Affect Specific Biological Processes. *Front. Mol. Neurosci.* 4, 28.
 150. Manoharan, I., Boopathy, R., Darvesh, S., and Lockridge, O. (2007). A medical health

- report on individuals with silent butyrylcholinesterase in the Vysya community of India. *Clin. Chim. Acta* 378, 128–135.
151. Pytel, E., Bukowska, B., Koter-Michalak, M., Olszewska-Banaszczyk, M., Gorzelak-Pabiś, P., and Broncel, M. (2017). Effect of intensive lipid-lowering therapies on cholinesterase activity in patients with coronary artery disease. *Pharmacol. Rep.* 69, 150–155.
 152. Massoulié, J. (2002). The Origin of the Molecular Diversity and Functional Anchoring of Cholinesterases. *Neurosignals* 11, 130–143.
 153. Perrier, A.L., Massoulié, J., and Krejci, E. (2002). PRIMA: the membrane anchor of acetylcholinesterase in the brain. *Neuron* 33, 275–285.
 154. Kilianova, Z., Ciznarova, N., Szmicekova, K., Slobodova, L., and Hrabovska, A. (2020). Expression of cholinesterases and their anchoring proteins in rat heart. *Can. J. Physiol. Pharmacol.* 98, 473–476.
 155. Prall, Y.G., Gambhir, K.K., and Ampy, F.R. (1998). Acetylcholinesterase: An enzymatic marker of human red blood cell aging. *Life Sci.* 63, 177–184.
 156. Altamirano, C.V., and Lockridge, O. (1999). Association of tetramers of human butyrylcholinesterase is mediated by conserved aromatic residues of the carboxy terminus. *Chem. Biol. Interact.* 119, 53–60.
 157. Liang, D., Blouet, J.P., Borrega, F., Bon, S., and Massoulié, J. (2009). Respective roles of the catalytic domains and C-terminal tail peptides in the oligomerization and secretory trafficking of human acetylcholinesterase and butyrylcholinesterase. *FEBS J.* 276, 94–108.
 158. Li, B., Duysen, E.G., Carlson, M., and Lockridge, O. (2008). The butyrylcholinesterase knockout mouse as a model for human butyrylcholinesterase deficiency. *J. Pharmacol. Exp. Ther.* 324, 1146–1154.
 159. Lockridge, O., and Masson, P. (2000). Pesticides and susceptible populations: people with butyrylcholinesterase genetic variants may be at risk. *Neurotoxicology* 21, 113–126.
 160. Broomfield, C.A., Maxwell, D.M., Solana, R.P., Castro, C.A., Finger, A.V., and Lenz, D.E. (1991). Protection by butyrylcholinesterase against organophosphorus poisoning in nonhuman primates. *J. Pharmacol. Exp. Ther.* 259, 633–638.
 161. Murthy, V., Gao, Y., Geng, L., LeBrasseur, N.K., White, T.A., Parks, R.J., and Brimijoin, S. (2014). Physiologic and metabolic safety of butyrylcholinesterase gene therapy in mice. *Vaccine* 32, 4155–4162.
 162. Hartmann, J., Kiewert, C., Duysen, E.G., Lockridge, O., Greig, N.H., and Klein, J. (2007). Excessive hippocampal acetylcholine levels in acetylcholinesterase-deficient mice are moderated by butyrylcholinesterase activity. *J. Neurochem.* 100, 1421–1429.
 163. Chatonnet, A., and Lockridge, O. (1989). Comparison of butyrylcholinesterase and acetylcholinesterase. *Biochem. J.* 260, 625–634.
 164. Dingová, D., Kucera, M., Hodbod, T., Fischmeister, R., Krejci, E., and Hrabovska, A.P. (2024). Cardiac acetylcholinesterase and butyrylcholinesterase have distinct localization and function. *bioRxiv* 1, 2024. <https://doi.org/10.1101/2024.05.29.596481>.
 165. Sankar, M., Karthikeyan, R., and Vigneshkumar, S. (2023). Synthesis and Characterization of Chitosan Acetylcholine Nanoparticles for Neural Disorders Associated with Cancer Treatment. *J. Inorg. Organomet. Polym.* 33, 2465–2484.
 166. Abdelwahab, G.M., Mira, A., Cheng, Y.B., Abdelaziz, T.A., Lahloub, M.F.I., and Khalil, A.T. (2021). Acetylcholine esterase inhibitory activity of green synthesized nanosilver by naphthopyrones isolated from marine-derived *Aspergillus niger*. *PLoS One* 16, e0257071.
 167. Alami, A.E., Lagarde, F., Huo, Q., Zheng, T., Baitoul, M., and Daniel, P. (2020). Acetylcholine and acetylcholinesterase inhibitors detection using gold nanoparticles coupled with dynamic light scattering. *Sensors Int.* 1, 100007.
 168. Roy, A., Guatimosim, S., Prado, V.F., Gros, R., and Prado, M.A.M. (2015). Cholinergic Activity as a New Target in Diseases of the Heart. *Mol. Med.* 20, 527–537.
 169. Foster, D.J., and Conn, P.J. (2017). Allosteric Modulation of GPCRs: New Insights and Potential Utility for Treatment of Schizophrenia and Other CNS Disorders. *Neuron* 94, 431–446.
 170. Andersson, K.-E. (2004). Antimuscarinics for treatment of overactive bladder. *Lancet Neurol.* 3, 46–53.
 171. Naicker, P., Anoopkumar-Dukie, S., Grant, G.D., and Kavanagh, J.J. (2017). Anticholinergic activity in the nervous system: Consequences for visuomotor function. *Physiol. Behav.* 170, 6–11.