Strategies for Vaccination and Control of Apicomplexan Protozoan Parasites

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Over the past several decades a great deal of effort has been invested in developing new control and vaccination strategies for apicomplexan protozoan parasites. These organisms are the cause of some of the most significant and harmful diseases in both humans and animals worldwide and include as examples; *Plasmodium* (malaria) and *Cryptosporidium* in humans, *Toxoplasma* in humans and animals, *Babesia* and *Neospora* in cattle, and *Eimeria* the cause of coccidiosis in animals (poultry, sheep, cattle, etc.). In spite of a great deal of progress made in understanding at the molecular level how these organisms invade, survive and transmit in their respective hosts, there has been a paucity of new vaccines commercially developed against these parasitic protozoa over the past few decades. In this chapter, we will discuss the types of strategies being developed to help control these parasites, which includes the development of live and subunit (both native and recombinant DNA based) vaccines, the search for and use of new or existing drugs (such as artemisinin combination therapy against malaria), as well as simpler management & hygiene strategies that can be employed to help alleviate the burden of parasitic diseases.

Keywords Apicomplexan parasites; vaccines; drug targets; ecoimmunology; immunotolerance

1. Introduction

As a result of the development of both recombinant DNA and hybridoma technologies in the early 1970s, it was believed that within 5-10 years new vaccines would be developed against malaria as well as several of the other major parasitic diseases affecting human and animal health. Unfortunately, those hopes have yet to be realized and although excellent progress has been made in unravelling some of the mysteries associated with parasite growth and developmental biology, little has been accomplished in the actual control of these devastating pathogens. Indeed, due to the development of drug resistance by the malaria parasite, the number of people infected in the world had actually grown to 244 million in 2005, but then decreased to 225 million in 2009 mainly due to the use of insecticide-treated mosquito nets (ITNs) and indoor residual spraying (IRS). Similarly, the number of deaths from malaria also increased during the 1970s-1990s, and then decreased from 985,000 per anum in 2000, to 781,000 in 2009. However, in some countries in Africa and elsewhere there has continued to be a marked increase in malaria morbidity and mortality over the past 5 years, which has been disappointing for the WHO and others involved in malaria control.

Much research has been devoted to the developmental biology of these parasites (i.e. their life cycle and mechanisms of switching from one developmental stage to another) and how they interact with the host in order to try and discern weaknesses in the parasite's armour. Clearly, parasitic protozoan parasites have come through several million years of coevolution with their hosts and have learned to invade and survive in very hostile environments. For example, the *Toxoplasma* parasite lives within the cell (macrophage) whose sole purpose is to destroy incoming foreign pathogens. *Plasmodium* and *Babesia* parasites have learned to invade host red blood cells or reticulocytes without lysing them using specific subcellular organelles (rhoptries and micronemes) present in the merozoite stage of development, and in that manner evade the host immune system. *Cryptosporidium* and *Eimeria* parasites reside within the intestinal epithelial cell and develop to an oocyst stage after which they are excreted in the feces. In this manner the parasite can survive for months or even years under harsh environmental conditions, enabling them to infect other host animals or humans. These apicomplexan parasites have also learned to move from host to host through arthropod vectors where they are transmitted to entire human or animal populations. In this chapter we will explore these various developmental and host/vector evasion strategies and discuss ways in which to inhibit or even prevent their growth and development.

It is clear that in order to control apicomplexan parasitic protozoa we must be able to rethink many of the current strategies that have been taken up in this battle against these pathogens. As mentioned above, apicomplexan parasites are very capable of evading host immune responses and surviving in hostile environments. In addition, they have learned to deal with antimicrobials produced to destroy them, developing resistance against a whole variety of drugs and toxic compounds. So where does the achilles heel of these parasites lie? This question will be discussed in the context of our current knowledge of host immune evasion and basic parasite biology.

Finally, in this chapter we will look at other potential control strategies involving improvements in how we manage the environment in which these parasites live and are transmitted (i.e. the ecosystem & ecoimmunology). This includes an understanding of the effect of host nutrition, environmental factors as well as infection with multiple pathogens on the immune response. Finally, we will discuss the question of whether or not we can really defeat these parasites or perhaps at least in certain cases we should be looking to develop a strategy of inducing immunotolerance to reduce the harmful effects of the infection. All in all, it is clear that a great deal of new and original research must be designed and carried out in order to help alleviate the terrible health burden and economic cost of these parasitic diseases.

2. Parasite developmental biology

Apicomplexan protozoan parasites undergo their life cycle in various tissues and cell types within the host animal or human. *Plasmodium*, the parasite which causes malaria in humans and animals, first infects the liver cells in the form of sporozoites where they undergo the extraerythrocytic, asexual life cycle. From there, daughter merozoites are released into the blood stream where they go on to infect red blood cells relying on organelles contained in the apical end of the parasite (rhoptries and micronemes) to invade the cell without rupturing it. They go on to form a parasitophorous vacuole where they can reside safe from the host immune response. Within this vacuole the parasite divides asexually producing new daughter cells (merozoites) which are released when the cell bursts open. After several such replication cycles during which the pathogenesis of malaria is manifested (fever and chills, anemia, cerebral malaria, etc.), the parasite once again invades red blood cells and commences the sexual cycle of development forming male microgametes and female macrogametes. These are taken up by a mosquito in a blood meal and in the midgut they undergo fertilization, zygote formation and development into ookinetes, which penetrate the gut wall and develop into single walled oocysts. The parasite undergoes mitotic replication in the oocyst forming numerous sporozoites that exit the oocyst and reach the salivary gland where they are injected into the next human host during the blood meal.

In the case of *Eimeria* and *Cryptosporidium* similar, less complex, developmental life cycles occur in the epithelial cells of the host intestinal mucosa. Here these parasites undergo both the asexual and sexual stages of their development life cycle, and over a period of 4-8 days form infectious oocysts which are secreted in the feces (in the case of *Cryptosporidium*, oocyst shedding can continue for several weeks post infection due to autoreinfection). These highly resistant and stable oocysts are then picked up by other animals or humans due to their presence in either litter, contaminated food, or water supplies. Due to their rapid replication and exponential increase in numbers (for each oocyst ingested up to 1,000,000 new ones are formed and secreted), the parasite is transmitted through the population where it can cause severe disease due to the rupture of host cells, secretion of toxic substances, as well as by inducing immunopathology. Indeed, it is due to the intense rearing practices of domestic animals, poor hygiene practices, and in the case of *Cryptosporidium*, difficulty to prevent the introduction of these parasites into drinking water due to their resistance against most disinfectants and very small size, that these parasites have caused such widespread damage.

Toxoplasma is an example of a parasite where in its definitive host, the cat, it undergoes its developmental cycle in epithelial cells of the intestinal tract, producing oocysts that are secreted in the feces into the environment. Once the oocysts are ingested by an intermediate host such as rodents, cattle or sheep, the sporozoite penetrates the intestine, is disseminated in the host and finally develops into tissue cysts where it can survive in undercooked meat until ingested by humans, cats or other animals. In the human or animal intermediate host, the parasite will develop in tissues and the bloodstream as tachyzoites, during which time the parasite invades macrophages and survives in phagosomes by preventing fusion with lysosomes as well as increasing the longevity of its host cell. After induction of a protective immune response by the host, they form bradyzoites that develop into cysts in the musculature, heart and brain. There they can reside for the entire lifetime of the host until activated either due to immunosuppression by viruses such as HIV or immunosuppresive drugs, forming tachyzoites that can directly damage host cells as well as induce an immunopathological response. This can lead to severe problems in the brain (encephalitis), eyes (chorioretinitis) as well as problems in the inner ear, heart and liver.

Over the past several years, live vaccines have been developed for some of these parasitic diseases. In the case of Coccidiosis, these vaccines were developed based on the finding that a single infection with an *Eimeria* species induces a very high level of immunity against reinfection against the homologous species [1]. As one example of a live coccidiosis vaccine, Paracox® was developed in which attenuated lines for each of the 7 species of *Eimeria* that cause the disease were combined and delivered to chickens via the drinking water [2]. Good results were obtained in that chickens that were vaccinated at 1 day of age with Paracox® were solidly immune against infection and disease by 2-3 weeks of age.

A similar approach has been taken for malaria where it has been shown that live infection can induce good immunity against reinfection. Researchers in Australia and the USA have begun the development of a live vaccine using parasites produced in tissue culture [3]. However, it is difficult to produce live malaria parasites in vitro at a low cost. In addition, storage, transport and use of such a vaccine in endemic countries would be very difficult to implement. Furthermore, malaria parasites have been shown to undergo antigenic variation so that it may be necessary to combine a variety of parasite strains in any live vaccine. In spite of these problems, it is hoped that a program to vaccinate against malaria in endemic countries using a live vaccine may be achievable in the future.

The fact that live vaccines can induce solid immunity show that there are chinks in the armour of these parasites that can be exploited to produce subunit or recombinant DNA based vaccines to inhibit or even prevent their growth and development. In this regard, one possible approach is to try and induce an immune response to prevent parasite invasion of the host cell by sporozoites, tachyzoites and/or merozoites. A great deal of information is available on the molecular composition of the organelles involved in invasion; the rhoptries, micronemes and dense granules. Many of these

proteins are well conserved between apicomplexan species and even genera, and several have been tested (either as a native or recombinant antigen or as a DNA based formulation) as vaccine candidates against a variety of apicomplexan parasites [4,5]. However, either partial or no significant levels of protection was observed in those studies. It is currently believed that by improving the methods of antigen presentation and delivery, better results with higher levels of protection can be obtained.

Another approach to vaccination against these parasites is to try and inhibit cyst/oocyst wall formation thereby reducing or blocking parasite transmission. In this regard, a subunit vaccine using purified antigens from the gametocyte and ookinete stages of development of Plasmodium gallinaceum and P. falciparum were tested to see if they can reduce the number of oocysts in mosquitos infected with blood from vaccinated birds or humans [6,7]. The results showed that a very high level of inhibition can be achieved reaching over 90% reduction of transmission. In *Eimeria*, vaccination of laying hens with gametocyte antigens or by live infection to induce a high level of maternal IgY antibody in their eggs, greatly reduced oocyst production in offspring chicks (by up to 80% using purified gametocyte glycoproteins to vaccinate the hens). This finding led to the production of a subunit vaccine containing 3 purified, native gametocyte antigens (CoxAbic[®]), which has already been tested and proven to be effective in controlling Coccidiosis in millions of broiler chickens around the world [8]. It was hoped that this approach could also be used to control diseases caused by other apicomplexan parasites in humans. However, the use of transmission blocking immunity in the control of *Plasmodium*, *Toxoplasma* or *Cyptosporidium* is much more difficult since it requires the induction of an antibody response that remains high for decades, while a chicken is only raised for 5-8 weeks of age before it is slaughtered for meat production. In addition, for malaria control a transmission blocking approach would require a great deal of eduction to convince people to participate in such a program in which no direct benefits are felt by the vaccinated individual. Finally, for Toxoplasma cats would need to be vaccinated to block parasite transmission.

3. Use of drugs in the control of apicomplexan parasites

Over the past 70 years, antimicrobial/antibiotic drugs have been developed to control diseases caused by apicomplexan parasites. In the case of malaria, in the late 1940s chloroquine was put into use and was extremely effective in combating this parasite. However, within 10 years of its use drug resistance appeared in Africa which spread worldwide. It was subsequently found that this resistance is based on point mutations occuring in key transport proteins (*pfcrt* and *pfmdr 1*), demonstrating the ability of the parasite to undergo mutations enabling its survival [9]. Similarly, the *Eimeria* parasite has become resistant against a multitude of anticoccidial drugs including potent ionophores, leading to the need for rotation and shuttle programs to reduce the build up of resistant parasites in the litter [10].

More recently artimisinin and artimisinin related compounds have been developed and used for the treatment of malaria, however, the WHO have already recommended that drug combination therapy be implemented to reduce the build up of drug resistance by the parasite. Once again mutations in transport proteins have accounted for the resistance phenotype [11]. New artimisimin related drugs and drug combinations are still being developed, however, only through strategic and limited use of these compounds will their effectiveness be preserved [12]. This is of course not only true for the malaria parasite, but for all parasites where new drugs are being developed. This coupled with the unwillingness of the pharmaceutical industry to invest large sums of money into developing new compounds, is leading to a situation in which we will have no effective means of treating malaria and other parasitic diseases.

4. Rethinking current strategies

As was mentioned above, apicomplexan parasites have been around for millions of years and have learned to grow and survive within a whole variety of hosts and environments. The strategies we have developed thus far to try and control them have largely been based on concepts that do not take into account the fundamental ability of these organisms to adapt, mutate and evade host immunity or environmental stresses. For example, parasite invasion is an attractive target to try and inhibit development, however, the organelles and proteins produced are well recognised by the host immune system, yet the parasite can still evade such an immune response.

Recent studies have indicated that it is not only a matter of the mounting of an immune response, but making sure it is of the right type and specificity. With regards to the cellular arm of the immune response, it has been found that the type of T cells and effector molecules and cytokines elicited can play a key role in determining the outcome of the infection [13]. This includes a proper balance between activation of 2 types of T helper cells (Th1 and Th2). Indeed, a given pathogen will employ a strategy to induce an immune response that benefits its survival such as by excreting substances that can inhibit the immune system, that activate the appropriate Th1 or Th2 cells for its own survival, as well as molecules that act to overstimulate the host immune system diverting it from the appropriate response. In the case of parasites such as *Eimeria* and *Cryptosporidium*, it was found that the host immune response can either play a key role in exacerbating intestinal lesions or inhibiting them depending on the type of immune response and host genetic background [14]. In addition, when multiple pathogens are present in the same host, the immune response becomes highly complex and unpredictable and may act to worsen one type of infection while preventing another [15].

A great deal of work using animal model systems with mixed infections have been performed, where it was found that the establishment and intensity of infection was effected by the combination of pathogens used (bacteria, virus, helminth or protozoan parasite), the strain of parasite being tested (highly virulent, mild, etc.), the host genetic background as well as the order in which the infectious organisms were administered.

So where does the achilles heel of these parasites lie? One approach already mentioned above is in the prevention or inhibition of cyst/oocyst wall formation. One thing is for certain, if an apicomplexan parasite cannot form its cyst/oocyst wall, it will not survive in the environment and be transmitted. A great deal of progress has been made in elucidating the molecular compostion and architecture of the oocyst wall of *Eimeria* and other apicomplexan parasites [15]. It is hoped that apart from immunological approaches to prevent oocyst formation, new drug targets can be discovered that may act to completely disrupt parasite developmental biology. Recent results showing artimisinin can prevent oocyst formation in *Eimeria* is very encouraging [16], and hopefully new drugs that are extremely potent in preventing cyst wall formation will be discovered and utilized.

Another potential strategy is by focusing on the pathogenicity of these parasites rather than trying to prevent their growth & development. For example, if the deleterious effect of *Cryptosporidium* is at least partly due to the toxin it secretes, then by either producing antibodies against these toxins or inhibitory drugs designed to block the toxic effect, then a new approach to controlling the severe diarrhea caused by this parasite may be accomplished [17]. In the case of *Plasmodium falciparum*, a great deal of the pathogenicity is due to its ability to attach to host cells and sequester itself away from the host immune system (the parasite modifies the erythrocyte surface membrane to bind to capillaries in the brain and thereby remove the infected host cell from the circulation). The sequestered parasites can then act to either directly block circulation in these capillaries, or to elicit an immunopathological response by host lymphocytes and macrophages leading to inflammation [18]. This process leads to the disease state known as cerebral malaria, which in severe cases can lead to coma and death. A vaccine that would induce antibodies that can block the binding of the parasitised red cell to the host cell membrane, or through the use of inhibitory peptides, etc., would alleviate this blockage in brain circulation and prevent the disease from occurring.

5. The role of management practices and the environment in parasite control.

Until vaccines or new effective drugs against most of these pathogenic protozoan parasites are developed, it is necessary to try and reduce infectivity and transmission through good management practices and improvements in environmental control. For example, simply by using insecticide impregnated bed nets and indoor residual spraying has led to a dramatic decrease in malaria transmission and a reduction in morbidity and mortality in countries that have implemented such a program effectively [19]. This together with the development of better, cheaper, and easier to use diagnostic procedures, has greatly helped in the fight against malaria worldwide.

In the case of veterinary diseases such as Coccidiosis or Neosporidiosis, good litter management in the case of *Eimeria* and better separation of cattle from infected dogs or dog feces for *Neospora*, has reduced the burden of these diseases on farms. In the case of raising chickens, the quality of the shed in which they are grown, the use of good ventilation systems, etc., have all helped to keep the litter dry and thereby reduce the effect of coccidiosis on flock performance. Cattle also need to be raised in well maintained sheds, and constantly monitored for exposure to a variety of pathogens and other environmental factors that can cause disease and stress.

Finally, good nutrition, breeding, and hygiene all play key roles in controlling infection by apicomplexan parasites. In Africa where poor nutrition coupled with multiple infections by immunosuppressive viruses, bacteria, helminthes and fungi, act to greatly exacerbate the rate of malaria transmission as well as the virulence of the infection. In animals, raw materials used in feed may contain endotoxins and other immunosuppressive factors. In addition, animal feed that is produced without proper quality control procedures in place may have lowered levels of key nutrients such as trace metals, amino acids, vitamins, etc. This may have a negative effect on the host immune response, which in turn can greatly effect the performance of flocks or herds.

6. Can we learn to live in harmony with apicomplexan parasites?

What is the explanation behind the fact that some individual animals or humans can harbour parasites even in large number without any detremental effect, while in others there are severe pathological consequences even from a light infection? One possible explanation is immunotolerance. For example, if cerebral malaria caused by *Plasmodium falciparum* in humans is at least in part due to an overly active immune system, than the induction of immunotolerance may reverse this pathogenic effect in the brain. The question is how to induce immunotolerance in a manner that benefits the host, without causing an increase in parasite load and as a result heightened pathogenicity?

In this regard, Ecoimmunology can be applied to ask the question what is the ideal type, quantity and quality of the immune response in controlling parasitic diseases, and what is an acceptable parasite load. From this point of view it can be argued that the complete killing off of all parasites is not necessarily good for the host; indeed host fitness may be maximized at a point of low parasite load together with only a medium or even relatively low immune response [20].

As mentioned above, the immune system can act as a two edge sword being both protective and pathological. Similarly, hosts that are exposed to parasites in a controlled manner can benefit from the induction of the appropriate immune response. The question from an ecologocal perspective is what is the nature of the host-parasite relationship and what drives it towards either harmony or one of disharmony and disease.

A good example of this is the application of the vaccine against *Eimeria*, CoxAbic®. This vaccine was developed in order to control the rate of parasite transmission in a chicken house and not to try and eradicate the parasite. Results from field studies carried out around the world showed that chicks from vaccinated hens, produced fewer numbers of oocysts during the first 3 weeks of growth due to the maternal antibody, and after the antibody waned they maintained a low level of oocysts in the litter that rather than causing disease in the birds induced an appropriate, protective immune response [8]. Thus, by inhibiting parasite transmission, oocyst numbers decreased by 60-80% in the litter which translated into good health and performance of the flocks.

The other important point to be made is that we must look at these parasitic diseases in the context of the whole population. As seen for CoxAbic®, by reducing parasite numbers in the litter, you can provide disease control for the whole flock. It is still possible that within a vaccinated flock there will be a few individual birds that develop mild disease, but overall there is a marked improval in growth, well-being and performance. Similarly, if we were able to do the same thing for malaria, we would greatly reduce the morbidity and mortality caused by the disease and then when necessary focus on the individuals that still require medical attention and care.

Finally, any measure taken to help control apicomplexan parasites must take into account the possibility that a reduction in parasite burden may induce an increase in parasite growth rates and even parasite virulence. Thus, any new measures must be applied with extreme caution with properly controlled field or clinical trials performed over lengthy periods of time. These parasites have already demonstrated their capabilities in adapting to adverse host and environmental conditions and do everything they can to continue to maintain their presence in human or animal populations. Therefore, well designed mathematical modelling and epidemiological studies must be used to accompany any new parasite control program.

7. Conclusions

Parasites have evolved over millions of years in order to be able to propagate and survive in hostile environments. Inherent in this survival is maintaining a proper balance between growth and development within its host animal or human, with its rate of transmission to new hosts. This also includes the need for the parasite to ensure that it does not cause undo harm to its host so that its own survival is ensured.

With that in mind, the host immune system has also developed in order to fight external pathogens while minimizing the harm caused by an overreaction to antigenic stimuli leading to immunopathology. Thus, simply by increasing the response to antigens which normally elicit an immune reaction is not going to be very effective in controlling infections and may indeed have the reverse effect. New approaches to immunotherapy need to be developed including the induction of immunotolerance to ensure a healthy balance between host and pathogen.

Clearly, environmental and management practices that can help reduce parasite transmission without effecting the virulence of the parasite is a useful practice. However, it must be remembered that by reducing parasite numbers in the environment could induce an increased rate of replication and transmission of these pathogenic organisms. This in turn may lead to a situation in which fewer individuals are infected, however, the pathogenicity in that individual may be increased. Therefore, any new practice should be implemented with caution and continuous monitoring backed up by well designed epidemiological studies.

Finally, the development of new vaccines and drugs to control apicomplexan parasites has been the goal of researchers for the past 40 years. Large government grants together with support from major pharmaceutical companies, have pushed this field forward greatly, however, we are still a long way off from being able to control malaria and other human and veterinary parasites. Clinical studies for malaria are in progress using existing antigens from both the sporozoite and merozoite stages of development, however, so far the results are not very encouraging. We must learn from this work how to bridge the work in the lab with what happens in the field and use it to develop better means of vaccination and control. In that way, once we do find the achilles heel of the parasite we will know how to implement its use to start to finally win the battle against apicomplexan parasites.

References

- [1] Rose ME, Long PL. Immunity to coccidiosis: protective effect of transferred serum and cells investigated in chick embryos infected with *Eimeria tenella*. *Parasitology*. 1971;63:299-313.
- [2] Shirley MW, Bedrnick P. Live attenuated vaccines against avian coccidiosis: Success with precocious and egg-adapted lines of *Eimeria. Parasitology Today.* 1997; 13:481-484.
- [3] Pombo DJ, Lawrence G, Hirunpetcharat C, Rzepczyk C, Bryden M, Cloonan N, Anderson K, Mahakunkijcharoen Y, Martin LB, Wilson D, Elliott S, Elliott S, Eisen DP, Weinberg JB, Saul A, Good MF. Immunity to malaria after administration of ultralow doses of red cell infected with *Plasmodium falciparum*. *Lancet*. 2002;360:610-617.

- [4] Ridley RG, Takacs B, Etlinger H, Scaife JG. A rhoptry antigen of *Plasmodium falciparum* is protective in *Saimiri* monkeys. *Parasitology*. 1990;101:187-192.
- [5] Alaeddine F, Keller N, Leepin A, Hemphill A. Reduced infection and protection from clinical signs of neosporosis in C57BL/6 mice vaccinated with recombinant microneme antigen NcMic1. *Journal of Parasitology*. 2005;91:657-665.
- [6] Carter R, Gwadz RW, Green I. *Plasmodium gallinaceum:* Transmission-blocking immunity in chickens: II. The effect of antigamete antibodies *in vitro* and *in vivo* and their elaboration during infection. *Experimental Parasitology.* 1979;47:194-208.
- [7] Rener J, Graves PM, Carter R, Williams JL, Burkot TR. Target antigens of transmission-blocking immunity on gametes of Plasmodium falciparum. Journal of Experimental Medicine. 1983;158:976-981.
- [8] Wallach MG, Ashash U, Michael A, Smith NC. Field Application of a subunit vaccine against an enteric protozoan disease. PLoS ONE. 2008; 3(12): e3948. doi:10.1371/journal.pone.0003948.
- [9] Nsobya SI, Kiggundu M, Nanyunja S, Joloba M, Greenhouse B, Rosenthal PJ. In vitro sensitivities of *Plasmodium falciparum* to different antimalarial drugs in Uganda. *Antimicrobial Agents and Chemotherapy*. 2010;54:1200-1206.
- [10] Bafundo K, Cervantes HM, Mathis GF. Sensitivity of Eimeria field isolates in the United States: Responses of nicarbozincontaining anticoccidials. Poultry Science. 2008;87:1760-1767.
- [11] Sidhu AB, Uhlemann AC, Valderramos SG, Valderramos JC, Krishna S, Fidock DA. Decreasing pfmdr1 copy number in *Plasmodium falciparum* malaria heightens susceptibility to mefloquine, lumefantrine, halofantrine, quinine and artemisinin. *Journal of Infectious Diseases*. 2006;194:528-535.
- [12] Bellot F, Cosledan F, Vendier L, Brocard J, Meunier B, Robert A. Trioxaferroquines as new hybrid antimalarial drugs. Journal of Medicinal Chemistry. 2010;53:4103-4109.
- [13] Denkers EY, Gazzinelli RT. Regulation and function of T-cell-mediated immunity during Toxoplasma gondii infection. Clinical Microbiology Reviews. 1998;11:569-588.
- [14] Lillehoj HS, Ruff MD, Bacon LD, Lamont SJ, Jeffers TK. Genetic control of immunity to Eimeria tenella. Interaction of MHC genes and non MHC-linked genes influences levels of disease susceptibility in chickens. Veterinary Immunology and Immunopathology. 1989;20:135-148.
- [15] Ferguson DJP, Belli SI, Smith NC, Wallach MG. The development of the macrogamete and oocyst wall in Eimeria maxima: immuno-light and electron microscopy. International Journal for Parasitology. 2003;33:1329-1340.
- [16] del Cacho E, Gallego M, Francesch M, Quilez J, Sanchez-Acedo C. Effect of artemisinin on oocyst wall formation and sporulation during *Eimeria tenella* infection. *Parasitology International*. 2010;59:506-511.
- [17] Guarino A, Canani RB, Casola A, Pozio E, Russo R, Bruzzese E, Fontana M, Rubino A. Human intestinal cryptosporidiosis: secretory diarrhea and enterotoxic activity in Caco-2 cells. Journal of Infectious diseases. 1995;171:976-983.
- [18] Ochola LB, Siddondo BR, Ocholla H, Siana N, Kimani EN, Williams TN, Makale JO, Liljander A, Urban BC, Bull P, Szestak T, Marsh K, Craig AG. *Plasmodium falciparum* cytoadherence to ICAM-1 is associated with cerebral malaria. *Malaria Journal*. 2010;9:27.
- [19] Russell TL, Lwetoijera DW, Maliti D, Chipwaza B, Kihonda J, Charlwood JD, Smith TA, Lengeler C, Mmanyangala MA, Nathan R, Knols BGJ, Takken W, Killeen, GF. Impact of promoting longer-lasting insecticide treatment of bed nets upon malaria transmission in a rural Tanzanian setting with pre-existing high coverage of untreated nets. *Malaria Journal*. 2010;9:187.
- [20] Graham AL, Shuker DM, Pollitt LC, Auld SKJR, Wilson AJ, Little TJ. Fitness consequences of immune responses: strengthening the empirical framework for ecoimmunology. Functional Ecology. 2011;25:5-17.

INTRODUCTION

The present edition is the third number of a Microbiology book series, which belongs to a more general line of books published by Formatex (Badajoz, Spain), aimed at communicating current scientific and technological research in a generalistic-didactic way. In this number, it aims at stimulating the presentation, exchange and dissemination of information and experiences on anti-microbe strategies (against bacteria, fungi or protozoans), in biotic or abiotic environments, in planktonic or adhered states, in biologically specific or unspecific ways, in vitro or in vivo, in a general context marked by the threat posed by the increasing antimicrobial resistance of pathogenic microorganisms. "Anti" is here taken in a wide sense as "against cell cycle, adhesion, or communication", when harmful for human health, industry or economy (infectious diseases, chemotherapy, food, biomedicine, agriculture, livestock, biotechnology, water systems...). It will include topics on antimicrobial resistance, (early) microbial and resistance detection, enhancement of innate defences against pathogens, as well as methods & techniques.

The topics covered are:

Antimicrobial chemistry (experimental and computational). Analytical detection of antibiotics in complex samples.

Synthesis and screening of novel chemical compounds for antimicrobial action. Natural, synthetic and semisynthetic antibiotics. Analogs. Structural determination. *In-silico/ab-initio/de-novo* antimicrobials discovery. New targets for antimicrobials. Rational design of antimicrobials. Bioinformatics and comparative genomics for the identification of antimicrobial targets...

Antimicrobial natural products.

Antimicrobial substances from terrestrial and marine organisms. Antimicrobial peptides. Antimicrobial enzymes. Essential oils. Bioactive phytochemicals. Plant/Herbal extracts. Purification. Structural determination...

Antimicrobials mechanisms of action.

Methods and Techniques.

Antimicrobial resistance. Superbugs. Multi-resistant strains. Emerging and re-emerging pathogens.

Microbial resistance to antibiotics and biocides. Molecular mechanisms. Resistance genes. Prevention of resistance. Surveillance & statistics. Genetics and Proteomics. Emerging and re-emerging bacteria and fungi in humans, animals, and plants. Methicillin-resistant *Staphylococcus aureus* (MRSA), Vancomycin Intermediate/Resistant *Staphylococcus aureus* (VISA/VRSA), *Clostridium difficile, Mycobacterium tuberculosis*, Vancomycin-resistant *enterococcus* (VRE), *Cryptosporidium, Plasmodium parasite, Plasmodium falciparum, Leishmania* species, *Klebsiella pneumoniae, Streptococcus pneumoniae, Acinetobacter baumannii, Cryptococcus, Escherichia coli* O157:H7, *Helicobacter* spp., *Enterobacter sakazakii, Serratia* spp., Fluoroquinolone-Resistant *Pseudomonas aeruginosa* (FQRP)...

Antimicrobial microbes.

Microbial-derived toxins. Bacteriocins (colicins, microcins, lantibiotics...). Archaeocins. Biocontrol approach to microbial invasions (probiotics, lactic acid bacteria...). Biosynthesis of antibiotics. Genetic and metabolic engineering. Gene regulation...

Antimicrobial viruses.

Bacteriophages. Phage therapy and biocontrol in humans, animals (agriculture-farm animals, aquaculture), plants, food industry... Materials functionalization with bacteriophages. Using bacteriophages for microbiological detection...

Antimicrobial materials science and surface chemistry. Biofilms.

Antimicrobial, anti-adhesive surfaces & coatings. Microbial adhesion to surfaces. Biofouling. Biofilm formation, control and eradication. Novel characterization techniques. Physical and chemical (inorganic (e.g.silver, copper compounds) and organic) surface modification. Cationic surfaces. Functionalization strategies for polymers, metals, metal oxides, ceramics. Drug-eluting concepts. Biofilms susceptibility to antimicrobials. Antibiotic resistance of microorganisms in biofilms. Genomics and Proteomics...

Antimicrobials in consumer products.

Textiles (hygienic clothing, activewear, medical textiles...), paper industry, active packaging (food industry...), public buildings (hospitals, schools, restaurants, day care centers, nursing homes...). Safety and toxicological aspects...

Antimicrobial physics.

Exploitation of physical properties for killing/inactivating microbes: surface tension (nano emulsions), radiation, ultrasounds, temperature, specific properties of nano-materials (nano-particles, nano-tubes/wires, nano-crystals, nano-grained materials...). Resistance to physical agents...

Non-antibiotic biocides. Hygiene and Sterilizing.

Disinfectants, antiseptics, preservatives... Mechanism of action. Resistance to non-antibiotic biocides. Combination of physical and chemical treatments. Hygiene and Sterilizing. Sanitizers. Regulatory issues. Good practices...

Techniques and Methods.

Susceptibility Testing. Rapid microbial and resistance detection. Detection of antibiotics in environmental samples. Microscopy, microanalysis & spectroscopy, single-cell studies, high-throughout studies, nanomechanical studies, microfluidics, lab-on-a-chip concepts, miniaturized science, analysis of microbial surfaces, heterogeneity, statistics. Interaction of antimicrobial drugs with model membranes. Analytical techniques...

The Intelligent war.

Interfering microbe-microbe communication (quorum sensing) as antimicrobial strategy.

Strengthening of innate immune system as antimicrobial strategy.

Immunotherapy, immunomodulating agents, cytokines (interleukins, colony-stimulating factors, interferons...), hormones... Novel vaccines for preventing or treating disease...

Antimicrobials evaluation. Pre-clinical and clinical trials.

Public awareness, learning & teaching, influence on policy-makers. Regional regulatory frameworks and experiences on antimicrobials.

We hope that you will find the articles included in this third edition interesting and stimulating, and look forward to receiving new proposals for future editions,

A. Méndez-Vilas, Editor

CONTENTS

VOL. 1

| Introduction | XVII |
|---|-------|
| 1111 ouucuonin antina antin | 11,11 |

Chemical and physical agents

| Antimicrobial activity of natural photosensitizing anthraquinones S.C. Núñez Montoya, L.R. Comini and J.L. Cabrera | 3-13 |
|--|--------|
| Examining the efficacy of silver and cadexomer iodine dressings in treating wounds compromised by bacterial burden: A review of the literature C. Miller | 14-22 |
| Mefloquine derivatives : synthesis, mechanisms of action, antimicrobial activities Alexandra Dassonville-Klimpt, Alexia Jonet, Marine Pillon, Catherine Mullié and Pascal Sonnet | 23-35 |
| Antimicrobial efficiency of functionalized cellulose fibres as potential medical textiles Tijana Ristić, Lidija Fras Zemljič, Monika Novak, Marjetka Kralj Kunčič, Silva Sonjak, Nina Gunde Cimerman and Simona Strnad | 36-51 |
| Fusogenic liposomes as new carriers to enlarge the spectrum of action of antibiotic drugs against Gram-negative bacteria Rosario Pignatello, Daria Nicolosi and Vito Mar Nicolosi | 52-60 |
| Antifungal free fatty acids: A Review Carolina H. Pohl, Johan L.F. Kock and Vuyisile S. Thibane | 61-71 |
| Quinoline scaffold as a privileged substructure in antimicrobial drugs R. Musiol, T. Magdziarz and A. Kurczyk | 72-83 |
| Immobilized Antimicrobial Agents: A Critical Perspective John-Bruce D. Green, Timothy Fulghum and Mark A. Nordhaus | 84-98 |
| Antimicrobial polymers for textile products A. Varesano, C. Vineis, A. Aluigi and F. Rombaldoni | 99-110 |

| Psychoactive drugs against effects of stress in infectious and non-infectious viral | |
|--|---------|
| diseases S. Novío, M.J. Núñez-Iglesias and M. Freire-Garabal | 111-121 |
| Polymer materials against the microorganism's attack Yu. Savelyev | 122-134 |
| Novel immune-pharmacological approaches for the treatment of bacterial invasive infections. Mónica D. Sparo and Sergio F. Sánchez Bruni | 135-143 |
| Antibacterial activity of materials synthesized from clay minerals M. E. Parolo, L. G. Fernández, I. Zajonkovsky, M. P. Sánchez and M. Baschini | 144-151 |
| The use of biocides for the protection of library documents: before and now T. Velikova, E. Trepova and T. Rozen | 152-159 |
| The structural requirement of direct InhA inhibitors for high potency against <i>M.</i> <i>Tuberculosis</i> based on computer aided molecular design A. Punkvang, P. Kamsri, A. Kumkong, K. Kunasa, P. Saparpakorn, S. Hannongbua, P. Wolschann and P. Pungpo | 160-168 |
| Structural modification of organic compounds by chemical synthesis to develop new antimicrobials. J.C. Espinoza-Hicks, A. Camacho-Dávila, G.V. Nevárez-Moorillón | 169-175 |
| Mini review: Antimicrobial strategies in the production of fresh-cut lettuce products Ö. Tirpanalan , M. Zunabovic, K. J. Domig and W.Kneifel | 176-188 |
| Nanostructured Carriers for Photodynamic Therapy Applications in microbiology João Paulo Figueiró Longo, Luis Alexandre Muehlmann and Ricardo Bentes de Azevedo | 189-196 |
| Nanoparticles and their potential application as antimicrobials Ravishankar Rai V and Jamuna Bai A | 197-209 |
| Metal nanostructures as antibacterial agents J. Díaz-Visurraga, C. Gutiérrez, C. von Plessing and A. García | 210-218 |
| Bactericidal silver nanoparticles present an antiangiogenic effect in the Chorioallantoic Membrane Model (CAM) S. E. A. Will, P. O. Favaron, M. A. Pavez, L. C. Florentino, D. Soares, F. C. Oliveira, R. E. G. Rici, M. A. Miglino, D. Alcântara, E. M. Mamizuka, R. S. Silva, I. M. Cuccovia, D. A.Maria and L. F. Gomes. | 219-227 |
| Bionanoparticles: synthesis and antimicrobial applications K. Sahayaraj and S. Rajesh | 228-244 |

| Antimicrobial activity of aluminium oxide nanoparticles for potential clinical | |
|--|---------|
| applications Amitava Mukherjee, Mohammed Sadiq I., Prathna T.C. and N. Chandrasekaran | 245-251 |
| Phenotypic switching: an opportunity to bacteria thrive A. M. Sousa, I. Machado and M. O. Pereira | 252-262 |
| Research on ozone application as disinfectant and action mechanisms on wastewater microorganisms | |
| M. N. Rojas-Valencia | 263-271 |
| Gamma radiation against toxigenic fungi in food, medicinal and aromatic herbs S. Aquino | 272-281 |
| Solar technologies for plant microbial pathogens inactivation on water M. I. Polo-López, I. García-Fernández, P. Fernández-Ibáñez | 282-290 |
| Factors that Influence the Electric Field Effects on Fungal Cells Maricica Stoica, Gabriela Bahrim and Geta Cârâc | 291-302 |
| Determination of heavy metals and other indicators in waters, soils and medicinal plants from Ave valley, in Portugal, and its correlation to urban and industrial pollution | |
| Pinto D, Fernandes A, Fernandes R, Mendes I, Pereira S, Vinha A, Herdeiro T, Santos E and Machado M. | 303-309 |

Clinical microbiology

| Antimicrobial Sutures: New Strategy in Surgical Site Infections Chatchai Mingmalairak | 313-323 |
|---|---------|
| Encapsulation of a biocide in a starch- oil microemulsion lotion: antimicrobial activity and clinical safety of benzalkonium chloride John J. Wille. | 324-330 |
| Extended-spectrum β-lactamase-producing bacteria: an emerging clinical concern Yong Chong | 331-337 |
| Treatment of respiratory tract infections with fluoroquinolones in Belgium S. Simoens | 338-343 |

| Separation of human immunodeficiency virus type 1 (HIV-1) from motile sperm using a continuous density gradient and subsequent swim-up | |
|--|---------|
| Naoaki Kuji, Shingo Kato, Hideji Hanabusa and Yasunori Yoshimura | 344-352 |
| | |
| Viral infections in the pediatric oncology patient Maria Moschovi, Maria Adamaki and Ioannis Kopsidas | 353-362 |
| , | |
| Clinical significance and antimicrobial susceptibility of rapidly growing mycobacteria | |
| L. García-Agudo and P. García-Martos | 363-377 |
| | |
| Novel Intervention Strategy against Tuberculosis: Insights from Graph Theory and Systems Biology | |
| Veeky Baths and Utpal Roy | 378-385 |
| | |
| Mini-Review: Biological control of bovine mastitis using bacteriophage therapy I.H. Basdew and M.D. Laing | 386-393 |
| I.H. Dasdew and M.D. Laing | 380-393 |
| Risk assessment and new developing strategies to reduce prevalence of <i>campylobacter</i> | |
| spp. In broiler chicken meat Djamel Djenane and Pedro Roncalés | 394-406 |
| | 394-400 |
| Adverse drug reactions following immunization in Danish children: retrospective | |
| analysis of spontaneous reports submitted to the Danish Medicines Agency | |
| Lise Aagaard, Erik Wind Hansen and Ebba Holme Hansen | 407-413 |
| Antimicrobial properties of plasma rich in growth factors (PRGF-ENDORET) | |
| technology | |
| Anitua E, Muruzabal F and Orive G | 414-421 |
| And the disc of Decision Decision of New 1 March 1996 Address of the State of the | |
| Application of Prime-Boost as a Novel Vaccination Strategy Against Microbial Pathogens | |
| De Gaspari | 422-428 |
| | |
| New Strategies to Control Vascular Catheter-Related Bloodstream Infection with Emphasis on Neonatal Intensive Care Unit | |
| M. L. Ribeiro de Souza da Cunha and L. T. Pazzini | 429-439 |
| | |
| Behaviour against β-lactams in <i>Aeromonas</i> spp. isolated from extraintestinal infections | |
| M. Quiroga and M. Vergara | 440-443 |
| | |
| Antimicrobial susceptibility of Streptococcus agalactiae isolated from pregnant | |
| women in Misiones, Argentina M. Quiroga, E. Pegels, P. Oviedo, M. Laczeski and M. Vergara | 444-447 |
| | |

| Importance of pre-operative skin and nail preparation of the foot and intra-operative surgical irrigation in reducing infection after surgical nail avulsion R. Becerro de Bengoa Vallejo, M.E. Losa Iglesias, L. Alou Cervera, D. Sevillano | |
|--|---------|
| Fernandez and J. Prieto Prieto | 448-451 |
| | |
| Selective Decontamination of the Digestive Tract (SDD), a standard of care Richard E Sarginson, Andy J Petros, Nia Taylor, Mark A Fox, Ian Weir, Luciano Silvestri, | |
| Hendrick KF van Saene and Miguel Angel de la Cal | 452-468 |
| Antimicrobial implants and bone allografts: new uses for old antibiotics | |
| Constantinos Ketonis, Noreen Hickok and Javad Parvizi | 469-482 |
| War against mastitis: Current concepts on controlling bovine mastitis pathogens | |
| Cristina Bogni, Liliana Odierno, Claudia Raspanti, José Giraudo, Alejandro Larriestra, | |
| Elina Reinoso, Mirta Lasagno, Mirian Ferrari, Edith Ducrós, Cecilia Frigerio, Susana Bettera, Matías Pellegrino, Ignacio Frola, Silvana Dieser and Claudina Vissio | 402 404 |
| Dettera, iviatias i enegrino, ignacio i ioia, Silvana Dieser and Ciaudina vissio | 483-494 |
| Molecular techniques for detection and control of nosocomial infections caused by | |
| <i>Acinetobacter baumannii</i> E. Sevillano and L. Gallego | 495-503 |
| L. Sevinano and L. Ganego | 495-505 |
| Sesamol attenuates systemic inflammation-associated acute kidney injury in | |
| polymicrobial infectious rats Y. H. Li, D. Z. Hsu and M. Y. Liu | 504-510 |
| | 504-510 |
| Genomics and proteomics approaches to understand virulence of <i>Entamoeba</i> | |
| <i>histolytica.</i> C. López-Camarillo, E. Azuara-Liceaga, A. Zamorano, O. Hernández de la Cruz, I. López | |
| Rosas and L. Marchat | 511-519 |
| | |
| Combination therapy: Synergism between natural plant extracts and antibiotics against infectious diseases | |
| Sumitra Chanda and Kalpna Rakholiya | 520-529 |
| In situ antimicrobial activity of chlorhexidine in the oral cavity | |
| Tomás I, Rubido S and Donos N | 530-541 |
| Chitosan as an oral antimicrobial agent | |
| Morgana Maria Souza Gadelha de Carvalho, Thayza C. Montenegro Stamford, Emerson | |
| Pereira dos Santos, Pedro Tenório and Fabio Sampaio | 542-550 |
| The <i>in silico</i> prediction of bacterial essential genes | |
| I.A.M. Cooper and M.L. Duffield | 551-559 |

| Bioinformatics for the identification of antimicrobial targets in oral bacteria Andréa Cristina Barbosa da Silva and Thaís Gaudêncio do Rêgo | 560-569 |
|---|---------|
| A formulation of olive oils (oHo®) shows potent antimicrobial activities <i>in vitro</i> and in patients with atopic dermatitis (AD) colonized by <i>S. aureus</i> . Other clinical results in AD and atopy | |
| V.G. Villarrubia, V. Pérez-Bañasco, J.M. Gil-Cunquero, F. Borrego-Utiel, R. Cisterna- Cáncer and S. Vidal-Asensi | 570-577 |
| Methods that discriminate immobilized from eluting mechanisms of kill John-Bruce D. Green | 578-585 |
| Quorum quenching – an alternative antimicrobial therapeutics | |
| Sunita Adak, Lakshmishri Upadrasta, S.P. Jeevan Kumar, Rahul Soni and Rintu Banerjee | 586-593 |
| Cytokines, key players to turn on/off the anti- <i>Trypanosoma cruzi</i> innate defense mechanisms | |
| Eugenio A. Carrera-Silva, Susana Gea and Natalia Guiñazú | 594-604 |
| Role of exogenous chemokines as immunotherapeutic tool against visceral leishmaniasis. | |
| G. Gupta, R. Dey, S. Bhattacharyya and S. Majumdar | 605-612 |
| Phagocyte and extra-phagocyte myeloperoxidase-mediated microbicidal action Robert C. Allen and Jackson T. Stephens, Jr. | 613-621 |
| New platforms for the diagnosis and identification of fungal and bacterial pathogens G. Gelsomino, R. Faedda, C. Rizza, G. Petrone and S.O. Cacciola | 622-630 |
| Antimicrobials: old tools, new approaches Melo, Geraldo Batista de and Moreira, Michel Rodrigues | 631-635 |
| Use of the xenobiotic extrusion pump, MexAB-OprM, of <i>Pseudomonas aeruginosa</i> as a reporter to construct a high throughput screening system for the development of novel antimicrobials | |
| H. Yoneyama, K. Akiba, T. Ando and E. Isogai | 636-643 |
| Strategies for Vaccination and Control of Apicomplexan Protozoan Parasites Michael Wallach | 644-649 |
| Advance photodynamic inactivation of dental pathogenic microorganisms with water-soluble and cationic phthalocyanines | |
| Vanya Mantareva, Veselin Kussovski, Ivan Angelov and Slavcho Dimitrov | 650-661 |

| Effectiveness of photodynamic therapy on Gram-negative bacteria Wanessa C. M. A Melo, Lucas F. Castro, Roberta M. M. T. S. Dal'Mas and Janice R. Perussi. | 662-667 |
|--|---------|
| Science against microbial pathogens: photodynamic therapy approaches Constance L.L. Saw | 668-674 |
| An Introduction to Photoantimicrobials: Photodynamic Therapy as a Novel Method of Microbial Pathogen Eradication Tyler G. St. Denis and Michael R. Hamblin | 675-683 |
| New techniques in antimicrobial photodynamic therapy: scope of application and overcoming drug resistance in nosocomial infections Faina Nakonechny, Marina Nisnevitch, Yeshayahu Nitzan and Michael A. Firer | 684-691 |

VOL. 2

| Introduction | XVII |
|--------------|------|
|--------------|------|

Antimicrobial Resistance

| Bacterial iron uptake: a promising solution against multidrug resistant bacteria S. Fardeau, C. Mullié, A. Dassonville-Klimpt, N. Audic, A. Sasaki and P. Sonnet | 695-705 |
|--|---------|
| Possibility of novel therapeutic strategy for multidrug resistant <i>Pseudomonas</i> <i>aeruginosa</i> using bactericidal activity in <i>Streptococcus sanguinis</i> secretion Masachika Senba and Kiwao Watanabe | 706-713 |
| Antimicrobial resistance in <i>Staphylococcus</i> spp. M. L. Ribeiro de Souza da Cunha and D. R. Ustulin | 714-721 |
| Mini Review: Novel antimicrobial compounds in the age of increasing bacterial resistance W.O. Chung, J.C. Watah and D.T. Hobbs | 722-727 |
| Induction of systemic resistance to anthracnose in cucumber by natural components of <i>Allium</i> vegetables and shiitake mushrooms H. Inagaki, A. Yamaguchi, K. Kato, C. Kageyama and H. Iyozumi | 728-735 |

| Antimicrobial resistance in biofilms M.G. Paraje | 736-744 |
|---|---------|
| <i>Helicobacter pylori</i> resistance to antibiotics Filipa F. Vale, Mónica Roxo Rosa and Mónica Oleastro | 745-756 |
| Altered Ergosterol biosynthetic pathway - an alternate multidrug resistance mechanism independent of drug efflux pump in human pathogenic fungi <i>C. albicans</i> Tulika Prasad, Sunesh Sethumadhavan and Zeeshan Fatima | 757-768 |
| Sensing the host niche: pH as a novel determinant of multidrug resistance Saif Hameed | 769-772 |
| Chinese medicinal herbs against antibiotic-resistant bacterial pathogens Ben Chung-Lap Chan, Clara Bik-San Lau, Claude Jolivalt, Sau-Lai Lui, Carine Ganem- Elbaz, Jean-Marc Paris, Marc Litaudon, Kwok-Pui Fung, Ping-Chung Leung and Margaret Ip | 773-781 |
| Treatment of methicillin-resistant <i>Staphylococcus aureus</i> otorrhea Chul Ho Jang, Yong Bum Cho, Cheol Hee Choi and Hun Cho | 782-785 |
| Antibiotic resistance traits of facultative <i>Enterobacter cloacae</i> strain studied with the PMEU (Portable Microbe Enrichment Unit) Elias Hakalehto | 786-796 |

Biofilms

| SOS-inducible biofilms Tao Weitao | 799-812 |
|---|---------|
| Opportunistic pathogens and their biofilm "Food for thought" Amro A. Amara | 813-825 |
| Antimicrobial resistance to disinfectants in biofilms P.Araújo, M.Lemos, F.Mergulhão, L. Melo and M.Simões | 826-834 |
| Catheters: a suitable surface for biofilm formation J. Treter and A. J. Macedo | 835-842 |
| Strategies to control <i>Staphylococcus epidermidis</i> biofilms F. Gomes, B. Leite, P. Teixeira and R. Oliveira | 843-852 |

| Bacteriocin activity and resistance in livestock pathogens H. C. Mantovani, A. M. O. Cruz and A. D. Paiva | 853-863 |
|--|---------|
| A rapid, high-throughput method for culturing, characterizing and biocide efficacy testing of both planktonic cells and biofilms N.D. Allan, A. Omar, M.W. Harding and M.E. Olson | 864-871 |
| A multi-well plate method for rapid growth, characterization and biocide sensitivity testing of microbial biofilms on various surface materials M.W. Harding, R.J. Howard, G.D. Daniels, S.L. Mobbs, S.L.I. Lisowski, N.D. Allan, A. Omar and M.E. Olson. | 872-877 |
| Nanotechnology applied to medical biofilms control C. Sousa, C. Botelho and R. Oliveira | 878-888 |
| <i>In-situ</i> study of early stages of biofilm formation under different environmental stresses by ATR-FTIR spectroscopy F. Humbert and F. Quilès | 889-895 |
| Biofilm formation, control and novel strategies for eradication Maria Esperanza Cortés, Jessika Consuegra Bonilla and Ruben Dario Sinisterra | 896-905 |
| Mechanisms and experimental models for the assessment of microbial biofilms' phenotypical resistance /tolerance V.Lazar and M.C. Chifiriuc | 906-911 |

Antimicrobial Peptides

| LL37, a human antimicrobial peptide with immunomodulatory properties Reinaldo Ramos, Lucília Domingues and Miguel Gama | 915-925 |
|--|---------|
| Antibacterial Peptides: A Review Christine Cézard, Viviane Silva-Pires, Catherine Mullié and Pascal Sonnet | 926-937 |
| Antimicrobial peptides modulate bilayer barrier properties using a variety of mechanisms of actions Md. Ashrafuzzaman | 938-950 |
| Structural and functional insights into plant bactericidal peptides E.S. Cândido, W.F. Porto, D.S. Amaro, J.C. Viana, S.C. Dias and O.L. Franco | 951-960 |

| Novel strategy for designing antimicrobial peptides: an answer to the development of drug resistance. | |
|---|-----------|
| N. B. Iannucci, R. González, O. Cascone and F. Albericio | 961-967 |
| Isolation of a New Antimicrobial/Antitumor Plant Peptide: Biotechnology Prospects for its Use in Cancer and Infectious Diseases Therapies | |
| María G. Guevara, Fernando F. Muñoz, María B. Fernández, Julieta R. Mendieta and Gustavo R. Daleo | 968-976 |
| Gram-positive antibiotic biosynthetic clusters: a review A. Argüelles Arias, M. Craig and P. Fickers | 977-986 |
| Antimicrobial Peptides of Probiotic <i>Lactobacillus</i> strains S. Pithva, P. Ambalam, J. M. Dave and B.R.M. Vyas | 987-991 |
| Production of eukaryotic antimicrobial peptides by bacteria – A review Rogier A Gaiser, Luis Rivas and Paloma López | 992-1002 |
| A preliminary study on antimicrobial peptides in the naturally damaged tunic of <i>Ciona intestinalis</i> (Tunicata) | |
| M. A. Di Bella, H. Fedders, M. Leippe and G. De Leo | 1003-1007 |

Natural products and biocontrol

| Antimicrobial natural products | |
|---|-----------|
| Kenneth G. Ngwoke, Damian C. Odimegwu and Charles O. Esimone | 1011-1026 |
| The potential anticariogenic effect of coffee A.G. Antonio, A. Farah, K.R.N. dos Santos and L.C. Maia | 1027-1032 |
| Control of plant diseases using extracts from medicinal plants and fungi | |
| J. R. Stangarlin, O. J. Kuhn, L. Assi and K. R. F. Schwan-Estrada | 1033-1042 |
| Pathogenesis Related (PR) Proteins in Plant Defense Mechanism Saboki Ebrahim, K.Usha and Bhupinder Singh | 1043-1054 |
| Antifungal plant extracts Marcel Pârvu and Alina E. Pârvu | 1055-1062 |
| Homeopathy for the control of plant pathogens M. V.Toledo, J. R. Stangarlin and C. M. Bonato | 1063-1067 |

| Structural and genetic alterations of fungal cells caused by mexican propolis ML. Quintero Mora, A. Londoño Orozco, CI. Soto Zárate, CG. García Tovar, L. Carrillo Miranda, JG. Penieres Carrillo and TA. Cruz Sánchez | 1068-1073 |
|--|-----------|
| Small cysteine-rich proteins from plants: a rich resource of antimicrobial agents Mrinal Bhave and Dinesh Raj Methuku | 1074-1083 |
| Plant antimicrobials in food applications: Minireview Yasmina Sultanbawa | 1084-1093 |
| Olive leaf extract and usage for development of antimicrobial food packaging Z.Ö. Erdohan and K.N. Turhan | 1094-1101 |
| Antimicrobial compounds produced by <i>Bacillus</i> spp. and applications in food F. Baruzzi, L. Quintieri, M. Morea and L. Caputo | 1102-1111 |
| Use of natural antimicrobials for the control of <i>Listeria monocytogenes</i> in foods C. A. Campos, M.P. Castro, M.F. Gliemmo and L.I. Schelegueda | 1112-1123 |
| Antimicrobial volatile essential oils in edible films for food safety Wen-Xian Du, Roberto J. Avena-Bustillos, Sui Sheng T. Hua and Tara H. McHugh | 1124-1134 |
| Essential oils against yeast and moulds causing food spoilage Judit Krisch, Tserennadmid Rentsenkhand and Csaba Vágvölgyi | 1135-1142 |
| The mode of antibacterial action of essential oils M. L. Faleiro | 1143-1156 |
| Shelf life prolongation of fruit juices through essential oils and homogenization: a review A. Bevilacqua, M.R. Corbo, D. Campaniello, D. D'Amato, M. Gallo, B. Speranza and M. Sinigaglia | 1157-1166 |
| Antidermatophytic activity of essential oils M. Zuzarte, M. J. Gonçalves, J. Canhoto and L. Salgueiro | 1167-1178 |
| Antimicrobial activity of plant natural extracts and essential oils M.A. Calvo, E.L. Arosemena, C. Shiva and C. Adelantado | 1179-1185 |
| Effects of selected plants on the survival of <i>Staphylococcus aureus</i> I. Steinka and A. Kukulowicz | 1186-1194 |
| Plant derived antifungals- trends and potential applications in veterinary medicine: A mini-review Nopamart Trakranrungsie | 1195-1204 |
| Antimicrobial potentials of <i>Allium roseum</i> : Recent Advances and Trends Najjaa Hanen, Sami Fattouch, Emna Ammar and Mohamed Neffati | 1205-1210 |

| Evaluation of <i>in vitro</i> and <i>in vivo</i> antibacterial and antifungal activity of "camelyn | |
|--|-----------|
| m" Benedikte Maglakelidze, Guguli Abashidze, Inga Dadeshidze, Vakxtang Mshvildadze, Andre Pichete, Vincent Perreten, Shota Tsanava, Nata Shubladze and Koba Gurielidze | 1211-1215 |
| GM flax as a source of effective antimicrobial compounds M. Czemplik, M.Żuk, A. Kulma, S. Kuc and J. Szopa | 1216-1224 |
| Antimicrobial properties of resveratrol: a review L. Paulo, M. Oleastro, Eugenia Gallardo, J. A. Queiroz and F. Domingues | 1225-1235 |
| Extending the benefits of antifungal proteins from plants David W.M. Leung and Hossein Alizadeh | 1236-1243 |
| Molecular defence responses of sugarcane (Saccharum officinarum L.) to smut | |
| (Sporisorium scitamineum (Syd.) Piepenbr & Oberw. 2002. | |
| M.E. Legaz, R. Santiago, R. de Armas, B. Alarcón, E.M. Díaz, E. Sánchez-Elordi, M. Sacristán, B. Fontaniella, A. M. Millanes, M. Blanch and C. Vicente | 1244-1250 |
| Indian nutraceutical plant leaves as a potential source of natural antimicrobial | |
| agents Sumitra Chanda and Mital Kaneria | 1251-1259 |
| Endophytic fungi from brazilian mangrove plant <i>Laguncularia racemosa</i> (L.) Gaertn. (Combretaceae): their antimicrobial potential M.R.O. Silva, A.C. Almeida, F.V.F. Arruda and N. Gusmão | 1260-1266 |
| Antimicrobial activity of aqueous and methanolic extracts from <i>Arthrospira maxima</i> N. B. Medina-Jaritz, D. R. Perez-Solis, S. L. Ruiloba de Leon F. and R. Olvera-Ramírez | 1267-1271 |
| Antimicrobial activities of microalgae: an invited review Helena M. Amaro, A. Catarina Guedes and F. Xavier Malcata | 1272-1284 |
| Marine microorganisms: the world also changes Pilar González-Párraga, Alberto Cuesta, J. Meseguer and M ^a Ángeles Esteban | 1285-1292 |
| Marine Compounds and their Antimicrobial Activities M. J. Abad, L. M. Bedoya and P. Bermejo | 1293-1306 |
| Influence of temperature on the production of antibiotic molecules in <i>Bacillus amyloliquefaciens strain HNA3</i> R.A. Nastro, Di Costanzo A., Gesuele R., Trifuoggi M., Inglese M. and Guida M | 1307-1310 |
| Bacteriocin producing lactic acid bacteria isolated from Boza, a traditional fermented beverage from Balkan Peninsula – from isolation to application Jean Guy LeBlanc and Svetoslav Dimitrov Todorov | 1311-1320 |

| Growth Inhibition Strategies Based on Antimicrobial Microbes/Toxins Shanow Uthman, Eyemen Kheir, Christian Bär, Daniel Jablonowski and Raffael Schaffrath | 1321-1329 |
|--|-----------|
| Enterocins: Bacteriocins with applications in the food industry Y.M. Alvarez-Cisneros, T.R. Sáinz Espuñes, C.Wacher, F.J.Fernandez and E.Ponce-Alquicira. | 1330-1341 |
| Importance of microbial antagonisms about food attribution Zerrin Erginkaya, Emel Ünal and Selin Kalkan | 1342-1348 |