IMMUNE REGULATION IN PLASMA CELL MYELOMA

Esther Aklilu

Submitted for the degree of Master of Science at the University of Technology, Sydney in December 2013

CERTIFICATE OF ORIGINAL AUTHORSHIP

I certify that the work in this thesis has not previously been submitted for a degree nor has it been submitted as part of requirements for a degree except as fully acknowledged within the text.

I also certify that the thesis has been written by me. Any help that I have received in my research work and the preparation of the thesis itself has been acknowledged. In addition, I certify that all information sources and literature used are indicated in the thesis.

Signature of Student:

Date:

Acknowledgements

I wish to express my sincere gratitude to my supervisors Dr Ross Brown, Mrs Narelle Woodland and Dr Najah Nassif for their magnanimous support, encouragement and patience throughout the duration of my degree.

To Ross, thank you for the opportunity to work for you and for allowing me to take on such an interesting project for my degree. I truly appreciate your exceptional advice and guidance throughout this project.

To Narelle and Najah, thank you for accepting me as your Masters Degree student and for all the excellent advice, support and encouragement. I truly appreciate all your immense help.

To my fellow research team and colleagues, your assistance and support is greatly appreciated. Thank you to Shihong Yang for imparting to me your knowledge of flow cytometry, cell sorting and cell culture work. It has been an absolute joy working with you. Thank you to James Favaloro for your editorial advice, support and friendship. Although we had only worked together for a short time, it has been a real pleasure working with you. Also, I would like to appreciate your assistance in the analysis of the 10 year survivor samples.

Thank you to the haematology staff of RPAH, for making my time in RPA thoroughly enjoyable and to the volunteers, for the kind donation of your blood.

Thank you to Professor Doug Joshua and Dr P Joy Ho, for your suggestions and advice during our weekly meetings.

I also wish to acknowledge the financial support provided by the Sydney Foundation for Medical Research and the Cancer Institute, NSW.

Finally, I wish to thank my family and friends for their support and encouragement.

Manuscripts and presentations arising from the work completed as part of this thesis

Articles published in peer-reviewed journals

Favaloro J, Brown R, Aklilu E, Yang S, Suen H, Hart D, Fromm P, Gibson J, Khoo L, Ho PJ, Joshua D (2013) Myeloma skews regulatory T and pro-inflammatory T helper 17 cell balance in favour of a suppressive state. *Leukaemia & lymphoma*. *55*(*5*):1090-8

Bryant C, Suen H, Brown R, Yang S, Favaloro J, Aklilu E, Gibson J, Ho PJ, Iland H, Fromm P, Woodland N, Nassif N, Hart D, Joshua D (2013) Long-term survival in multiple myeloma is associated with a distinct immunological profile, which includes proliferative cytotoxic T-cell clones and a favourable Treg/Th17 cell balance. *Blood Cancer journal 3:e148*.

Conference abstracts

Favaloro J, Brown R, Aklilu E, Yang S, Suen H, Gibson J, Ho PJ, Joshua D. (2013) The ratio of Treg/Th17 cells is of prognostic significance in multiple myeloma. Australian Flow Cytometry group (AFCG) meeting.

Bryant CE, Brown RD, Yang S, Suen H, Aklilu E, Favaloro J, Hart DNJ, Fromm P, Woodland N, Nassif N, Iland H, Gibson J, Ho PJ, Joshua DE. (2011) Immunological Biomarkers in 10 year Survivors of Multiple Myeloma. *Proceedings of the Haematology Society of Australia and New Zealand*. O/021.

Bryant CE, Brown RD, Yang S, Suen H, Aklilu E, Favaloro J, Hart DNJ, Fromm P, Woodland N, Nassif N, Iland H, Gibson J, Ho PJ, Joshua DE. (2011) Ten year survivors of multiple myeloma demonstrate a differential expression of immunological biomarkers including a high incidence of cytotoxic T-cell clones which have not acquired myeloma-associated anergy. *Blood.* 118:S38865.

Aklilu E, Brown R, Yang S, Kabani K, Woodland N, Nassif N, Ho P, Gibson J, Joshua D. (2010) Treg number, Treg function and Th17 cells in plasma cell dyscrasias. *Proceedings of the Haematology Society of Australia and New Zealand*. *A94*.

Aklilu E, Brown R, Yang S, Kabani K, Woodland N, Nassif N, Ho PJ, Gibson J, Joshua D (2010) Treg number, Treg function and Th17 cells in plasma cell dyscrasias. Australian Flow Cytometry group (AFCG) meeting. A87.

Aklilu E, Brown R, Yang S, Kabani K, Woodland N, Nassif N, Ho PJ, Gibson J, Joshua D (2010) Treg number, Treg function and Th17 cells in plasma cell dyscrasias, The XXVIIth Annual scientific research meeting. P42.

Abbreviations

Ab/ Abs	Antibody (-ies)
AF-647	Alexa Fluor 647
AML	Acute myeloid leukaemia
Anti-	Antibody against
APC	Allophycocyanin
APCs	Antigen presenting cells
ATLL	Adult T-cell leukaemia/lymphoma
B cell	B lymphocyte
β ₂ M	Beta-2 microglobulin
BM	Bone marrow
ВММС	Bone marrow mononuclear cells
Bort-	Bortezomib (Velcade)
°C	Degrees Celsius
CD	Cluster of differentiation
CFSE	Carboxyfluorescein succinimidyl ester
CLL	Chronic lymphocytic leukaemia
CLMF	Cytotoxic lymphocyte maturation factor
CRAB	Hypercalcaemia, renal failure, anaemia and bone lesion
CRP	C-reactive protein
CTLA-4	Cytotoxic T lymphocyte-associated antigen-4
CTLA-8	Cytotoxic T lymphocyte-associated antigen-8
DC	Dendritic cells
Dex	Dexamethasone
DMSO	Dimethyl sulfoxide
DVT	Deep vein thrombosis
EDTA	Ethylenediamine tetra-acetic acid
FACS	Fluorescence activated cell sorting
FISH	Fluorescent in situ hybridisation
FITC	Fluorescein isothiocyanate

FoxP3	Forkhead box P3
FSC	Forward scatter
g	Gravitational force
G-CSF	Granulocyte-colony stimulating factor
GITR	Glucocorticoid-induced tumour necrosis factor -receptor-related
GvHD	Graft versus host disease
HCI	Hydrochloric acid
HEPES	4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid
hi	High
HLA	Human leukocyte antigen
IFN	Interferon
lg	Immunoglobulin
IL	Interleukin
IMiD/ IMiDs	Immunomodulatory drug (s)
iTreg	Induced regulatory T cell
ISS	International staging system
kDa-	Kilo Dalton
LAP	Latency associated peptide
Len	Lenalidomide
Len + Dex	Lenalidomide in combination with dexamethasone
mAB	Monoclonal Antibody
MFI	Mean fluorescence intensity
MGUS	Monoclonal gammopathy of undetermined significance
МНС	Major histocompatibility complex
min	Minutes
MM	Plasma cell myeloma
MRI	Magnetic resonance imaging
NK	Natural killer
NKSF	Natural killer cell stimulatory factor
NS	Not significant (P>0.05)
nTreg	Natural regulatory T cell

РВ	Peripheral blood
РВМС	Peripheral blood mononuclear cells
PBS	Phosphate buffered saline
PCLI	Plasma cell labelling index
PE	Phycoerythrin
PerCP	Peridinin chlorophyll protein
PMA	Phorbol 12-myristate 13-acetate
Pom	Pomalidomide
Rh	Recombinant human
ROR	Retinoid-related orphan receptor
RPMI	Roswell Park Memorial Institute
RT	Room temperature
SAM	Significance analysis of microarray
SSC	Side scatter
SCT	Stem cell transplantation
SD	Standard deviation
SEM	Standard error of mean
SM	Smouldering myeloma
SPE	Serum protein electrophoresis
STAT3	Signal transducer and activator of transcription 3
T cell	T lymphocyte
TCR	T cell receptor
TGF-β	Transforming growth factor β
Th	T helper cell
Thal-	Thalidomide
TILs	Tumour infiltrating lymphocytes
TNF-α	Tumour necrosis factor -α
Treg	Regulatory T lymphocyte
WM	Waldenström macroglobulinaemia

Abstract

The pathogenesis of monoclonal gammopathies, particularly plasma cell myeloma (MM), is multifaceted and complex. In recent years, Treg and Th17 cells have emerged as key factors in the development and progression of malignancies including MM. However, there are still conflicting reports on whether regulatory T (Treg) cells and Th17 cells are increased or decreased in the peripheral blood (PB) of patients with MM. This is partly due to technical difficulties associated with the use of the transcriptional repressor, forkhead box P3 (FoxP3) to identify Treg cells. Studies have shown FoxP3 results to be dependent on the clones, fluorochromes attached and fixation/permeabilisation methods used. More recent studies have defined Treg cells as CD4⁺CD25^{hi} cells which do not express CD127, an IL-7 receptor. This methodology was used to determine Treg cell number and develop assays for the assessment of Treg cell function. The study also extends to exploring Th17 cell equilibrium on the survival of patients with MM.

CD4⁺CD25^{hi}CD127⁻ expression was used to quantitate Treg cell numbers and an intracellular IL-17 assay on CD3⁺CD4⁺ cells was used for Th17 cell enumeration. Treg cell function was determined using carboxyfluorescein succinimidyl ester (CFSE) tracking of Treg depleted lymphocyte preparations stimulated by anti-CD3,CD2,CD28 beads at a 1:1 ratio for 4 days \pm 1:1 fluorescence-activated cell sorted Treg cells. This functional assay was also used to investigate the effect of recombinant human (rh) TGF- β and rhIL-12 on Treg cells.

The mean proportion of Treg cells in the CD4⁺ compartment of PB of patients with MM (n=32) was 8.9±0.6% and this was increased compared to the mean of the normal cohort (n=36) at 6.5±0.4% (p=0.009). However, no significant difference was observed between the frequency of PB Treg cells in the control group compared to patients with monoclonal gammopathy of undetermined significance (MGUS) (n=20) (mean=7.5±0.8%; P=0.24) and patients with Waldenström macroglobulinaemia (WM) (n=13) (mean=6.0±0.5%; P=0.48). Interestingly, a comparison of the absolute numbers exhibited different results. A significantly lower number of Treg cells was observed in patients with MM [(3.2 ± 0.4) x10⁷/L; P<0.01] and WM [(3.0 ± 0.6) x10⁷/L; P<0.01] compared to the control group [(6.4 ± 0.7) x10⁷/L]. However, no significant difference was observed when comparing patients with MGUS [(4.3 ± 0.8) x10⁷/L; P=0.06] to the normal cohort. It was observed that a significantly

higher proportion of PB Treg cells in patients with MM (85.9±1.8%; P<0.01) and WM (86.4±2.1%; P=0.02) to be of the CD45RO⁺ memory phenotype compared to the normal cohort (76.7±2.5%). However this was not observed in patients with MGUS (73.4±4.0%; P=0.47). In addition, the study revealed that PB Treg cell proportions were not influenced by MM stage. Thalidomide treated patients with MM appeared to have an increased PB Treg cell proportion, however only a small number of thalidomide treated patients were tested due to the use of thalidomide therapy being phased out and its replacement with lenalidomide. Treg cells from bone marrow (BM) were compared to matched PB samples from patients with MM, demonstrating a significantly greater proportion (p=0.02) of Treg cells in the CD4⁺ compartment of the BM (9.7±1.2%) compared to PB (6.7±1.4%).

Regarding Th17 cells, a significant decrease (p=0.03) in the mean proportion and absolute number of Th17 cells was observed in the PB of patients with MM (n=22) compared with the controls (n=20) ($0.7\pm0.1\%$ and $2.0\pm0.6\%$ respectively). However, the mean number of Th17 cells in patients with MGUS ($2.2\pm0.6\%$; n=15) and WM ($1.1\pm0.2\%$; n=12) was not significantly different from normal. No correlation was observed between Th17 cell number and MM staging or therapy.

Additionally, the study explored the Treg/Th17 cell ratio in PB of patients with monoclonal gammopathies with comparison made to normal subjects. The mean Treg/Th17 cell ratio of patients with MM (16.1±2.4) was significantly higher (p=0.0002) than the healthy control group (6.6±1.0). The Treg/Th17 cell ratio of WM and MGUS patients was 7.0±1.0 and 4.9±0.5 respectively, neither of which were statistically different to the ratio of the normal controls. Most interestingly, patients who have survived with MM for 10 or more years possessed a Treg/Th17 cell ratio similar to the normal controls (7.04±2.47; p=0.84) and this was shown to affect overall survival. The data demonstrated that patients with MM observed to have a high Treg/Th17 cell ratio had an overall shorter survival compared to those whose Treg/Th17 cell ratio was lower (p<0.025).

The suppressive capability of Treg cells from MM patients (n=15) was variable. The Treg cell function of patients treated with lenalidomide (n=5) was increased (mean=68%) compared to patients treated with thalidomide (n=5; mean=23%), Velcade (n=3; mean=12%), untreated patients (n=5; mean=36%) and normal controls (n=11; mean=31%). The suppression exerted upon the CD4⁺ T cell subset in patients treated with bortezomib was

significantly lower when compared to the normal cohort. However, no significant difference in CD8⁺ T cell suppression was found between patients with MM and the normal controls. rhTGF- β increased the suppressive capabilities and rhIL-12 reduced the function of Treg cells from both MM and normal PB samples.

In conclusion, immune regulation is dysfunctional in patients with MM as the proportion of PB Treg cells is increased and Th17 cells are reduced. Also, the cytokine microenvironment and treatment have a major impact on the function of Treg cells. The data clearly delineate the importance of the PB Treg/Th17 cell equilibrium, revealing a strong association between the Treg/Th17 cell homeostatic balance and disease progression and survival in MM, indicating an imbalance may cause either or both the innate and adaptive immune system to be dormant and incapacitate the anti-tumour response.

Table of Contents

ACKNOWLE	DGEMENTS II		
Manuscrii	MANUSCRIPTS AND PRESENTATIONS ARISING FROM THE WORK COMPLETED AS PART OF THIS THESIS		
ABBREVIAT	ABBREVIATIONS IV		
ABSTRACT.			
TABLE OF C	ONTENTSX		
LIST OF FIG	URES XV		
LIST OF TAB	LESXVII		
Introducti	ON1		
Plasma ce	LL MYELOMA2		
1.1 Int	RODUCTION TO PLASMA CELL MYELOMA: EPIDEMIOLOGY, INCIDENCE AND MORTALITY2		
1.2 CLI	NICAL PRESENTATION, DIAGNOSIS AND PROGNOSTIC INDICATORS IN MM2		
1.2.1	Clinical presentation2		
1.2.2	Diagnosis and classification of monoclonal gammopathies		
1.2.3	Staging and prognosis5		
1.3 Тн	ERAPY FOR MM		
1.3.1	Thalidomide8		
1.3.2	Lenalidomide and Pomalidomide9		
1.3.3	Bortezomib10		
1.3.4	Haematopoietic stem cell transplantation11		
1.3.4.1	Autologous haematopoietic stem cell transplantation11		
1.3.4.2	Allogeneic haematopoietic stem cell transplantation12		
IMMUNE RE	GULATION		
1.4 Тн	E IMMUNE SYSTEM		
1.4.1	Lymphocytes12		
1.5 Rec	GULATORY T (TREG) CELLS15		
1.5.1	Treg cells in PB15		
1.5.2	Classes of Treg cells15		
1.5.3	Treg cell identification by flow cytometry16		

1.	5.4	Treg cells and cancer	17
1.	5.5	Treg cells and haematological malignancies	18
1.	5.6	Treg cells and GvHD	18
1.	5.7	Controversy	19
1.6	Тн1	.7 CELLS AND THEIR CYTOKINES	21
1.	6.1	Th17 cell development and differentiation	21
1.	6.2	The IL-17 cytokine	21
1.	6.3	Th17 cells in cancer	22
1.7	Тне	ROLE OF THE CYTOKINES TGF-B AND IL-12 IN CANCER	23
1.	7.1	TGF-β	23
1.	7.2	IL-12	23
1.8	BAG	KGROUND TO THE PROJECT	24
1.9	HYF	OTHESIS	24
1.10) Air	IS OF THE PROJECT	25
ΜΑΤΕ	RIALS	AND METHODS	26
Мате 2.1		AND METHODS	
	Paf		27
2.1 2.2	Paf	TICIPANTS	27 27
2.1 2.2 2.	Paf San	ITICIPANTS IPLE COLLECTIONS Blood sample collection	27 27
2.1 2.2 2.	Paf San 2.1 2.2	ITICIPANTS IPLE COLLECTIONS Blood sample collection	27 27 27 27
2.1 2.2 2. 2. 2.3	Paf San 2.1 2.2	TICIPANTS IPLE COLLECTIONS Blood sample collection BM collection	27 27 27 27 27 28
2.1 2.2 2. 2. 2.3 2.3	Paf San 2.1 2.2 Me	TICIPANTS IPLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent	27 27 27 27 27 28 28
2.1 2.2 2. 2.3 2.3 2.	Par San 2.1 2.2 Me 3.1	APLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent Preparation of phosphate buffered saline	27 27 27 27 27 28 28 28
2.1 2.2 2. 2.3 2.3 2. 2. 2.	Par San 2.1 2.2 Me 3.1 3.2	APLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent Preparation of phosphate buffered saline Preparation of Roswell Park Memorial Institute -10 medium	27 27 27 27 28 28 28 28 28
2.1 2.2 2. 2.3 2.3 2. 2. 2. 2.	Par San 2.1 2.2 Me 3.1 3.2 3.3 3.4	ITICIPANTS IPLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent Preparation of phosphate buffered saline Preparation of phosphate buffered saline Preparation of Roswell Park Memorial Institute -10 medium Preparation of RPMI cell culture medium	27 27 27 27 28 28 28 28 28 28 28
2.1 2.2 2. 2.3 2.3 2. 2. 2. 2. 2. 2.	PAF SAN 2.1 2.2 ME 3.1 3.2 3.3 3.4 3.5	TICIPANTS MPLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent Preparation of phosphate buffered saline Preparation of phosphate buffered saline Preparation of Roswell Park Memorial Institute -10 medium Preparation of RPMI cell culture medium Preparation of MACSBeads	27 27 27 27 28 28 28 28 28 28 29 29
2.1 2.2 2. 2.3 2.3 2. 2. 2. 2. 2. 2.	PAF SAM 2.1 2.2 ME 3.1 3.2 3.3 3.4 3.5 3.6	TICIPANTS APLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent Preparation of phosphate buffered saline Preparation of Roswell Park Memorial Institute -10 medium Preparation of RPMI cell culture medium Preparation of MACSBeads Preparation of carboxyfluorescein succinimidyl ester stain	27 27 27 27 28 28 28 28 28 29 29 29 29
 2.1 2.2 2. 2.3 2. 2. 2. 2. 2. 2. 2. 2.4 	PAF SAN 2.1 2.2 ME 3.1 3.2 3.3 3.4 3.5 3.6 GEN	APLE COLLECTIONS MPLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent Preparation of ammonium chloride lysing reagent Preparation of phosphate buffered saline Preparation of phosphate buffered saline Preparation of Roswell Park Memorial Institute -10 medium Preparation of RPMI cell culture medium Preparation of MACSBeads Preparation of carboxyfluorescein succinimidyl ester stain NERAL TECHNIQUES	27 27 27 27 28 28 28 28 28 28 29 29 29 29 31
 2.1 2.2 2. 2.3 2. 2. 2. 2. 2. 2. 2.4 2. 	PAF SAN 2.1 2.2 ME 3.1 3.2 3.3 3.4 3.5 3.6 GEN 4.1	ITICIPANTS IPLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent Preparation of phosphate buffered saline Preparation of phosphate buffered saline Preparation of Roswell Park Memorial Institute -10 medium Preparation of RPMI cell culture medium Preparation of MACSBeads Preparation of carboxyfluorescein succinimidyl ester stain NERAL TECHNIQUES Cell wash	27 27 27 27 28 28 28 28 28 28 29 29 29 29 31
2.1 2.2 2. 2.3 2.3 2. 2. 2. 2. 2. 2.4 2.4 2.	PAF SAN 2.1 2.2 ME 3.1 3.2 3.3 3.4 3.5 3.6 GEN 4.1 4.2	APLE COLLECTIONS MPLE COLLECTIONS Blood sample collection BM collection THODOLOGY FOR PREPARATION OF COMMON REAGENTS Preparation of ammonium chloride lysing reagent Preparation of ammonium chloride lysing reagent Preparation of phosphate buffered saline Preparation of phosphate buffered saline Preparation of Roswell Park Memorial Institute -10 medium Preparation of RPMI cell culture medium Preparation of MACSBeads Preparation of carboxyfluorescein succinimidyl ester stain NERAL TECHNIQUES	27 27 27 27 28 28 28 28 28 29 29 29 29 29 29 31 31

2.5	M	ONOCLONAL ANTIBODIES USED DURING EXPERIMENTATION	33
2.6	As	SAY FOR IDENTIFICATION OF TREG CELLS	34
2	.6.1	Cell staining for flow cytometric identification of CD127 ^{lo/-} Treg cells	
2	.6.2	Preparation of PBMC Treg cell identification using FoxP3	34
2	.6.3	Preparation of BM Treg cells for flow cytometric analysis	35
2.7	M	ETHODOLOGY FOR THE IDENTIFICATION OF TH17 CELLS IN PB	36
2	.7.1	Preparation of Th17 cells for flow cytometric analysis	36
2.8	Tr	EG CELL FUNCTIONAL ASSAY USING CFSE	37
2	.8.1	Fluorescence-activated cell sorting of PBMC	37
2	.8.2	FACS purity test	38
2	.8.3	CFSE staining of target cells	38
2	.8.4	Cell count	39
2	.8.5	Preparation of the MACSbead suspension	39
2	.8.6	Cell culture of Treg and CFSE stained target cell	39
2	.8.7	Analysis of the CFSE assay after culture	41
2.9	Ev	ALUATION OF THE IMPACT OF CYTOKINES (IL-12 AND TGF- eta) and anti-TGF- eta on Treg	CELLS 41
2	.9.1	IL-12 preparation	41
2	.9.2	rhTGF-β1 preparation	42
2	.9.3	Anti TGF- β preparation	42
2	.9.4	Plating of the cell culture wells with addition of cytokines	42
2.1	0 Sт	ATISTICS AND ANALYSIS	44
Resu	LTS		45
3.1	٥v	/ERVIEW	46
3.2	Tr	EG CELLS IN MONOCLONAL GAMMOPATHIES	46
3	.2.1	Identification of Treg cells using CD127	46
3	.2.2	Identification of Treg cells using FoxP3	49
3	.2.3	Enumeration of Treg cells in PB	52
3	.2.4	Phenotypic analysis of Treg cells	55
3	.2.5	Effect of IMiD treatment on Treg cell proportions	58

3.2	2.6	MM staging and Treg cell quantitation	60
3.2	2.7	Enumeration of Treg cells in BM versus PB	60
3.2	2.8	Treg cells in patients with MM who have survived more than 10 years	63
3.3	Тн1	7 CELLS IN MM AND OTHER MONOCLONAL GAMMOPATHIES	66
3.3	3.1	Identification of Th17 cells in PB	66
3.3	3.2	Enumeration of Th17 cells in PB	68
3.3	3.3	Th17 cells and stage of MM	70
3.3	3.4	Th17 cells in MM patients who have survived more than 10 years	71
3.4	Tre	G AND TH17 CELL EQUILIBRIUM	73
3.4	4.1	Treg/Th17 cell ratio in monoclonal gammopathies	73
3.4	4.2	Effect of a high Treg/Th17 cell ratio on overall survival in MM	73
3.5	Tre	G CELL FUNCTIONAL ASSAY	77
3.5	5.1	Development and optimisation of Treg cell functional assay	77
3.5	5.2	Treg cell functional assay results	79
3.5	5.3	Suppression of T cell proliferation	79
3.6	Eff	ECT OF CYTOKINES ON TREG CELL FUNCTION	84
Discus	SION		86
4.1	Ovi	ERVIEW	87
4.2	Тне	ROLE OF TREG CELLS IN MONOCLONAL GAMMOPATHIES	88
4.2	2.1	The phenotype and identification of Treg cells	88
4.2	2.2	Enumeration and phenotype of Treg cells in monoclonal gammopathies	89
4.2	2.3	Naïve and memory Treg cells in monoclonal gammopathies	91
4.2	2.4	Effect of MM treatment on Treg cell proportion	91
4.2	2.5	Effect of MM stage on Treg cell proportion	92
4.2	2.6	Quantification of Treg cells in the PB and BM of MM patients	93
4.2			
	2.7	Treg cell function in MM	93
4.2	2.7 2.8	Treg cell function in MIM	
		с С	95

4.3.1	Th17 cell enumeration in MM and other monoclonal gammopath	ies97
4.3.2	Association of MM stage and Th17 cell proportion	99
4.4 T	REG AND TH17 EQUILIBRIUM	100
4.4.1	Treg/Th17 cell ratio in monoclonal gammopathies	100
4.4.2	Treg/Th17 cell ratio and survival	101
4.5 C	ONCLUSIONS AND FUTURE DIRECTIONS	102
4.5.1	Future directions	102
4.5.2	Conclusion	103

FERENCES

List of Figures

Figure 1.1: Diagnostic characteristics of MM4
Figure 1.2: Gene expression patterns and overall survival in the different molecular
subgroups of MM7
Figure 1.3: The chemical structure of the IMiDs thalidomide, lenalidomide and
pomalidomide10
Figure 1.4: Hierarchy of lymphocytes and T cell lineages involved in the innate and adaptive
immune response14
Figure 2.1: Flow cytometric analysis of a CFSE stained cell population
Figure 2.2: Whole blood before and after centrifugation with Ficoll-Paque
Figure 2.3: Cell culture plate set up for investigation of Treg cell function40
Figure 2.4: Cell culture plate set up to evaluate the effect of IL-12, TGF- β , and anti-TGF- β on
the function of Treg cells43
Figure 3.1: Isotype control for each fluorochrome used in the Treg cell assay47
Figure 3.2: Flow cytometry results from a representative normal subject showing the gating
strategy for identification of Treg cells in PB48
Figure 3.3: Flow cytometry results from a representative normal subject showing the gating
strategy for identification of PB Treg cells using FoxP350
Figure 3.4: Comparison of FoxP3 expression on sorted CD25 ^{hi} CD127 ^{lo/-} Treg cells compared
to the Treg depleted T cells51
Figure 3.5: Enumeration of PB Treg cells in patients with monoclonal gammopathies and
normal control individuals54
Figure 3.6: Flow cytometry results showing CD45RA and CD45RO expression on Treg cells in
PB of a representative normal subject56
Figure 3.7: Expression levels of CD45RA and CD45RO on Treg cells in PB of patients with
monoclonal gammopathies and normal individuals57
Figure 3.8: Effect of IMiD treatment on PB Treg cell proportions in patients with MM59
Figure 3.9: Comparison of the percentage of PB Treg cells and the different stages of MM.61
Figure 3.10: Comparison of Treg cell numbers in the BM and PB of patients with MM62
Figure 3.11: Relative Treg cell numbers in PB of normal subjects, patients with MM for less
than 10 years and MM patients who have survived for 10 or more years since diagnosis64

Figure 3.12: Absolute number of PB Treg cells in normal subjects, patients with MM for less
than 10 years and patients who have survived 10 or more years with MM65
Figure 3.13: Flow cytometric results from a PB sample of a representative normal subject
showing the gating strategy for the identification of Th17 cells67
Figure 3.14: Relative Th17 cells number in the PB of patients with monoclonal
gammopathies and normal controls69
Figure 3.15: Comparison of the percentage of Th17 cells of the $CD4^+$ compartment in PB and
different stages of MM70
Figure 3.16: Enumeration of Th17 cells in PB of patients with MM and normal subjects72
Figure 3.17: Treg/Th17 cell ratio in patients with monoclonal gammopathies and normal
individuals74
Figure 3.18: Treg/Th17 cell ratio in 10 year MM survivors75
Figure 3.19: The effect of Treg/Th17 cell ratio on overall survival of patients with MM76
Figure 3.20: Optimisation of MACSbead stimulation for the Treg functional assay
Figure 3.21: Flow cytometry gating strategy for FACS of Treg cells in PB
Figure 3.22: A representative flow cytometric analysis of CFSE stained CD3 ⁺ cells after 4 days
culture
Figure 3.23: Effect of treatment on Treg cells suppressive activity on lymphocytes target
cells
Figure 3.24: Effect of IL-12, TGF- β and anti-TGF- β cytokines on the suppressive function of
Treg cells

List of Tables

Table 1.1: International Staging System for MM
Table 1.2: Comparison of studies of Treg identification, enumeration and function in MM. 20
Table 2.1: List of monoclonal antibodies (mAb) used during experimentation
Table 2.2: Fluorescent antibodies used for the flow cytometric identification of Treg cells
using CD127
Table 2.3: Fluorescent antibodies used for the flow cytometric identification of Treg cells
using FoxP335
Table 2.4: Fluorescent antibodies used for the flow cytometric identification of BM Treg cells
Table 2.5: Fluorescent antibodies used to identify Th17 cells for analysis by flow cytometry
Table 2.6: Fluorescent antibodies used to identify Treg cells for flow cytometric sorting38
Table 2.7: Fluorescent antibodies used for analysis of the CFSE assay by flow cytometry41