

# **The Molecular Biology of Venom Genes from Death Adder and King Brown Snake**

**Kah-Leong Sung**

Submitted for the degree of Masters of Science at the University of Technology, Sydney 2009

## CERTIFICATE OF AUTHORSHIP/ORIGINALITY

I certify that the work in this thesis has not previously been submitted for a degree nor has it been submitted as part of requirements for a degree except as fully acknowledged within the text.

I also certify that the thesis has been written by me. Any help that I have received in my research work and the preparation of the thesis itself has been acknowledged. In addition, I certify that all information sources and literature used are indicated in the thesis.

Production Note:

Signature removed prior to publication.

---

Kah Leong SUNG



## **Acknowledgements**

I would like to thank my supervisor Asso. Professor Kevin Broady. Without his support, his sense of humour, his patience and his insights, this project would not have been successful. I could not have asked for a better supervisor. Thank you.

I would also like to thank my brother, Kah-Kui Sung for his support throughout this project. He has been selfless in offering his time and resources.

Finally, I would like to thank the members of the now disbanded immunobiology unit at UTS. In particular, Ms Joyce To, Mr Matt Padula, Mr Darren Jones, Ms Sakura Narasimhan and Ms Susan Lemke deserve special mention here. Their encouragement and positive feedback have helped me overcome the numerous problems encountered in this project. In addition, I would also like to thank Asso. Professor Graham Nicholson for his help, his support and kind words throughout this project.

## Table of Contents

	Pages
Table of Contents	iv
List of Figures and Tables	
Abstract	
Chapter 1 - Introduction	Pages
1.1 Uniquely Australian	1
1.2 The Death Adder	1
1.3 Death Adder Venom Components	2
1.4 The King Brown Snake	3
1.5 King Brown Snake Venom Components	4
1.6 Habitat destruction and loss of natural resources	4
1.7 Phospholipase A <sub>2</sub> enzymes	5
1.8 The PLA <sub>2</sub> reaction	5
1.9 Classification	6
1.9.1 Group I	7
1.9.2 Group II	8
1.9.3 Group III	8
1.9.4 Group IV	8
1.9.5 Subgroups of PLA <sub>2</sub> enzymes	8
1.10 The PLA <sub>2</sub> fold	9
1.11 Toxicity of PLA <sub>2</sub>	11
1.11.1 Neurotoxicity	12
1.11.2 Myotoxicity	13
1.11.3 Haemotoxicity	13
1.12 Toxicity and PLA <sub>2</sub> activity	13
1.13 The Venom PLA <sub>2</sub> genes	15
1.14 The Alpha Neurotoxins	16
1.15 Classification	17
1.16 Structure and Function of Three Finger Proteins	17
1.17 The $\alpha$ -neurotoxin genes	19

1.18	Accelerated Evolution	20
1.19	Aims and Summary	21
Chapter 2 -	Construction of complementary DNA libraries	23
2.0	Introduction	24
2.1	Preparation of DPEC water	24
2.2	Precautions taken in messenger RNA extraction	24
2.3	The Modified Chomczynski and Sacchi Method	24
2.4	The RNeasy Midi Kit method	26
2.5	Protocol for Agarose Gel Electrophoresis of RNA	26
2.6	Method for Purification of mRNA from total RNA	27
2.7	Complementary DNA Library construction protocol	28
2.8	Method for verification of proper gene insertions	35
2.9	Results for the messenger RNA extraction	32
2.10	Screening the library for Insertion of random genes	35
2.11	Conclusion	37
Chapter 3 -	Screening for neurotoxin genes in Death Adder cDNA library	38
3.1	Introduction to screening procedure	39
3.2	DNA Primers for Screening of PLA <sub>2</sub> genes	39
3.3	DNA Primers for $\alpha$ -neurotoxin genes	41
3.4	Screening Procedure	44
3.5	Results for the first screen	47
3.6	The Problem of Preferential Amplification in PCR	49
3.7	The Protocol for Optimisation	50
3.8	Elution of the PCR product	51
3.9	Ligation to pGEM-T vector	51
3.10	Precipitation and Transformation	52
3.11	Plasmid Amplication and Purification	52
3.12	How many colonies should be sampled?	53
3.13	Analysis	54

3.14	Results for optimum number of cycles	55
3.15	Sampling and Sequencing the PLA <sub>2</sub> genes	57
3.16	Categorizing Groups A, B and C	57
3.17	Comparison of the Long Chain PLA <sub>2</sub> to other PLA <sub>2</sub> genes	68
3.18	Comparison of the Short Chain PLA <sub>2</sub> to other PLA <sub>2</sub> genes	68
3.19	Translation into protein sequences and alignment	69
3.20	Short Chain amino acid sequences	73
3.21	Long Chain amino acid sequences	75
3.22	Sequencing results for the $\alpha$ -neurotoxin genes	80
Chapter 4 -	Screening for neurotoxin genes in King Brown cDNA library	81
4.1	Introduction	82
4.2	PLA <sub>2</sub> enzymes in Death Adder Crude Venom	82
4.3	Aim - Determining possible neurotoxin genes in Death Adder	87
4.4	Methodology	87
4.5	Results for Israel's PCR screening method	87
4.6	Results for PCR approach for PLA <sub>2</sub> genes	89
4.7	Results for Automated Sequencing	89
4.8	Results for Protein Analysis	91
4.9	Screening Results for $\alpha$ -neurotoxin genes	96
4.10	Conclusion	96
Chapter 5 -	Conclusions, Caveats and new directions.	97
5.1	The screening results from King Brown snake venom gland cDNA library	98
5.1.1	The validity of the putative proteins	100
5.2	Further work :- definitive proof of accelerated evolution by sequencing introns	101
5.3	The Death Adder conundrum	101
5.4	Intra-species Variation	101

5.5	Multiple venom genes or single copy	103
5.6	Further work	104
5.7	Proper Phylogenetic Study	104
5.8	Concluding Remarks - a cautionary tale	105
	Bibliography	



## List of Figures and Tables

Figure 1.1	The Catalytic Reaction of PLA <sub>2</sub>	6
Figure 1.2	Classification of PLA <sub>2</sub>	7
Figure 1.3	Three Dimensional Ribbon Representation Oof PLA <sub>2</sub>	10
Figure 1.4	The Synaptic Vesicle Cycle	12
Figure 1.5	Ribbon representation of $\alpha$ -neurotoxin	18
Figure 2.1	Quality of messenger RNA	34
Figure 2.2	Verification of gene insertions into vector	36
Figure 3.1.1	Primers used for Screening PLA <sub>2</sub> genes	42
Figure 3.1.2	Alignment of various $\alpha$ -neurotoxin amino acid sequences	43
Figure 3.2	Example of a hypothetical screen	45
Figure 3.3	Screening Results for King Brown library (PLA <sub>2</sub> genes) showing positive wells	48
Figure 3.4	Optimisation for King Brown snake cDNA library based on gene specific PLA <sub>2</sub> primers	56
Figure 3.5	ClustalW alignment of the novel PLA <sub>2</sub> genes from King Brown snake (All Groups)	59
Figure 3.6	Phylogram of the Novel Nucleotide Sequences	63
Figure 3.7	Novel King Brown PLA <sub>2</sub> genes (Group B)	65
Figure 3.8	Phylograms of Group AC and B sequences	67
Figure 3.9	The clustalW alignment of the putative proteins sequences	70
Figure 3.10	Clustal Alignment of Known Proteins from King Brown	74
Figure 3.11	Clustal W alignment of Novel Long Sequences	77
Figure 4.1	Chromatogram of Crude Death Adder Venom	83
Figure 4.2	A chromatogram of Region A, showing further separation	84
Figure 4.3	Results from Screening Death Adder venom gland cDNA library with Gene specific primers	88
Figure 4.4	The Results for Optimizing the number of Cycles used for screening	90
Figure 4.5	Novel PLA <sub>2</sub> gene for Death Adder	92
Figure 4.6	Clustal alignment of unknown with Acanthin I & II	93



Figure 4.7	Phylogram of Alignment with other proteins	95
Table 1.1	Danse's Classification of Group I and II PLA <sub>2</sub>	9
Table 1.2	Location And Function of the Conserved Disulfide Bridges in PLA <sub>2</sub>	11
Table 1.3	Overseas and native snakes which have been identified with positive PLA <sub>2</sub> genes	15
Table 1.4	α-neurotoxins from Australian snakes	20
Table 4.1	Size and PLA <sub>2</sub> activity of protein components from Crude Death Adder venom	86
Table 5.1	Various PLA <sub>2</sub> enzymes from <i>A. antarcticus</i> from different regions of Australia	103
Diagram 3.1	a graphical representation of reach the end point for sampling	54

## Abstract

A complex mixture of post-synaptic and pre-synaptic neurotoxins have been identified in both King Brown snake and Death Adder venom. However, since the experiments were conducted using pooled venom samples from different snakes, it could be argued that the large number of homologous toxins previously reported were due to individual (intra-species) variation. Furthermore, previous studies on King Brown snake and Death Adder toxins were mainly at a protein level and there have been few studies at a genotypic level. Thus, a series of experiments were conducted to ascertain whether individual King Brown snakes and Death Adders express an array of toxins, or whether the results were due to the use of pooled venom samples.

In order to isolate all venom genes and their homologues from the genome of both snakes, individual snakes from both species were sacrificed and cDNA libraries were prepared from their excised venom glands. The venom gland cDNA libraries were then screened exhaustively for the presence of any post-synaptic and pre-synaptic neurotoxin genes i.e. the Phospholipase A<sub>2</sub> gene and its homologues, as well as the alpha-neurotoxin genes (both long and short chain  $\alpha$ -neurotoxins genes). While the results from King Brown snake cDNA library supported the hypothesis that individual snakes express a number of homologues of PLA<sub>2</sub> enzymes, some of which might be toxins, the actual number of venom genes and the variation between each homologue were lower than previously reported values. Multiple copies of PLA<sub>2</sub> genes were isolated supporting the contention that there was gene duplication of these venom genes. Regions of hyper-variability were also observed within the PLA<sub>2</sub> genes, further supporting the idea that these neurotoxin genes evolved at an accelerated rate. However, with respect to Death Adder cDNA library, the results indicated that there was only a single copy of the PLA<sub>2</sub> gene, and supported the hypothesis that variation may be due to pooled venom samples.

Unfortunately, no results were obtained for  $\alpha$ -neurotoxin genes despite numerous attempts at isolating the long and short chain  $\alpha$ -neurotoxin genes. Thus no conclusions could be drawn regarding the evolution and complexity of post-synaptic neurotoxins.