Development of Procedures for Casework Specimen Collection and Processing for Organic Gunshot Residue Analysis

A Thesis submitted in fulfilment of the requirements for the award of the degree

Doctor of Philosophy

from

University of Technology Sydney

by

Regina Verena Taudte

M.Sc., B.Sc.

Centre for Forensic Science University of Technology Sydney

Certificate of Authorship and Originality

CERTIFICATE OF AUTHORSHIP AND ORIGINALITY

I, Regina Verena Taudte, certify that the work in this thesis has not previously been

submitted for a degree nor has it been submitted as part of the requirements for a

degree except as fully acknowledged within the text.

I also certify that the thesis has been written by me. Any help that I have received in

my research work and the preparation of the thesis itself has been acknowledged. In

addition, I certify that all the information sources and literature used are indicated in

the thesis.

Regina Verena Taudte

01.03.2016

~ i ~

LIST OF PUBLICATIONS

The chapters presented in this thesis have been published, accepted for publication or prepared for submission to journals as follows:

Chapter 1 - This chapter includes some parts of the literature review:

R.V. Taudte, A. Beavis, L. Blanes, N. Cole, P. Doble, C. Roux, Detection of Gunshot Residues Using Mass Spectrometry, *BioMed Research International*, 2014, Article ID 965403, http://dx.doi.org/10.1155/2014/965403

Chapter 2 – R.V. Taudte, A. Beavis, L. Wilson-Wilde, C. Roux, P. Doble, L. Blanes, A portable explosive detector based on fluorescence quenching of pyrene deposited on coloured wax-printed μPADs, *Lab Chip*, 2013, 13, 4164-4172, DOI: 10.1039/C3LC50609F

Chapter 3 – This chapter has been submitted to the *Journal of Mass Spectrometry*:

R. V. Taudte, C. Roux, D. Bishop, C. Fouracre, A. Beavis, High-throughput screening for smokeless powders and gunshot residues using RapidFire® with tandem mass spectrometry

Chapter 4 – R.V. Taudte, C. Roux, D. Bishop, L. Blanes, P. Doble, A. Beavis, Development of a UHPLC method for the detection of organic gunshot residues using artificial neural networks, *Analytical Methods*, 2015, 7, 7447-7454, DOI: 10.1039/C5AY00306G

Chapter 5 – R.V. Taudte, C. Roux, L. Blanes, M. Horder, K.P. Kirkbride, A. Beavis, The Development and Comparison of Collection Techniques for Inorganic and Organic Gunshot Residues, *Analytical and Bioanalytical Chemistry*, 2016, **408**, 2567-2576, DOI: 10.1007/s00216-016-9357-7

Chapter 6 - This chapter has been prepared for submission to *Forensic Science International:*

R.V. Taudte, C. Roux, A. Beavis, Stability of smokeless powder compounds on collection devices

LIST OF CONFERENCES

The research conducted during this project was presented at several international conferences listed below.

Year	Conference	Presentation
2015	7 th European Academy of Forensic Science Conference, Prague (Czech Republic)	The Development and Comparison of Procedures for the Combined Collection of Organic and Inorganic Gunshot Residues
2014	22 st International Symposium on the Forensic Sciences, Adelaide (Australia)	The Development and Comparison of Procedures for the Combined Collection of Organic and Inorganic Gunshot Residues
2012	21 st International Symposium on the Forensic Sciences, Hobart (Australia)	Development of Procedures for Casework Sample Collection and Processing for Organic Gunshot Residue Analysis

ACKNOWLEDGEMENT

Once upon a time, a girl moved to Australia and started a PhD at the University of Technology Sydney. A few years later...she was finally about to finish. The end of an exciting, mind broadening, fulfilling and sometimes exhausting journey is coming close and I would have never made it to this stage without the help and support of many wonderful people.

First and foremost, I would like to express my deepest appreciation to my supervisors, Associate Professor Alison Beavis and Professor Claude Roux. Thank you for giving me the opportunity to undertake this project, supporting me throughout this incredible journey, encouraging me when encouragement was needed, giving me the freedom to try and follow my ideas and the opportunity to grow as a researcher. Thank you so so much Alison, for spending weekends with me on the shooting range, being incredibly supportive and positive throughout this project and making me always feel accomplished after all our meetings. Thank you Claude! I am indebted to you for your support and trust that made it possible for me to come to UTS and the many opportunities you gave me along the way. Both of you have given invaluable contributions to the preparation of the research output from my doctoral work and I cannot put in words how grateful I am for everything you have done for me.

I would like to thank Dr Lucas Blanes, Dr David Bishop and Professor Philip Doble for their constant help and support especially in regards to micropads and analytical chemistry. Thank you Lucas for the many Brazilian barbecues I enjoyed very much and inspiring me to finally learn the guitar.

I am truly grateful to Sergeant Mark Horder. Without you, this research would not have been possible!

In addition, I would like to thank Elizabeth Chan from NSW Health and Joanna Pryke who initialised this project and provided me with valuable feedback specifically in regards to case work applicability.

I would also like to thank Katie McBean from the Microstructural Analysis Unit (UTS), Dr Richard Wuhrer at the University of Western Sydney and Ken Mason for the support with the SEM and the automated gunshot residue software. My apologies for the many desperate emails I sent you when my computer skills repeatedly abandoned me and I experienced issues with the software.

I would like to show my gratitude to everyone involved in the preparation and review process of manuscripts resulting from this research and in the preparation of this thesis. Your constructive feedback greatly improved the written work related to this PhD and I am extremely grateful for your time and efforts.

I would like to thank Microsoft for developing such an amazing program as office. I am in awe of everyone who had to write a PhD thesis without the possibility to automatically update List of Figure and Tables, as well as references using endnote.

I would like to thank my fellow PhD candidates and colleagues at the university: Ali, Fiona, Nadine, Matt, Dan, Anna, Scott, Marie, Joyce, and many more. Our communal lunches, coffee breaks, Friday bar evenings and other social activities made the last four years more than enjoyable and I am so grateful to have found wonderful friends in many of you.

A big thank you to Ronald and the whole Shimoninski family! Every morning I go to work with a big smile on my face – and this is because of you!

Thank you Claire and everyone involved in the UTS Volleyball Club! You managed to distract my mind from PhD work for at least a few hours per week and reminded me that there is a life outside university.

I would like to thank Maiken, who showed me that every obstacle in life is conquerable and who found surprisingly fun in counting particles.

I want to thank my partner Gabriele for his support throughout the whole time. I know the last few years have not been easy for you and I greatly appreciate all the sacrifices you made to make my life easier even though they made your life harder. You have been there for me, always provided constructive feedback and advices. You have been my motivation and inspiration for undertaking a PhD and I greatly appreciate everything you have done for me! I love you so so much, ti amo, LoYuMuMo! You mean the world to me! Always remember how beautiful you are.

Words cannot express how grateful I am and how much I love my family and friends back home in Germany. Despite the 14,000 kilometres including an ocean separating us, you have been there for me every step of the way. You have given me the strength and endurance to not give up when it became difficult and always focus on the positives. You have always believed in me, even when I doubted myself. You have been my rock, my hope and my strength! I could not think of any better support system and cannot thank you enough for your love and friendship.

To all PhD candidates out there:

As my mum would say:"Bleib übrig!".

TABLE OF CONTENTS

CERTIFICATE OF AUTHORSHIP AND ORIGINALITY	•••••
LIST OF PUBLICATIONS	I
LIST OF CONFERENCES	IV
ACKNOWLEDGEMENT	
TABLE OF CONTENTS	VII
LIST OF FIGURES	XIV
LIST OF TABLES	XXV
ABBREVIATIONS	XXXIV
ABSTRACT	XI
CHAPTER 1:INTRODUCTION	
1.1 PROJECT RATIONALE	2
1.1 GUNSHOT RESIDUE BACKGROUND	2
1.2 SCREENING TESTS FOR GSR	7
1.3 GSR COLLECTION	8
1.4 GSR Analysis	10
1.4.1 IGSR Analysis	
1.4.2 OGSR Analysis	
1.5 GSR INTERPRETATION	12
1.5.1 Discharge of a Firearm	14
1.5.2 Time since Discharge	14
1.5.3 Linkage of Firearms and/or Ammunitions	
1.5.4 Occupational and Environmental Sources	
1.6 CONTAMINATION	20
1.7 Persistence	21

1.8	CONCLUSION	22
1.9	PREVIOUS RESEARCH FOCUSING ON COMBINED IGSR AND OGSR ANALY	SIS 23
1.10	RESEARCH AIMS	26
	ER 2:DEVELOPMENT OF A PORTABLE SCREENING METHOD I	
2.1	BACKGROUND	30
2.2	MATERIALS AND METHODS	33
2.2	l Chemicals and Reagents	33
2.2	2 μPAD Fabrication	34
,	2.2.1 Fabrication Process Optimisation	34
2	2.2.2 Wax Barrier Optimisation	34
,	2.2.3 Influence of Solvents on Wax Barriers	35
,	2.2.4 Optimised μPAD Design	35
2.2	3 Pyrene Application	35
2	2.3.1 Increasing the Temperature	36
2	2.3.2 Surfactant Additive	36
2	2.3.3 Solvent Ratios	36
2	2.3.4 Concentration of Pyrene	36
2.2	4 Detection	37
2.2	5 Fluorescence Quenching	37
2	2.5.1 Preliminary Test	37
,	2.5.2 Sensitivity Test	38
2	2.5.3 Selectivity Test	38
2.2	6 Portable Explosive Detector Prototype	38
2.3	RESULTS AND DISCUSSION	39
2.3	l μPAD Fabrication	39
2.3	2 Application: Explosive Detection by Fluorescence Quenching	45

2.3.3	Portable Explosive Detector Prototype	51
2.3.4	Optimisation	53
2.4	CONCLUSION	54
AND GUI	R 3:HIGH-THROUGHPUT SCREENING FOR SMOKELESS I NSHOT RESIDUES USING RAPIDFIRE® WITH TANDEM MA OMETRY	ASS
3.1	INTRODUCTION	57
3.2	MATERIALS AND METHODS	58
3.2.1	Reagents and Standards	58
3.2.2	RapidFire® – Automated On-line Solid Phase Extraction	59
3.2	.2.1 Instrument	59
3.2	.2.2 Optimisation	60
3.2	.2.3 Calibration Curves	61
3.2.3	Triple Quadrupole Mass Spectrometer	62
3.2.4	Simulated Case Specimens	62
3.3	RESULTS AND DISCUSSION	62
3.3.1	Optimisation	62
3.3.2	Simulated Case Specimens	66
3.4	CONCLUSION	67
	R 4:DEVELOPMENT OF A UHPLC METHOD FOR THE DET R USING ARTIFICIAL NEURAL NETWORKS	
4.1	BACKGROUND	70
4.2	MATERIALS AND METHODS	73
4.2.1	Reagents and Standards	73
4.2.2	Instrumentation	77
4.2	.2.1 Ultra-high Performance Liquid Chromatography	77
4.2	.2.2 Triple Quadrupole Mass Spectrometry	77
4.2.3	Experimental Design	81

4.2.4	Artificial Neural Network	82
4.2.5	Additional Separation Optimisation	83
4.2.6	Method Validation	83
4.2.7	Ammunitions, Firearms and Specimen Preparation	83
4.2.7	.1 OGSR Collection from Hands and Specimen Preparation	. 83
4.2.7	.2 Unburned Smokeless Powder Collection and Sample Preparation	. 85
4.3 RI	ESULTS AND DISCUSSION	86
4.3.1	Artificial Neural Network Training	86
4.3.2	Additional Optimisation	94
4.3.3	Method Validation	97
4.4 Co	ONCLUSION	.04
	5:DEVELOPMENT AND COMPARISON OF COLLECTION UES FOR THE COMBINED COLLECTION OF OGSR AND IGSR	107
5.1 BA	ACKGROUND1	07
5.2 M	ATERIALS AND METHODS	.09
5.2.1	Reagents and Standards	109
5.2.2	Instrumentation	110
5.2.2	.1 Ultra-high Performance Liquid Chromatography	110
5.2.2	.2 Triple Quadrupole Mass Spectrometry	111
5.2.2	.3 Gas Chromatography Mass Spectrometry	111
5.2.2	.4 Scanning Electron Microscopy with Energy Dispersive X-ray Spectroscopy 1	111
5.2.3	Protocol 1 (Swabbing followed by Liquid Extraction)	!12
5.2.3	.1 Extraction Solvent Comparison	114
5.2.3	.2 Extraction Technique Comparison	115
5.2.3	.3 Effect of Multiple Extractions	116
5.2.3	.4 Sonication Times	116
5 2 3	5 Ontimised Condition	116

5.2.3.6	Interference Test	116
5.2.4 Pro	otocol 2 (GSR Stubs followed by Liquid Extraction)	117
5.2.4.1	Extraction Solvent Comparison	118
5.2.4.2	Effect of Temperatures and Multiple Extractions	118
5.2.4.3	Sonication Times	119
5.2.4.4	Optimised Conditions	119
5.2.4.5	Interference Test	119
5.2.5 Pro	otocol 3 (GSR Stubs followed by Solid Phase Microextraction)	120
5.2.5.1	Heating Temperature	121
5.2.5.2	Effect of GSR Stub Adhesive	121
5.2.5.3	Effect of Liquid Immersion	122
5.2.6 Sin	nulated Case Specimens	122
5.3 Resu	LTS AND DISCUSSION	123
5.3.1 Pro	otocol 1	123
5.3.1.1	Extraction Solvent Comparison	123
5.3.1.2	Extraction Technique Comparison	127
5.3.1.3	Effect of Multiple Extractions	132
5.3.1.4	Sonication Time Optimisation	136
5.3.1.5	Optimised condition	140
5.3.1.6	Interference test	141
5.3.2 Pro	otocol 2	143
5.3.2.1	Extraction Solvent comparison	143
5.3.2.2	Effect of Temperatures and Multiple Extractions	147
5.3.2.3	Sonication Times	151
5.3.2.4	Optimised Condition	154
5.3.2.5	Interference Test	155
5.3.3 Pro	otocol 3.	157

5.3.3.1 Heating Temperature 158	
5.3.4 Comparison of Protocol 1 and Protocol 2	
5.3.5 Simulated Case Specimens	
5.3.5.1 Efficiency for IGSR Analysis	
5.3.5.2 Efficiency for OGSR Analysis	
5.4 CONCLUSION 168	
CHAPTER 6:STABILITY OF SMOKELESS POWDER COMPOUNDS ON COLLECTION DEVICES	71
6.1 BACKGROUND 171	
6.2 MATERIALS AND METHODS	
6.2.1 Reagents and Standards	
6.2.2 Instruments and Conditions	
6.2.3 Experimental Design	
6.2.4 Data Analysis and Definitions	
6.3 RESULTS AND DISCUSSION	
6.4 CONCLUSION 184	
CHAPTER 7:CONCLUSIONS AND FUTURE RESEARCH 18	86
REFERENCES	93
APPENDICES22	10

LIST OF FIGURES

rigure	aluminium stubs
Figure	1-2: Prevalence of inorganic composition found in cartridge cases collected during 2008-2010. Overall, 201 cartridge cases from 69 different ammunitions were submitted for analysis corresponding to 49 criminal cases [98]
Figure	1-3: Plume formation influenced by the weapon construction. a: pistol-Walther P38 Series (gases and particles at the muzzle begin forming a cone-shape); b: pistol-Glock 17 pistol (vertical jet from the ejection port); c: revolver-Casull 454 (strong emission from the drum/barrel gap); d: shotgun-Pumpgun Winchester Defender 1300 (cloud from the ejection port) [100].
Figure	2-1: Fabrication process of the microfluidic paper-based analytical device (μPAD) and the application of pyrene (fluorophore) on it. The process consists of: 1. designing a μPAD pattern on the computer; 2. printing the pattern on filter paper; 3. heating the printed wax on the paper using a heat press in order to create fully functioning hydrophobic barriers; 4. pipetting pyrene (in 80:20 MeOH:water) to finally create the finished μPAD (5.)
Figure	2-2: Electronic diagram of the microfluidic paper-based analytical device reader (Bat = battery; D1-D4 = photo (D1), red (D2), green (D3) and ultraviolet light-emitting (D4) diodes; OA2 LM358 and OA3 LM358 = dual operational amplifiers (8 pin integrated circuit); Q1 BC547B and Q2 BC547B = bipolar transistors; R1-R7 = resistors with 10 (R1 and R2), 1 (R3 and R5), 330 (R4 and R6) and 12 (R7) kiloohms ($k\Omega$)).

Figure 2-3: Schematic illustration of the spreading process of the wax, where $W_H = W_{H} + 2W_{H} +$
$W_P + 2L$ with W_H representing the width after heating, W_P the printed
width and L the distance between the spreaded wax and the edge of the
printed line, adapted from [159]
Figure 2-4: Same microfluidic paper-based analytical device design heated at four
different temperatures for 5 min. Heating temperature starting from the
left side: 250, 200, 150, and 140 °C
Figure 2-5: Identical microfluidic paper-based analytical device design heated at the
same temperature (150 °C) for different times (top line from left: 30s, 1
min, 2 min, 4 min; bottom line from left: 4 min, 5min, 6 min, 7 min) 41
Figure 2-6: Upper line: Printed lines with 0.300 mm W _N using a FujiXerox
ColorQube 8870 colour printer, whereby $W_{\rm N}$ represents the nominal
width. Lower line: Printed lines after heating for 5 min at 150°C using a
swing-away heat press (GEO Knight & Co, Inc). Colours tested were
yellow (a, b), cyan (c, d), black (e, f), magenta (g, h) and green (i, j). The
lines were measured and the pictures taken with an EZ4D microscope
(Leica)
Figure 2-7: Nominal widths against widths after heating for the CMYK colours
present in the four different solid inks used in the ColorQube 8870 colour
printer
Figure 2-8: Coloured hydrophobic circles in black (a), magenta (b), cyan (c), yellow
(d) and green (e) with minimal widths of the inner circle from 0.050 to
0.450 mm with increments of 0.050 mm (see left column). All circles are
filled with 5 μL of a 1 mg/mL Terasil Blue aqueous solution. The marked
areas highlight inner circles which are not reliable hydrophobic barriers as
the solution does not stay within them. Image taken under visible light
[159]44
Figure 2-9: Schematic of explosive detection based on fluorescence quenching. a)

Pyrene is deposited in the circled area on the microfluidic paper-based

analytical device (μ PAD) and emits light upon excitation by a light source with 365 nm. b) When explosives are present on top of pyrene on
the circled area on the µPAD, no light is emitted upon excitation 46
Figure 2-10: Interaction mechanism between pyrene and trinitrotoluene (TNT; representing the quencher). Left side: Pyrene alone exhibits fluorescence upon excitation. Right side: No fluorescence is detection due to the formation of a charge transfer complex between the electron-rich pyrene and electron-withdrawing explosive (here TNT) [167]
Figure 2-11: Column A-MeOH:water mixture, B-EtOH:water mixture, C-propanol:water mixture and D-ACN:water mixture. Ratios of organic solvents:water are given in the left column starting from 10 % organic solvent at the top to 100 % at the bottom. Picture is taken with visible light. All solutions are coloured with 1 mg/mL Terasil Blue
Figure 2-12: Comparison of various techniques to increase the pyrene solubility in aqueous solution: blank (a), saturated pyrene solution after heating (30 min, 80 °C), saturated pyrene solution with sodium dodecyl sulfate (c), 0.5 mg/mL pyrene solution (80:20 methanol:water) (d)
Figure 2-13: Fluorescence of pyrene in various concentrations: blank (a), 0.05 mg/mL (b), 0.1 mg/mL (c), 0.25 mg/mL (d), 0.5 mg/mL (e), 0.75 mg/mL (f), and 0.1 mg/mL (d)
Figure 2-14: Microfluidic paper-based analytical device (μPAD) with 1 μL of 0.5 mg/mL pyrene solution in methanol : water (80:20) on circles with 5.000 mm diameter generated under the same conditions as in Figure 2-3. 2-Same μPAD after the deposition of 1 μL of 10 different explosives (A: TNB, B: 1,3-DNB, C: NB, D: TNT, E: 2,4-DNT, F: 4-NT, G: 4-A-2,6-DNT, H: RDX, I:tetryl, J:PETN) demonstrating fluorescence quenching. 3- μPAD generated under the same conditions with nine different non-explosive substances and one explosive (A: negative control, B: water, C: milk, D: coffee, E: tea, F: coke, G: beer, H: wine, I: Mylanta Antacid
Dual Action and J: TNT (positive control))

Figure 2-15: Illustration of the portable explosive detector prototype. The first step
(not shown) includes inserting the calibration point between the
ultraviolet light-emitting diode (LED) and the photodiode and turning the
calibration knob until the green LED flashes. The second step (displayed
in the Figure) shows the detection of explosives on the microfluidic
paper-based analytical device
Figure 3-1: Scheme of the RapidFire® connected to an Agilent triple quadrupole
mass spectrometer (QQQ-MS)
mass spectrometer (QQQ 1415)
Figure 3-2: Sum of the % recoveries of the target compounds using nine different
cartridge types. Different cartridges were loaded with a 10 ppm mixed
standard (10 µL) of the target compounds and were eluted using
isopropanol (0.75 mL/min). Error bars represent standard deviations
(n = 3)
Figure 3-3: Extraction efficiencies of different solvents/solvent compositions
presented as sum of the % recoveries of the target compounds. A C18
Type C cartridge was loaded with a 10 ppm mixed standard and eluted
using the solvent/solvent system at 0.75 mL/min. Error bars represent
standard deviations $(n = 3)$. IPA = isopropanol, MeOH = methanol,
ACN = acetonitrile, DCM = dichloromethane
Figure 3-4: Total ion chromatogram demonstrating the very short analysis time per
sample
sample03
Figure 4-1: Representation of gradients defining the experimental space for input
data to the Artificial Neural Network. Five gradients were used as training
points to train the network, two gradients were used as verification data to
mitigate overlearning.
oz
Figure 4-2: Schematic diagram of the 1:1-19-33:33 multilayer perceptron network
providing the smallest error for the prediction of the retention times of the
33 compounds of interest. The gradient slope represents the input data,
the retention times are given through the output data
the retention times are given unough the output data

Figure 4-3: Response Resolution Plot. The minimum peak pair is plotted versus the
gradient (% MeOH/min), whereby MeOH stands for methanol. The run
times of the maxima of the minimum peak pairs (representing the best
resolution) are shown in the brackets. The gradient with 4.6 %/min
MeOH increase was used as it provided efficient resolution and short
analysis time91
Figure 4-4: Graph showing the correlation between observed retention time [min]
and predicted retention time [min] for the gradient 0.7 % MeOH/min (a)
and the gradient 4.6 % MeOH/min (b). MeOH = methanol
Figure 4-5: Early sections of the chromatograms when using different initial
methanol (MeOH) concentrations (5-30 %). The blue circled area
highlights the relationship between RDX (a) and 1,3-DNG (b) with
different initial MeOH %, while the red area shows the relationship
between EGDN (c) and HMX (d)95
Figure 4-6: Optimised separation of 32 organic gunshot residue compounds under
214 nm. 1 = NGU, 2 = resorcinol, 3 = DDNP, 4 = RDX, 5 = 1,3-DNG,
6 = 1,2-DNG, 7 = EGDN, 8 = HMX, 9 = TNB, 10 = 1,3-DNB, 11 = NB,
12 = NG, $13 = tetryl$, $14 = TNT$, $15 = 4-A-2,6-DNT$, $16 = 3,4-DNT$,
17 = DMP, 18 = 2,4-DNT, 19 = 2,6-DNT, 20 = 2,3-DNT, 21 = 2-naphthol
(internal standard), $22 = m-NT$, $23 = DEP$, $24 = N,N'-DPU$, $25 = PETN$,
26 = 4 - nDPA, $27 = MC$, $28 = N - nDPA$, $29 = DPA$, $30 = 2,4 - DNDPA$,
31 = 2-NDPA, $32 = EC$, $33 = DBP$. 20 ng of each compound were
injected97
Figure 4-7: Overlayed chromatograms of smokeless powder before shooting (40
S&W, Winchester, Australia; red dashed line) and the gunshot residues
collected from the hands of a shooter after discharge using a 22 Glock
pistol (blue line). $1 = 1,2-DNG$, $2 = 1,3-DNG$, $3 = NG$, $4 = 2$ -naphthol
(internal standard), 5 = DEP, 6 = MC, 7 = DPA, 8 = 2,4-DNDPA, 9 = EC,
10 = DBP

Figure 5-1: Swabbing kit used for the collection of gunshot residues by medi wipes
The kit includes a pair of gloves, plastic tweezers, a scintillation vial
Kendall TM alcohol swab, and pen
Figure 5-2: Scheme of specimen preparation using alcohol wipes as collection
devices. After collection, the swab is liquid extracted in 5 mL solvent and
the extract filtered using two syringe filters (10 μm and 0.8μm). The
inorganic particulates are hereby collected on the second syringe filte
which is directly mounted on a gunshot residue stub for subsequen
analysis by scanning electron microscopy with energy dispersive x-ray
spectroscopy (SEM-EDX). The extract is dried under a stream of nitrogen
and reconstituted in 196 μL solvent and 4 μL volumetric internal standard
are added for ultra-high performance liquid chromatography (UHPLC
analysis114
Figure 5-3: Scheme of specimen collection and preparation using gunshot residue
(GSR) stubs as collection device and liquid extraction. After collection
using the GSR stubs, the stubs are analysed for inorganic GSR using
scanning electron microscopy with energy dispersive x-ray spectroscopy
(SEM-EDX). This is followed by liquid extraction in 5.5 mL solvent, the
extract is dried under a stream of nitrogen and reconstituted in 196 μI
solvent and 4 µL volumetric internal standard for organic GSR analysis
using ultra-high performance liquid chromatography (UHPLC) 117
Figure 5-4: Scheme of specimen preparation using gunshot residue (GSR) stubs as
collection devices and solid phase microextraction (SPME). Afte
collection, the organic compounds are heated and absorbed by the SPMI
fibre. The stub is analysed by scanning electron microscopy with energy
dispersive x-ray spectroscopy (SEM-EDX) for inorganic GSR. The
organic compounds are desorbed from the fibre by direct immersion o
the fibre in 196 µL solvent and 4 µL volumetric internal standard (5 min
and analysed using a previously developed ultra-high performance liquid
chromatography method

Figure 5-5: Percentage recoveries of the target organic gunshot residues extracted
from spiked swabs (25 ng) by liquid extraction (5 mL solvent, 15 min
sonication followed by 5 min centriguation) using eight different
solvents/solvent systems. Error bars represent standard deviations $(n = 3)$.
ACN = acetonitrile, MeOH = methanol, IPA = isopropanol,
DCM = dichloromethane, MTBE = methyl tertbutyl ether. After liquid
extraction, the extracts were dried under a steady stream of nitrogen and
reconstituted in 196 μL ACN:MeOH (1:1) and 4 μL volumetric internal
standard

Figure 5-6: Percentage recoveries of the target compounds when liquid extracted (5 mL methyl tertbutyl ether) from spiked alcohol wipes (presented here the overall % recoveries from 12, 20, and 30 ng) using four different techniques, i.e. sonication (15 min at ambient temperatures), centrifugation (5 min). comb technique (sonication (15)min)+centrifugation (5 min)) at room temperature, comb technique + T (15 min heated (45 °C) sonication followed by centrifugation). Error bars represent standard deviations (n = 3). After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 µL aceonitrile:methanol (1:1) and 4 µL volumetric internal standard. 128

Figure 5-8: Comparison of the % recoveries of the target organic gunshot residue compounds spiked on alcohol wipes (25 ng) when performing single (15 min sonication at ambient temperatures) or double liquid extraction (2 x 15 min sonication at ambient temperatures and combining the extracts) of the alcohol wipes using 5 mL methyl tert-butyl ether (MTBE). After

nitrogen and reconstituted in 196 μL acetonitrile:methanol (1:1) and 4 μL volumetric internal standard. Error bars represent standard deviations (n = 3)
Figure 5-9: Comparison of the recoveries of the target organic gunshot residue compounds spiked on alcohol swabs (25 ng) when performing single (15 min sonication at ambient temperatures) or double extraction (2 x 15 min sonication and combining the extracts) of the alcohol wipes using 5 mL acetone. After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μ L acetonitrile:methanol (1:1) and 4 μ L volumetric internal standard. Error bar represent standard deviations (n = 3).
Figure 5-10: Percentage recoveries of the target compounds spiked on alcohol swabs (25 ng) when liquid extracted using 5 mL methyl tert-butyl ether and four different sonication times (5, 10, 15, and 20 min) at ambient temperatures. After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL acetonitrile:methanol (1:1) and 4 μL volumetric internal standard. Error bars represent standard deviations (n=3).
Figure 5-11: Percentage recoveries of 15 tested organic gunshot residue compounds from spiked hands (25 ng) collected using alcohol swabs that were liquid extracted using the optimised extraction conditions (5 min sonication at ambient temperatures using 5 mL methyl tert-butyl ether). After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μ L acetonitrile:methanol (1:1) and 4 μ L volumetric internal standard. The recovery of DBP from hands is excluded in the chart. Interferences to DBP were extracted from hands and prohibited the determination of its recovery. Error bars represent standard deviations (n = 3)
Figure 5-12: Percentage recoveries of the targeted organic gunshot residue

compounds from spiked gunshot residue stubs (30 ng) liquid extracted

liquid extraction, the extracts were dried under a steady stream of

Figure 5-15: Percentage recoveries of the target compounds liquid extracted from spiked (15 ng) gunshot residue stubs using three different conditions and 5.5 mL acetone. The conditions were: s, nh = single extraction, non-heated (15 min sonication at ambient temperatures); s, h = single extraction, heated (15 min sonication at 45 °C); d, nh = double extraction, non-heated (2 x 15 min sonication at ambient temperatures and

combining the extracts). After liquid extraction, the extracts were dri	ed
under a steady stream of nitrogen and reconstituted in 196	ιL
acetonitrile:methanol (1:1) and 4 μL volumetric internal standard. Error	ror
bars represent standard deviations (n = 3).	48

Figure 5-19: X-ray spectra and picture of a spherical 8.50 μm wide gunshot residue particle incorporating the elements lead (Pb), antimony (Sb), and barium

(Ba)	analysed	using	scanning	electron	microscopy	coupled	with	energy
dispe	rsive X-ra	ay spec	ctroscopy.					162

Figure 6-1: Percentage recoveries of the different target compounds, namely resorcinol (a), RDX (b), HMX (c), TNB (d), m-DNB (e), NG (f), tetryl (g), TNT (h), 4-A-2,6-DNT (i), 2,4-DNT (j), N-nDPA (k), DPA (l), and EC (m) extracted using the optimised protocols (Chapter 5) from spiked swabs and stubs on several days after initial spiking. The days involved day 0, 1, 2, 4, 8, 15, 22, 29, 40, 49, 63. The spike amount of each compound was 10 ng. Error bars represent standard deviations (n = 3).178

LIST OF TABLES

Table 1-1: List containing some of the most common organic compounds present in
gunshot residues [4, 6, 10, 11, 14, 21-24]
Table 2-1: List of the width before (W _p) and after heating (W _H) of vertical lines
with 0.300 mm $W_{\rm N}$ (nominal width) at 150°C for 5 min using a swing-
away heat press
Table 2-2: Minimum detectable masses of 10 explosives using the prototype
explosive detector and the optimised microfluidic paper-based analytica
device (1µL 0.25 mg/mL pyrene, 5 mm diameter circle)
Table 3-1: Limits of detection (LODs) and limits of quantification (LOQs) of the
target compounds in ng when loaded onto a C18 cartridge and eluted
using isopropanol (0.75 mL/min)
Table 3-2: Recoveries [ng] of the target compounds from simulated gunshot residue
specimens collected at a firing range. Shooting A: Three shots using a
pistol with 44 Rem Magnum (PMC) (Smith&Wesson); Shooting B: Three
shots using shotgun (Remington, USA) with SuperX (12 gauge
Winchester, Australia). ND = not detected67
Table 4-1: List of target compounds, abbreviations, and functions in propellan
powder or primer (indicated in brackets), the standard concentrations, and
brand [4, 6, 11, 21, 22]74
Table 4-2: Triple quadrupole mass spectrometric conditions in multiple reaction
monitoring mode for the target organic gunshot residues
Table 4-3: List of firearms and ammunition combinations used at the indoor
shooting range. LF primer = lead free primer

Table 4-4: Input data for supervised training of the Artificial Neural Network. The data consists of the average retention times (n = 2) [min] of each target compound at five different gradient conditions. MeOH = methanol 87	
Table 4-5: List of predicted minimum peak pair (MPP) and runtimes [min] of gradients ranging from 0.6-6 %/min with increments of 0.1 %/min. MeOH = methanol	
Table 4-6: List of predicted and observed retention times [min] and % relative standard deviation (% RSD) values of the target compounds using the gradients 0.7 % MeOH/min and 4.6 % MeOH/min. rt = retention time, MeOH = methanol	
Table 4-7: Figures of merit for the detection of gunshot residue compounds by ultraviolet detection at 214 nm with $n = 7$; % RSD = % relative standard deviation	
Table 5-1: Gradient reversed phase program for the ultra-high performance liquid chromatographic separation of the targeted organic gunshot residues 110	
Table 5-2: Percentage recoveries of the compounds of interest liquid extracted (5 mL solvent, 15 min sonication followed by 5 min centrifugation) from spiked swabs (25 ng) using eight extraction solvents (ACN, MeOH, IPA:water (70:30), ACN:water (1:1), water, acetone, DCM, MTBE) measured in triplicates. ACN = acetonitrile, MeOH = methanol, IPA = isopropanol, DCM = dichloromethane, MTBE = methyl tertbutyl ether. After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL ACN:MeOH (1:1) and 4 μL volumetric internal standard	
Table 5-3: Percentage relative standard variations (n = 3) for the extraction of the target compounds spiked on alcohol swabs (25 ng) by liquid extraction (5 mL solvent, 15 min sonication followed by 5 min centrifugation) using the eight tested extraction solvents. ACN = acetonitrile, MeOH = methanol, IPA = isopropanol, DCM = dichloromethane, MTBE = methyl	

tertbutyl ether. After liquid extraction, the extracts were dried	d under a
steady stream of nitrogen and reconstituted in 196 µL ACN:Me	eOH (1:1)
and 4 µL volumetric internal standard.	125

- Table 5-5: Percentage relative standard variations of the recoveries of target compounds (n = 3) from spiked swabs (12, 20, and 30 ng) using 5 mL methyl tertbutyl ether and the four different extraction techniques, i.e. sonication (15 min at ambient temperatures), centrifugation (5 min), comb technique (15 min sonication+5 min centrifugation) at ambient temperatures, comb technique + T (heated (45 °C) sonication (15 min) followed by centrifugation (5 min)). After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL acetonitrile:methanol (1:1) and 4 μL volumetric internal standard. 130
- **Table 5-6:** Student's t-test (paired, two-tailed) results between the recoveries of the spiked (12, 20, and 30 ng) organic gunshot residues on alcohol swabs using 5 mL methyl tertbutyl ether and the different extraction techniques, i.e. sonication (15 min), centrifugation (5 min), comb technique (15 min sonication+5 min centrifugation) at ambient temperatures, comb technique + T (heated (45 °C) sonication (15 min) followed by centrifugation (5 min)). After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL acetonitrile:methanol (1:1) and 4 μL volumetric internal standard. Table a shows the results for conc 1 (12 ng), b for conc 2 (20 ng), c for conc 3 (30

< 0.05 indicates a significant difference. P-values indicating no significant
between the different techniques are shown in bold and italics
Table 5-7: Percentage recoveries of the target organic gunshot residue compounds
spiked on alcohol swabs (25 ng) when extracting the swabs using 5 mL of
different solvents, i.e. acetonitrile (ACN), methyl tert-butyl ether
(MTBE), and acetone and sonication (single extraction: 15 min, double
extraction: 2 x 15 min and combining the extracts) at ambient
temperatures. After liquid extraction, the extracts were dried under a
steady stream of nitrogen and reconstituted in 196 µL ACN:methanol
(1:1) and 4 μL volumetric internal standard. Interferences (% recoveries >
100 %) are indicated in bold and italics
Table 5-8: Percentage relative standard variations of the $\%$ recoveries (n = 3) of the
spiked (25 ng) target organic gunshot residues on alcohol swabs liquid
extracted using 5 mL of different solvents (acetonitrile (ACN), methyl
tert-butyl ether (MTBE), and acetone) and single (sonication for 15 min)
and double extraction (2 x sonication for 15 min and combining the
extracts) at ambient temperatures. After liquid extraction, the extracts
were dried under a steady stream of nitrogen and reconstituted in 196 μL
ACN:methanol (1:1) and 4 μL volumetric internal standard
Table 5-9: Percentage recoveries of the spiked (25 ng) target compounds on alcohol
swabs liquid extracted using 5 mL methyl tert-butyl ether as extraction
solvent and four different sonication times (5, 10, 15, and 20 min) at
ambient temperatures. After liquid extraction, the extracts were dried
under a steady stream of nitrogen and reconstituted in 196 μL
acetonitrile:methanol (1:1) and 4 μL volumetric internal standard. The
numbers in italics highlight the presence of interference indicated by %

ng), and d overall. The p-values are listed in the tables, whereby a p-value

Table 5-10: Percentage relative standard variations (n = 3) of the liquid extractions of the on alcohol swabs spiked (25 ng) target organic gunshot residues using 5 mL methyl tertbutyl ether for the different sonication times

(methyl tert-butyl ether as extraction solvent). After liquid extraction, the
extracts were dried under a steady stream of nitrogen and reconstituted in
196 μL acetonitrile:methanol (1:1) and 4 μL volumetric internal standard.

- **Table 5-12:** Percentage recoveries and % relative standard variations (% RSDs) of the targeted compounds from spiked hands (25 ng) collected using alcohol swabs that were liquid extracted using 5 mL methyl tert-butyl ether and 5 min sonication (normalised to the extraction efficiencies of the solvents). After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL acetonitrile:methanol (1:1) and 4 μL volumetric internal standard. Interferences are indicated by % recoveries > 100 % (italics, bold)...... 142
- **Table 5-14:** Percentage relative standard variations (n = 3) of the recoveries of the target compounds spiked (30 ng) on gunshot residues and liquid extracted from stubs using 5.5 mL of six different solvents/solvent systems

Table 5-16: Percentage relative standard deviations (n = 3) of the target compounds liquid extracted from spiked (15 ng) gunshot residue stubs using three different techniques and 5.5 mL of different solvents (acetonitrile (ACN), methanol (MeOH) and acetone). The techniques are: s,nh: single extraction without heating (15 min sonication at ambient temperatures); s,h: single extraction with heating (15 min sonication at 45 °C); d,nh: double extraction without heating (2 x 15 min sonication at ambient temperatures and combining the extracts). After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL ACN:MeOH (1:1) and 4 μL volumetric internal standard........ 150

Table 5-17: Results (p-values) of the Student's t-tests (paired, two-tailed) between the recoveries when liquid extracting the spiked (15 ng) target compounds from gunshot residue stubs using 5.5 mL of different solvents (acetonitrile (ACN), methanol (MeOH) and acetone) and the three different conditions: s,nh = single extraction without heating (15 min sonication at

ambient temperatures); s,h = single extraction with heating (15 min
sonication at 45 °C); d, nh = double extraction without heating (2 x 15
min sonication at ambient temperatures and combing the extracts). After
liquid extraction, the extracts were dried under a steady stream of
nitrogen and reconstituted in 196 μL ACN:MeOH (1:1) and 4 μL
volumetric internal standard. P-values < 0.05 (bold, italics) indicate a
significant difference

- **Table 5-19:** Percentage relative standard deviations (n = 3) of the different organic gunshot residue compounds spiked (13 ng) on gunshot residue stubs and liquid extracted using 5.5 mL acetone and four different sonication times (5, 10, 15, 20 min) at ambient temperatures. After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL acetonitrile:methanol (1:1) and 4 μL volumetric internal standard.
- **Table 5-21:** Percentage recoveries and % relative standard deviations (% RSDs) of the spiked (20 ng) target compounds on hands collected by gunshot

- Table 5-22: Percentage recoveries of the spiked (20 ng) organic gunshot residue (GSR) compounds from GSR stubs heated at different temperatures ranging from 30 to 150 °C with 20 °C increments extracted using solid microextraction (SPME). The **SPME** fibre phase $(65 \mu m)$ polydimethylsiloxane/divinylbenzene) was exposed for 1 hour followed by 5 min direct immersion in the solvent system (196 µL) acetonitrile:methanol (1:1) and 4 µL volumetric internal standard) that by subsequently analysed ultra-high performance liquid
- Table 5-23: Overview and comparison of the two optimised and superior collection protocols. The optimised collection protocol involving alcohol swabs consists of liquid extracting the swab using 5 mL methyl tertbutyl ether and 5 min sonication at ambient temperatures. After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL acetonitrile (ACN):methanol (MeOH) (1:1) and 4 μL volumetric internal standard. The optimised collection protocol involving gunshot residue stubs consists of liquid extracting the stub using 5.5 mL acetone and 5 min sonication at ambient temperatures. After liquid extraction, the extracts were dried under a steady stream of nitrogen and reconstituted in 196 μL ACN:MeOH (1:1) and 4 μL volumetric internal standard............ 161
- **Table 5-24:** List of average number of characteristic (incorporating Pb, Ba and Sb) gunshot residue (GSR) particles (sizes between 0.8 and 10 μ m) using three different ammunition-firearm combinations per mm² when collected using medi swabs or GSR stubs (one shot, n = 3). The stubs and swabs

were not carbon coated before analysis using scanning electron

microscopy coupled with energy dispersive X-ray spectroscopy.......... 163

min sonication at ambient temperatures using 5.5 mL acetone). The

results of the remaining ammunition-firearm combinations are presented

Table 6-1: Degradation [%] and mean standard deviations (SDs) [%] (n = 3) of the target compounds over 63 days extracted using the optimised protocol (Chapter 5) from spiked (10 ng) gunshot residue stubs and swabs. Stable compounds are defined as compounds degrading less than 15 % on GSR stubs and alcohol swabs.

ABBREVIATIONS

1,2-Dintiroglycerine

1,3-Dinitroglycerine

2,3-DNT 2,3-Dinitrotoluene

2,4-DNDPA 2,4-Dinitrodiphenylamine

2,4-DNT 2,4-Dinitrotoluene

2,6-DNT 2,6-Dinitrotoluene

2-NDPA 2-Nitrodiphenylamine

3,4-DNT 3,4-Dinitrotoluene

3-NT 3-Nitrotoluene

4-A-2,6-DNT 4-Amino-2,6-dinitrotoluene

4-nDPA 4-Nitrosodiphenylamine

4-NT 4-Nitrotoluene

AAS Atomic absorption spectroscopy

ACN Acetonitrile

ANN Artificial Neural Network

APCI Atmospheric chemical ionisation

AUS Australia

Ba Barium

CE Collision energy

DBP Dibutyl phthalate

DCM Dichloromethane

DDNP Diazodinitrophenol

DEP Diethyl phthalate

DMP Dimethyl phthalate

DNB 2,3-Dinitrobenzene

DNT Dinitrotoluene

DPA Diphenylamine

EC Ethyl centralite

EGDN Ethylene glycol dinitrate

EtOH Ethanol

FID Flame ionisation detector

GC Gas chromatography

GC-TEA Gas chromatography coupled to thermal energy analyser

GSR Gunshot residue/s

h Hour/s

HMF Heavy-metal free

HMX Octahydro-1,3,5,7-tetranitro-1,3,5,7-tatrazocine

HPLC High performance liquid chromatography

IED Improvised explosive device

IGSR Inorganic gunshot residues

IMS Ion mobility spectroscopy

IPA Isopropanol

ISTD Internal standard

ISDN Isosorbide dinitrate

 $k\Omega$ Kiloohm

LA-ICP-MS Laser ablation inductively coupled plasma mass

spectrometry

LC Liquid chromatography

LC-QTOF-MS Liquid chromatography coupled to quadrupole time of

flight mass spectrometry

LED Light-emitting diode

LF Lead free

LOD Limit of detection

LOQ Limit of quantification

MC Methyl centralite

MECE Micellar electrokinetic capillary electrophoresis

MeOH Methanol

mg Milligram

min Minute/s

mL Millilitre

MLP Multilayer perceptron

mm Millimetre

mm² Square millimetre

MPP Minimum peak pair

MRM Multiple reaction monitoring

ms Milliseconds

MS Mass spectrometry/ mass spectrometer

MS/MS Tandem mass spectrometry

MTBE Methyl tert-butyl ether

NAA Neutron activation analysis

NB Nitrobenzene

NC Nitrocellulose

ng Nanogram

NG Nitroglycerine

NGU Nitroguanidine

nm Nanometre

N-nDPA N-Nitrosodiphenylamine

N,N'-DPU N,N'-Diphenylurea

NO Nitrogen monoxide

NO₂ Nitrogen dioxide

NPD Nitrogen phosphorus detector

NSW New South Wales

NSWPF New South Wales Police Force

OGSR Organic gunshot residues

Pb Lead

PDMS/DVB Polydimethylsiloxane/divinylbenzene

PETN Pentaerythitol tetranitrate

pg Picogram

PMDE Pendant mercury drop electrode

ppm Parts per million

PTFE Polytetrafluoroethylene

QQQ-MS Triple quadrupole mass spectrometer

R² Coefficient of determination

RDX Hexahydro-1,3,5-trinitro-1,3,5-triazine

resorcinol 1,3-Benzenediol

rpm Revolutions per minute

RSD Relative standard deviation

RT Room temperature

Sb Antimony

SDS Sodium dodecyl sulfate

sec Seconds

SEM-EDX Scanning electron microscopy with energy dispersive

x-ray spectroscopy

SOP Standard operating procedure

SPE Solid phase extraction

SPME Solid phase microextraction

Sr Strontium

SD Standard deviation

TEA Thermal energy analyser

tetryl 2,4,6-Trinitrophenylmethylnitramine

TNB 1,3,5-Trinitrobenzene

TNT 2,4,6-Trinitrotoluene

UHPLC Ultra-high performance liquid chromatography

UP Ultrapure

USA United States of America

UV Ultraviolet

TOF-SIMS Time of flight secondary ion mass spectrometry

W_H Width after heating

W_N Nominal width

W_P Width after printing

μg Microgram

 μL Microlitre

ABSTRACT

The detection and interpretation of gunshot residues (GSR) plays a crucial role in the investigation of firearm related events. Specimens are commonly collected using GSR stubs with double sided adhesive carbon tape. After collection, the stubs can directly be analysed using scanning electron microscopy with energy dispersive x-ray spectroscopy (SEM-EDX), which is widely used for the detection of inorganic gunshot residues (IGSR) as it provides simultaneous elemental and morphological information of discrete particles. Since SEM-EDX analysis focuses on the detection of characteristic GSR particles incorporating the elements lead, antimony and barium, the relatively recent introduction of lead free (LF) and heavy-metal free (HMF) ammunition challenges the current standard operating procedure (SOP) for GSR analysis. Other problems arise from the recent findings of GSR-like particles from environmental and occupational sources. The incorporation of organic gunshot residues (OGSR) into the current SOP can provide additional and complementary information that is alleged to overcome these limitations. This project focused on the detection and incorporation of OGSR to current GSR SOPs on different levels.

A screening technique was developed for the in-field detection of compounds potentially present in smokeless powders and GSR. The technique was based on microfluidic paper-based analytical devices (μPAD) and fluorescence quenching of pyrene and showed promising results for detecting energetic compounds in OGSR. A portable μPAD reader was built and showed potential for in-field detection of GSR (and explosives).

A second screening technique was developed based on solid phase extraction (SPE). This technique can allow pre-concentration and clean-up of samples before

OGSR analysis, which might be necessary considering the low amounts of OGSR that are commonly detected on specimens directly collected after discharge. A proof-of-concept study using a completely automated on-line SPE robot, the RapidFire[®], connected to a triple quadrupole mass spectrometer (QQQ-MS) was conducted showing promising results for the pre-concentration and/or screening of OGSR.

To allow the detection of a broad range of OGSR, an ultra-high performance liquid chromatography (UHPLC) method with ultraviolet (UV) detection and mass spectrometric confirmation using a QQQ-MS was developed using a statistical approach (Artificial Neural Networks (ANN)). This approach was applied for the first time to GSR analysis. The network was trained and used for the prediction of retention times of the target compounds in relation to different gradients. The final UHPLC-UV method was fully validated and tested using simulated case specimens collected at an indoor firing range. It proved sufficiently sensitive and selective for the detection of OGSR from hands and the establishment of smokeless powder profiles.

Three different collection protocols for the recovery of OGSR and IGSR from hands were conceptualised to enable both subsequent OGSR analysis by UHPLC-UV and IGSR analysis by SEM-EDX. Comparing the two superior protocols, the extraction efficiencies of OGSR from alcohol swabs and GSR stubs were found to be comparable, while GSR stubs proved to be more efficient in collecting OGSR. Testing the protocols using simulated case specimens taken at the shooting range confirmed that GSR stubs followed by liquid extraction are more suitable than wipes for a combined collection of OGSR and IGSR.

Finally, the stability of OGSR on collection devices, i.e. alcohol swabs and GSR stubs, was investigated for a time period of 63 days. Interestingly, energetic compounds were found to be relatively stable, while stabilisers, often the target compounds for OGSR, degraded mostly following a negative logarithmic curve. This could be problematic for the developed SOP for the collection and analysis of both OGSR and IGSR, since SEM-EDX analysis is preceding OGSR analysis causing the degradation of compounds of interest.

In summary, an SOP for GSR collection and analysis was developed that could potentially overcome problems arising from LF and HMF ammunitions. Further research studies into persistence and background are necessary to test the value of the developed SOP in a forensic framework.