Comparison of the Performance of Submerged Membrane Bioreactor (SMBR) and Submerged Membrane Adsorption Bioreactor (SMABR)

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Abstract

This study focuses on comparing the performance of submerged membrane bioreactor (SMBR) and submerged membrane adsorption bioreactor (SMABR) over a period of 20 days at a hydraulic retention time (HRT) of 3.1 hours. The effects of PAC on critical flux and membrane fouling were also investigated. The SMABR exhibited better results in terms of mixed liquor suspended solids (MLSS) growth, DOC removal (over 96%), COD removal (over 95%), transmembrane pressure (TMP) and oxygen uptake rate. Nearly 100% of bacteria and 100% removals of total coliforms were removed in both systems. The addition of PAC could maintain the critical flux at a lower TMP value (7.5 kPa), while irreversible fouling caused by PAC occurred when the filtration flux exceeded critical flux.

Keywords: Submerged membrane bioreactor; Oxygen uptake rate; Critical flux; Adsorption

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1. Introduction

The use of membrane filtration technology has been advancing in a rapid place in replacing the conventional water and wastewater treatment processes to produce high quality treated water. Among the membrane processes, membrane bioreactor (MBR) technology is becoming an innovative and promising option for wastewater treatment and reuse. MBR comprises of a suspended growth bioreactor and a filtration on porous membrane, which leads to the total retention of biomass (high microbial concentration) and improved biological reactor operation (high sludge ages) in the bioreactor (Lee et al., 2003). In MBR system design, the submerged membrane configuration can assist in significantly reducing power consumption.

Although MBR offers the effective separation of pollutants and persistence to high or shock loadings, membrane fouling is still an unavoidable obstacle. The occurrence of fouling affects the performance of the membrane either by deposition of a layer onto the membrane surface which introduces additional resistance to permeate flow, or by blockage or partial blockage of the pores which changes the effective pore size distribution (Field et al., 1995). The characteristics of activated sludge (AS) in MBR are one of the cardinal factors to membrane fouling. The sludge matrix within MBR is a mixed liquor of two main fractions: (i) biological flocs formed by a large range of living microorganisms, and (ii) supernatant containing soluble and colloidal compounds. Each element has its own physicochemical and biological properties affecting membrane fouling (Lee et al., 2003; Le Clech at al., 2003). Various attempts have been made to reduce the membrane fouling in submerged MBR (SMBR). Yamamoto et al. (1989) examined the influence of operational modes and found that intermittent suction greatly reduced membrane fouling compared to continuous suction. Lee at al. (2001) indicated that alum and natural zeolite addition to a SMBR not only reduced membrane fouling, but also increased the removal of COD. Furthermore, the association of SMBR and powdered activated carbon (PAC) became a promising unit process for advanced water treatment, because the addition of PAC as pretreatment to membrane processes (such as microfiltration (MF) or ultrafiltration (UF)) could achieve more dissolved organic carbon (DOC) and disinfection by-products (DBPs) removal and mitigate membrane fouling by reducing organic loading to membrane adsorbing organic matters (Kim et al., 2001; Clark and Heneghan, 1991). Tsai et al. (2005) summarized the advantages of PAC addition in MBR system, which indicated that PAC has adsorptive affinity for removal of biologically resistant compounds that may be toxic to the microbial community and it provides an excellent surface for the attachment of microorganisms (Ying and Weber, 1979; Pirbazari et al., 1990a). In SMBR, the entire treatment activity (such as adsorption/biodegradation, liquid-solid separation, and sludge accumulation and withdrawal) can be carried out in a single unit.

The bacterial activity during operation of MBR can be evaluated by measuring the oxygen consumption (by respirometric procedure). Nowadays, the oxygen demand measurement gains great interest because it is directly linked to the biological activity. Thus, respirometry is considered as an essential parameter for controlling AS process in MBR (Rodde-Pellegrin et al., 2002). It is well known that respirometry has the following advantages: (i) it can be used for those substrates that cannot be easily determined analytically, (ii) it is much more sensitive than the methods based on biomass growth or substrate removal, (iii) it is detectable even for substrate

concentrations below 1 mg/L, (iv) it determines the kinetic constants of mixed cultures without changing the qualitative and quantitative composition of the culture, and (v) it is simple and easy method (Pitter and Chudoba, 1990).

The objective of this study is to compare the performances of two MBR systems, namely SMBR alone and submerged membrane adsorption bioreactor (SMABR) in treating a synthetic secondary wastewater. Oxygen uptake rate (OUR) was used to measure the biological activities in both the bioreactors, including the suspended growth in SMBR and suspended/attached growth in SMABR. Dissolved organic carbon (DOC) removal, COD removal, mixed liquor suspended solids (MLSS), transmembrane pressure (TMP), total viable counts and total coliform counts were also investigated. After 20 days operation, critical flux (the critical flux is the flux below which there is no presence of TMP increase in resistance) experiments were conducted to examine the filtration flux stability in both the systems.

2. Methods

2.1 Wastewater

The experiments were conducted using a synthetic wastewater to avoid any fluctuation in the feed concentration and provide a continuous source of completely biodegradable organic pollutants. It was used to simulate high strength domestic wastewater (just after primary treatment process). The synthetic wastewater has DOC of 120-130 mg/L and COD of 320-350 mg/L (COD: N: P = 100:5:1). The composition of synthetic wastewater is given in Table 1 (Lee et al., 2003). NaHCO₃ or H₂SO₄ were added to the wastewater to maintain a constant pH around 7.

2.2 SMBR and SMABR set-up

SMBR and SMABR systems were operated at a constant permeate flux of 10 L/m².h under the same hydraulic retention time (HRT) of 3.1 hours. Each MBR consisted of an activated sludge bioreactor having an effective volume of 6 L. Initially, SMBR and SMABR were filled with sludge and acclimatized to synthetic wastewater for 12 days. The source of the seeding sludge was from Castle Hill Wastewater Treatment Plant in Sydney. The MLSS concentration of the activated sludge was 1.25 g/L. Synthetic wastewater was then added gradually during the acclimatization time to support the microbial growth in both the MBR systems. In SMABR system, a predetermined amount of PAC (5 g/L) was added into the reactor at the beginning of the acclimatization period to adsorb the dissolved organic substances. The PAC amount was predetermined according to the previous study of the authors (Guo et al., 2005). The PAC (80% min finer than 75 micron) used was wood based carbon with a surface area of 882 m²/g and a mean pore diameter of 30.61 Å. There was no further addition of PAC during the experimental period.

A polyethylene hollow fiber membrane module was used with the pore size of 0.1 μ m and surface area of 0.195 m². The schematic diagram of the submerged hollow fiber microfiltration system is shown in Fig. 1. Synthetic wastewater was pumped into the reactor using a feeding pump to control the feed rate while the effluent flow rate was controlled by a suction pump. Level sensor was used to control the wastewater volume in the reactor. A pressure gauge was used to measure the transmembrane pressure (TMP)

and a soaker hose air diffuser was used to maintain a high air flow rate (9 L/min or 2.77 $m^3/m^2_{(membrane area)}$.h). The bubbling of air has three functions in the systems: (i) sweeping the membrane surface, (ii) mixing the PAC in SMABR and (iii) supplying oxygen to facilitate the biological degradation organics during the long term operation of MBR. For physical cleaning of membranes, filtrate backwash was used every 1 hour for 1 min duration at a backwash rate of 30 L/m².h.

Fig. 1. Experimental set-up of SMBR

2.3 Analysis

YSI 5300 Biological Oxygen Monitor was used to measure oxygen uptake rate due to its useful tool for measuring samples including respiration, oxidative activity, and cellular metabolism studies. The oxygen consumption measurement can be achieved through use of oxygen electrode with oxygen permeable Teflon membrane. Voltage generated from the reaction is proportional to the oxygen concentration of the sample and produces oxygen uptake or evolution curves in 2 to 15 minutes. During the acclimatization, the wastewater withdrawn from the aeration tank at different periods was monitored. DOC of the influent and effluent was measured using the Analytikjena Multi N/C 2000. For measuring MLSS, three samples were taken each time and the average values were then calculated. Total viable counts and total coliform counts were carried out using spread plate technique on nutrient agar and MacConkey agars as media respectively. All samples were diluted using 0.1% bacteriological peptone water. Nutrient agar, MacConkey agar and bacteriological peptone were obtained from OXOID[®].

3. Results and Discussion

3.1 Comparison of MLSS growth

The average concentrations of MLSS in the two systems were investigated. After 12 days of acclimatization, SMBR and SMABR started with the MLSS concentration of 2.58 g/L and 1.81 g/L respectively. The growth of MLSS in SMBR was steady and gradual, whereas a significantly higher growth was observed in SMABR system due to the increase of adsorption surface area made available through the incorporation of PAC. The average concentrations of MLSS in SMABR remained constant (around 10 g/L) after a 10 day-operation.

3.2 Comparison of organic removal

DOC and COD removal efficiencies were measured during the 20 days of operation, which are shown in Fig. 2. The results indicated that both systems achieved excellent DOC and COD removals of over 95% and 94% respectively. SMABR had slightly higher DOC and COD removal efficiencies as compared to SMBR (reaching up to 99% DOC removal and 100% COD removal occasionally). This is due to PAC had the simultaneous functions of biodegradation by attached microorganisms on its surface as well as adsorption to improve the DOC and COD removal efficiencies. During the operation of SMABR, bioreaction took place due to the growth of the biomass supported by PAC. The adsorbed organics on the PAC were biodegraded with time by the biomass, which hence created sites for further adsorption of organics on the PAC. Since the simultaneous activity of biodegradation and adsorption on PAC attained at its peak after 5 days, the DOC and COD removal efficiencies decreased slightly from day 6 to 12. However, after the 13th day of operation, the DOC and COD removal restored

again as a permeable activated sludge layer was formed on some membrane fibers. The later was made through visual observation.

Fig. 2. DOC and COD profile of SMBR and SMABR systems (filtration flux = 10 L/m^2 .h; PAC dose = 5 g/L; backwash rate = 30 L/m^2 .h; backwash = 1 minute every 1 hour; HRT = 3.1)

In this study, SMABR and SMBR performance had a marginal difference in terms of the organic removals because the synthetic wastewater used in this study was completely biodegradable organic pollutants. The advantage of SMABR is to remove persistent organic compounds. SMABR showed much higher organic removal efficiency than SMBR when persistent organic compounds were present in the wastewater. Another study conducted by the authors showed the capability of SMABR in removing persistent organic matters (Guo et al., 2005).

3.3 Comparison of TMP

In any membrane bioreactor, the TMP generally increases with the operational time. Usually, it can only be reduced by membrane cleaning. In the field works, SMBR together with automation backwash control are normally used to minimize the membrane fouling thus to extend the operation period of the MBR system. PAC plays a significant role in fouling reduction as PAC adsorbs a part of the organic matter. The variation of TMP values were measured during the operation of both SMBR and SMABR systems. The TMP in both cases increased slightly during the 20-day of operation (e.g. 9 kPa and 7.5 kPa of TMP developed in SMBR and SMABR respectively). The SMABR system had lower TMP development compared to SMBR system. This is due to the direct adsorption of dissolved organic matters onto PAC. Thus, PAC can mitigate the membrane fouling.

3.4 Comparison of Oxygen uptake rate (OUR)

OUR was used to study the dissolved oxygen (DO) consumption rates in both SMBR and SMABR systems. This relates to the microbial activity at different periods of experiment. Figs. 3 and 4 present the DO variation of the mixed liquor from the aeration tank of both the systems. In SMBR system, the OUR had a lower value during the first 3 days (55% on the first day and 72% on the third day). Moreover, the OUR reached the equilibrium within 16 and 28 minutes with the mixed liquor taken after 1 and 3 days respectively. After that, the OUR of SMBR system was over 94% and equilibrated within 14 minutes. On the other hand, the OUR in SMABR system had better performance from the initial stage of the experiment (also reached the equilibrium within 14 minutes), which meant that there were more microbial substances in SMABR system. The same conclusions were drawn from MLSS and DOC removal efficiencies.

> Fig. 3. OUR variation of the mixed liquor in SMBR system Fig. 4. OUR variation of the mixed liquor in SMABR system

3.4 Comparison of total viable counts and total coliform counts

A quantitative microbiological analysis was carried out with influent, effluent and mixed liquor of SMBR and SMABR on a regular basis. In order to estimate the number of viable bacteria in these samples, viable counts were carried out with the spread plate technique using nutrient agar as medium. Uniform increase in number of viable bacteria was observed in the mixed liquor of both the systems (Table 2). This may be due to the fact that the composition of synthetic wastewater used in these experiments was very rich in nutritional sources such as glucose and yeast extract. In SMBR, the viable

numbers increased rapidly and reached a stationary phase in 10 days (around 6×10^2 cfu/ml). On the other hand, higher degree of growth was noticed in SMABR and bacterial numbers increased rapidly from 3.5×10^2 to 2.24×10^4 cfu/ml during the first 15-day of operation. Adsorption of bacteria on to PAC particles must have contributed to this high numbers. In reality, the bacterial amount in this sample should even be more. The underestimation was due to the difficulty in detaching them from PAC particles.

In order to test the microbiological quality of treated wastewater, viable counts were carried out in both influent and effluent samples. Total coliform counts were also measured. The synthetic wastewater had the viable count of 2.8×10^3 cfu/ml and total coliform of 270 cfu/ml. After treatment, the viable count was less than 15 cfu/ml in both systems. 100% removals of total coliforms were also observed in treated effluent samples of SMBR and SMABR systems.

Table 2 Total viable counts and total coliform counts at different periods of operation

3.5 Comparison of critical flux

Critical flux experiments were carried out after 20-day operation of SMBR and SMABR systems to examine the membrane fouling. The membrane was physical cleaned by using backwash and the cake layer formed by activated sludge was brush off before starting the critical flux experiment. After each 40 minute-flux-step, 1 minute-backwash was provided at a backwash rate of 30 L/m².h using membrane filtrate. The purpose of backwash was mainly to minimize the TMP increase due to reversible fouling during every experimental flux-step, which could lead to TMP development. Figs. 5 and 6 show the critical flux of SMBR and SMABR systems. According to the

figures, both of the systems had the same critical flux value of 20 L/m².h. However, the TMP value of the SMBR system was much higher than that of the SMABR systems, which were 33 and 7.5 kPa respectively at a filtration flux of 20 L/m².h. This indicated that PAC can reduce membrane fouling. During the cycle test, it was observed that the TMP values obtained during the descending (filtration flux) phase were greater than the corresponding values recorded during the ascending phase. Especially for SMABR system, the TMP values were nearly three folds of the ascending (filtration flux) phase values. For example, at the critical flux-step of 20 L/m².h, TMP were 7.5 and 21 kPa for the ascending and descending phases, respectively. These observations indicated that SMABR system formed an initial irreversible fouling due to small PAC particles when the filtration flux was higher than critical flux. The formation of some reversible fouling led to less TMP developments of the SMBR system in the descending phases when compared to SMABR.

Fig. 5. Constant filtration flux vs. TMP of SMBR systemFig. 6. Constant filtration flux vs. TMP of SMABR system

4. Conclusions

SMBR and SMABR systems were compared based on different membrane performance during 20 days operation. It was demonstrated that SMABR system had better performance than SMBR system. PAC addition could mitigate the membrane fouling and led to less TMP development in SMABR system. After acclimatization, the SMABR showed more stable OUR (over 94%) than that of SMBR system. Adsorption of bacteria on to PAC particles presented higher growth in terms of total viable counts in SMABR system. Nearly 100% of bacteria in terms of viable count were removed in both the systems and 100% removals of total coliforms were also observed in treated effluent. Both systems showed the same critical flux of 20 L/m^2 .h after 20 days running except that the SMBR had higher TMP value to maintain the sustainable flux.

Acknowledgment

This research was funded by DEST International Science Linkages Competitive Grant.

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Compounds	Molecular weight	Concentration
	(g/mol)	(mg/L)
Organics and nutrients		
Glucose ($C_6H_{12}O_6$)	180.0	280
Ammonium sulfate ((NH ₄) ₂ SO ₄)	132.1	72
Potassium phosphate (KH ₂ PO ₄)	136.1	13.2
Trace nutrients		
Calcium chloride (CaCl ₂ ·2H ₂ O)	147.0	0.368
Magnesium sulfate (MgSO ₄ ·7H ₂ O)	246.5	5.07
Manganese chloride (MnCl ₂ ·4H ₂ O)	197.9	0.275
Zinc sulfate ($ZnSO_4 \cdot 7H_2O$)	287.5	0.44
Ferric chloride anhydrous (FeCl ₃)	162.2	1.45
Cupric sulfate (CuSO ₄ ·5H ₂ O)	249.7	0.391
Cobalt chloride (CoCl ₂ ·6H ₂ O)	237.9	0.42
Sodium molybdate dihydrate (Na ₂ MoO ₄ ·2H ₂ O)	242.0	1.26
Yeast extract		30

Table 1 Constituents of the Synthetic Wastewater

Day	System	Total Count, cfu/mL	Total Coliforms, cfu/mL
0	SMBR	1.6×10^{2}	90
	SMABR	3.5×10^2	35
10	SMBR	6.5×10^2	38
	SMABR	1.6×10^4	43
15	SMBR	6×10^{2}	30
	SMABR	2.24×10^4	23

Table 2Total viable counts and total coliform counts at different periods of operation

Samples were taken from the middle part of the reactor



Fig. 1. Experimental set-up of SMBR



Fig. 2. DOC and COD profile of SMBR and SMABR systems (filtration flux = 10 L/m^2 .h; PAC dose = 5 g/L; backwash rate = 30 L/m^2 .h; backwash = 1 minute every 1 hour; HRT = 3.1)



Fig. 3. OUR variation of the mixed liquor in SMBR system



Fig. 4. OUR variation of the mixed liquor in SMABR system



Fig. 5. Constant filtration flux vs. TMP of SMBR system (LMH = L/m^2 .h)



Fig. 6. Constant filtration flux vs. TMP of SMABR system (LMH = L/m^2 .h)